



ISSN: (Print) (Online) Journal homepage: https://www.tandfonline.com/loi/gnpl20

Cytoprotective, antioxidant, and anti-migratory activity of *Pistacia lentiscus L*. supercritical carbon dioxide extract on primary human endothelial cells

Roberta Giordo, Annalisa Cossu, Maria Cristina Porcu, Roberto Cappuccinelli, Grazia Biosa, Javad Sharifi-Rad, Luca Pretti, Gheyath K. Nasrallah, Gianfranco Pintus & Anna Maria Posadino

To cite this article: Roberta Giordo, Annalisa Cossu, Maria Cristina Porcu, Roberto Cappuccinelli, Grazia Biosa, Javad Sharifi-Rad, Luca Pretti, Gheyath K. Nasrallah, Gianfranco Pintus & Anna Maria Posadino (2022): Cytoprotective, antioxidant, and anti-migratory activity of *Pistacia lentiscus L*. supercritical carbon dioxide extract on primary human endothelial cells, Natural Product Research, DOI: <u>10.1080/14786419.2022.2130304</u>

To link to this article: <u>https://doi.org/10.1080/14786419.2022.2130304</u>

+	View supplementary material 🗗	Published online: 06 Oct 2022.
	Submit your article to this journal 🛽 🖉	Article views: 131
ď	View related articles 🗷	Uiew Crossmark data 🗹



Check for updates

Cytoprotective, antioxidant, and anti-migratory activity of *Pistacia lentiscus L*. supercritical carbon dioxide extract on primary human endothelial cells

Roberta Giordo^a (**b**), Annalisa Cossu^b, Maria Cristina Porcu^c, Roberto Cappuccinelli^c, Grazia Biosa^c, Javad Sharifi-Rad^{d,e} (**b**), Luca Pretti^c, Gheyath K. Nasrallah^{f,g} (**b**), Gianfranco Pintus^{b,h} (**b**) and Anna Maria Posadino^b (**b**)

^aCollege of Medicine, Mohammed Bin Rashid University of Medicine and Health Sciences, Dubai, United Arab Emirates; ^bDepartment of Biomedical Sciences, University of Sassari, Sassari, Italy; ^cPorto Conte Ricerche S.r.I, Alghero, Sassari, Italy; ^dPhytochemistry Research Center, Shahid Beheshti University of Medical Sciences, Tehran, Iran; ^eFacultad de Medicina, Universidad del Azuay, Cuenca, Ecuador; ^fDepartment of Biomedical Sciences, College of Health Sciences Member of QU Health, Qatar University, Doha, Qatar; ^gBiomedical Research Center, Qatar University, Doha, Qatar; ^hDepartment of Medical Laboratory Sciences, University of Sharjah, Sharjah, United Arab Emirates

ABSTRACT

Green chemistry is a useful tool for producing valuable chemicals from biomass. However, extracted compounds need to be tested for safety and efficacy before their use in humans. Here we investigate the chemical composition and biological effects of a leaves Pistacia lentiscus L. supercritical carbon dioxide (SCCO₂) extract. Terpenes represented the main extract fraction, with Germacrene D (11.18%), delta-cadinene (10.54%), and alpha-pinene (8.7%) the most abundant molecules. Challenged with endothelial cells (ECs), increasing extract concentrations failed to affect cell proliferation or promote cell toxicity. ROS assessment in unstressed and H₂O₂treated ECs revealed an extract dose-dependent antioxidant activity. Exposition of H₂O₂-treated ECs to increasing extract concentrations dose-dependently counteracted H₂O₂-induced cell impairments. The extract significantly counteracted fetal calf serum-induced ECs migration. For the first time, we report that a SCCO₂ extract obtained from PL leaves is safe on ECs and may be a useful source of valuable compounds with vasculoprotective properties.

ARTICLE HISTORY

Received 8 July 2022 Accepted 24 September 2022

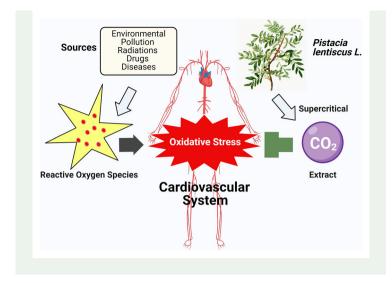
KEYWORDS

Pistacia lentiscus; supercritical CO₂; endothelial cells; reactive oxygen species; antioxidants; cardiovascular diseases;

CONTACT Gianfranco Pintus g gpintus@uniss.it;Luca Pretti pretti@portocontericerche.it; Gheyath K. Nasrallah g gheyath.nasrallah@qu.edu.qa

Supplemental data for this article can be accessed online at https://doi.org/10.1080/14786419.2022.2130304.

 $\ensuremath{\mathbb{C}}$ 2022 Informa UK Limited, trading as Taylor & Francis Group



1. Introduction

Biomass exploitation is emerging as an important component in producing chemicals from renewable sources (Posadino et al. 2012, 2018; Cho et al. 2020; Posadino et al. 2021). In this regard, wild aromatic plants are invaluable sources of new potential drugs. Indeed, plant essential oils (EO) and their constituents have been widely used in the pharmaceutical, cosmetic, food, and beverage industries (Fabian et al. 2006). Different beneficial properties, such as antioxidant, anti-inflammatory, antiviral, antibacterial, antidiabetic and anticancer have been reported for EO, which are increasingly employed in complementary therapies (Ali et al. 2008). Among aromatic plants, Pistacia lentiscus L (PL) has been extensively used in folk medicine for several therapeutic uses, including anti-hypertensive, anti-inflammatory and antiseptic (Bozorgi et al. 2013). PL is an aromatic bush indigenous to Mediterranean and Middle East countries (Milia et al. 2021). PL extracts and EO have shown several properties, including antioxidant, antimicrobial, anticancer and cytoprotective (Loutrari et al. 2006; Benhammou et al. 2008; Xanthis et al. 2021). Among the different extraction processes, supercritical CO_2 (SCCO₂) is emerging as a superior extraction technology since it can be performed at low temperatures, thus protecting matrix components from thermal degradation (Abbas et al. 2008). SCCO₂ emerged as the most desirable technique for separating natural products used in foods and medicines because of its inertness, non-toxicity, critical temperature and low pressure (Starmans and Nijhuis 1996; Santos et al. 2013). Reactive oxygen species (ROS) are recognized as by-products of the aerobic metabolism and essential second messengers regulating vital cellular functions. ROS's physiological levels are tuned by the orchestrated action of ROS-generating enzymes and cellular antioxidant mechanisms (Costa et al. 2021). However, dysregulation of the above-mentioned homeostasis can generate oxidative stress, a phenomenon linked to several pathological conditions (Vono et al. 2016; Thuan et al. 2018; Giordo, Ahmed, et al. 2021; Giordo, Thuan, et al. 2021). This linkage suggests that

counteracting oxidative stress with antioxidants might prevent disease occurrence or ameliorate its associated complications, and for this reason, natural antioxidants are now seen as potential therapeutic candidates (Ceriello et al. 2016). However, although plant-derived compounds are recognized as valuable adjuvants or therapeutic tools (Phu et al. 2020; Shaito et al. 2020; Giordo, Nasrallah, et al. 2021; Giordo, Zinellu, et al. 2021), their efficacy and safety in humans are still significant concerns (Posadino et al. 2013; Goszcz et al. 2015; Shaito et al. 2020). Moreover, the extracted compounds' quality may be affected by the technological process employed; therefore, it is essential to test their safety, efficacy, and quality before therapeutic utilization. To our knowledge, no previous reports have investigated the effects of PL leaves SCCO₂ extract on human ECs. In this light, the present work aims to examine the safety and potential cytoprotective, antioxidant and anti-migration properties of a SCCO₂ extract obtained from PL on human ECs.

2. Results and discussion

2.1. Essential oil composition

Percentages of individual components of PL extract were calculated based on gas chromatography (GC) peak areas without flame ionization detection (FID) response factor correction. Chromatogram and main chemical components of PL-EO are reported, respectively, in Figure S1 and Table S1. The analytical results were consistent with those reported in the literature concerning terpene compounds, representing the more important fraction with antioxidant activity (Said et al. 2011). The total EO yield, after 4 h of extraction was 0.14% and the main abundant constituents included germacrene D (11.18%), delta-cadinene (10.54%), alpha-pinene (8.7%), beta-caryophyllene (5.74%), myrcene (4.56%), beta-phellandrene (4.33%), terpin-4-ol (4.306%), epi-alpha-muurolol (3.262%) and beta-pinene (3.02%). The complete profiling of the EO components is reported in Table S1.

2.2. PL extract does not affect ECs viability and proliferation

Using primary human ECs (Zinellu et al. 2009), we investigated the extract's ability to modulate different biological cell functions. Compounds safety is of paramount importance for their therapeutic employment; therefore, we first examined the potential toxicity of the obtained extract by assessing its effects on cell viability, cell proliferation, and intracellular ROS production. Indeed, intracellular ROS generation is closely related to EC survival, proliferation and apoptosis (Shaito et al. 2022). Based on previously reported data (Catalani et al. 2017), we evaluated the possible harmful effects of three extract concentrations [50, 150 and $600 \mu g/mL$] on ECs viability and proliferation. To this end, cells were treated for 24 h with the PL extract, then cell viability and proliferation were assessed as previously reported (Posadino et al. 2019). As depicted in Figure S2A, the PL extract had no toxic effect at any of the tested concentrations indicating the safety of both the extract and the extraction process applied. Likewise, the data in Figure S2B indicate that the tested extract concentrations failed to induce detrimental effects on ECs proliferation, further confirming the extract's safety.

2.3. PL extract showed an antioxidant effect against H_2O_2 -induced oxidative stress

Whether PL SCCO₂ extract can exert antioxidant effects in biological models remains to be elucidated. Therefore, we sought to investigate whether the obtained extract could counteract oxidative stress in H₂O₂-treated ECs. For this purpose, H₂DCFDAloaded cells were pre-treated for 3 h with the indicated extract concentrations and then incubated for 6 h in the presence or absence of H₂O₂ (75 μ M). The data derived from five pooled measurements were expressed as percentages of untreated control cells (Boin et al. 2014). As depicted in Figure S3A, exposure of H₂O₂-treated cells to increasing concentrations of PL extract showed a significant dose-dependent antioxidant effect compared to cells treated with only H₂O₂. We next wondered whether the PL extract per se could exert any antioxidant or prooxidant effect in the absence of oxidative insults. In this regard, exposure of unstressed cells to increasing extract concentrations induced a significant antioxidant effect at 150 and 600 μ g/mL while failing to affect the intracellular redox state at 50 μ g/mL. (Figure S3B). These findings agree with the cell viability and proliferation results showing no extract toxicity up to 24 h of cell treatment (Figure S2A, B).

2.4. PL extract showed a protective effect against H_2O_2 -induced oxidative stress

Oxidative-induced endothelial damage triggers and sustains cardiovascular diseases (CVD) (Shaito et al. 2022). Therefore, much research is now focused on finding natural antioxidants capable of preventing or countering CVD-associated ROS increases. To determine whether the observed extract antioxidant effect could be protective against the H_2O_2 -induced oxidative damage, we measured the ECs viability. To this end, cells were exposed for 3 h to increasing concentrations of PL extract and then incubated for 24 h in the absence or presence of H_2O_2 (75 µM). As indicated in Figure S4, cell exposition to increasing doses of PL extract provided a significant dose-dependent cytoprotective effect with respect to the H_2O_2 -induced cell damage. Indeed, all the tested PL concentrations were able to significantly counteract H_2O_2 -induced cell damage. Consonant with these findings is the data in Figure S3B, reporting the extract's ability to dose-dependently prevent the detrimental effect on HUVEC proliferation elicited by H_2O_2 .

2.5. PL extract showed an antimigratory effect against seruminduced migration

EC migration is an essential step of the angiogenic process; indeed, ECs, which generally are maintained in a quiescent state, are stimulated to degrade the basement membrane and migrate into the perivascular stroma in response to either proangiogenic factors or by the downregulation of antiangiogenic factors (Lamalice et al. 2007). For this reason, we investigated the ability of PL extract to modulate ECs migration using the matrigel transfilter cell invasion assays (Pintus et al. 2018). Figure S5 demonstrated that the PL extract dose-dependently counteracted the serum-induced cell migration, eliciting a significant reduction at the doses of 150 and $600 \,\mu\text{g/mL}$.

Acknowledgements

Open Access funding was provided by the Qatar National Library.

Ethics statement

This research does not involve Human Participants and/or Animals.

Authors' contributions

RG, GP, AMP: Conceptualization. RG, AC, MCP, RC, GB, JSR, AMP: Methodology. RG, AC, MCP, RC, GB, AMP: Investigation. GKN, LP, GP: writing—original draft preparation. RG, AC, MCP, RC, GB, JSR, GKN, LP, GP, AMP: writing—review and editing. GP: supervision. GKN, LP, GP: funding acquisition.

Disclosure statement

No potential conflict of interest was reported by the authors..

Funding

This work has been made possible thanks to grants from Progetto Fondazione di Sardegnabando 2022-2023 and FAR2020-Pintus.

ORCID

Roberta Giordo D http://orcid.org/0000-0002-2187-4458 Javad Sharifi-Rad D http://orcid.org/0000-0002-7301-8151 Gheyath K. Nasrallah D http://orcid.org/0000-0001-9252-1038 Gianfranco Pintus D http://orcid.org/0000-0002-3031-7733 Anna Maria Posadino D http://orcid.org/0000-0002-6155-159X

Data availability statement

The data presented in this study are available in this article.

References

- Abbas K, Mohamed A, Abdulamir A, Abas H. 2008. A review on supercritical fluid extraction as new analytical method. Am J Biochem Biotechnol. 4(4):345–353.
- Ali BH, Blunden G, Tanira MO, Nemmar A. 2008. Some phytochemical, pharmacological and toxicological properties of ginger (*Zingiber officinale* Roscoe): a review of recent research. Food Chem Toxicol. 46(2):409–420.
- Benhammou N, Bekkara FA, Panovska TK. 2008. Antioxidant and antimicrobial activities of the *Pistacia lentiscus* and *Pistacia atlantica* extracts. Afr J Pharm Pharmacol. 2(2):022–028.
- Boin F, Erre GL, Posadino AM, Cossu A, Giordo R, Spinetti G, Passiu G, Emanueli C, Pintus G. 2014. Oxidative stress-dependent activation of collagen synthesis is induced in human pulmonary smooth muscle cells by sera from patients with scleroderma-associated pulmonary hypertension. Orphanet J Rare Dis. 9:123.

- Bozorgi M, Memariani Z, Mobli M, Salehi Surmaghi MH, Shams-Ardekani MR, Rahimi R. 2013. Five *Pistacia* species (*P. vera*, *P. atlantica*, *P. terebinthus*, *P. khinjuk*, and *P. lentiscus*): a review of their traditional uses, phytochemistry, and pharmacology. ScientificWorldJournal. 2013: 219815
- Catalani S, Palma F, Battistelli S, Benedetti S. 2017. Oxidative stress and apoptosis induction in human thyroid carcinoma cells exposed to the essential oil from *Pistacia lentiscus* aerial parts. PLoS One. 12(2):e0172138.
- Ceriello A, Testa R, Genovese S. 2016. Clinical implications of oxidative stress and potential role of natural antioxidants in diabetic vascular complications. Nutr Metab Cardiovasc Dis. 26(4): 285–292.
- Cho EJ, Trinh LTP, Song Y, Lee YG, Bae H-J. 2020. Bioconversion of biomass waste into high value chemicals. Bioresour Technol. 298:122386.
- Costa TJ, Barros PR, Arce C, Santos JD, da Silva-Neto J, Egea G, Dantas AP, Tostes RC, Jimenez-Altayó F. 2021. The homeostatic role of hydrogen peroxide, superoxide anion and nitric oxide in the vasculature. Free Radical Biol Med. 162:615–635.
- Fabian D, Dusan F, Sabol M, Marián S, Domaracká K, Katarína D, Bujnáková D, Dobroslava B. 2006. Essential oils—their antimicrobial activity against *Escherichia coli* and effect on intestinal cell viability. Toxicol in Vitro. 20(8):1435–1445.
- Giordo R, Ahmed Y, Allam H, Abusnana S, Pappalardo L, Nasrallah GK, Mangoni AA, Pintus G. 2021. EndMT regulation by small RNAs in diabetes-associated fibrotic conditions: potential link with oxidative stress. Front Cell Dev Biol. 9:683594.
- Giordo R, Nasrallah GK, Posadino AM, Galimi F, Capobianco G, Eid AH, Pintus G. 2021. Resveratrol-elicited pkc inhibition counteracts nox-mediated endothelial to mesenchymal transition in human retinal endothelial cells exposed to high glucose. Antioxidants 10(2):224.
- Giordo R, Thuan DTB, Posadino AM, Cossu A, Zinellu A, Erre GL, Pintus G. 2021. Iloprost attenuates oxidative stress-dependent activation of collagen synthesis induced by sera from scleroderma patients in human pulmonary microvascular endothelial cells. Molecules 26(16):4729.
- Giordo R, Zinellu A, Eid AH, Pintus G. 2021. Therapeutic potential of resveratrol in COVID-19associated hemostatic disorders. Molecules 26(4):856.
- Goszcz K, Deakin SJ, Duthie GG, Stewart D, Leslie SJ, Megson IL. 2015. Antioxidants in cardiovascular therapy: panacea or false hope? Front Cardiovasc Med. 2:29.
- Lamalice L, Le Boeuf F, Huot J. 2007. Endothelial cell migration during angiogenesis. Circ Res. 100(6):782–794.
- Loutrari H, Magkouta S, Pyriochou A, Koika V, Kolisis FN, Papapetropoulos A, Roussos C. 2006. Mastic oil from *Pistacia lentiscus* var. chia inhibits growth and survival of human K562 leukemia cells and attenuates angiogenesis. Nutr Cancer. 55(1):86–93.
- Milia E, Bullitta SM, Mastandrea G, Szotáková B, Schoubben A, Langhansová L, Quartu M, Bortone A, Eick S. 2021. Leaves and fruits preparations of *Pistacia lentiscus* L.: a review on the ethnopharmacological uses and implications in inflammation and infection. Antibiotics 10(4): 425.
- Phu HT, Thuan DT, Nguyen TH, Posadino AM, Eid AH, Pintus G. 2020. Herbal medicine for slowing aging and aging-associated conditions: efficacy, mechanisms and safety. Curr Vasc Pharmacol. 18(4):369–393.
- Pintus G, Giordo R, Wang Y, Zhu W, Kim SH, Zhang L, Ni L, Zhang J, Telljohann R, McGraw KR, et al. 2018. Reduced vasorin enhances angiotensin II signaling within the aging arterial wall. Oncotarget 9(43):27117–27132.
- Posadino A, Biosa G, Zayed H, Abou-Saleh H, Cossu A, Nasrallah G, Giordo R, Pagnozzi D, Porcu M, Pretti L, et al. 2018. Protective effect of cyclically pressurized solid–liquid extraction polyphenols from Cagnulari grape pomace on oxidative endothelial cell death. Molecules 23(9): 2105.
- Posadino AM, Cossu A, Giordo R, Piscopo A, Abdel-Rahman WM, Piga A, Pintus G. 2021. Antioxidant properties of olive mill wastewater polyphenolic extracts on human endothelial and vascular smooth muscle cells. Foods 10(4):800.

- Posadino AM, Cossu A, Giordo R, Zinellu A, Sotgia S, Vardeu A, Hoa PT, Deiana L, Carru C, Pintus G. 2013. Coumaric acid induces mitochondrial damage and oxidative-mediated cell death of human endothelial cells. Cardiovasc Toxicol. 13(3):301–306.
- Posadino AM, Giordo R, Cossu A, Nasrallah GK, Shaito A, Abou-Saleh H, Eid AH, Pintus G. 2019. Flavin oxidase-induced ROS generation modulates PKC biphasic effect of resveratrol on endothelial cell survival. Biomolecules 9(6):209.
- Posadino AM, Porcu MC, Marongiu B, Cossu A, Piras A, Porcedda S, Falconieri D, Cappuccinelli R, Biosa G, Pintus G, et al. 2012. Antioxidant activity of supercritical carbon dioxide extracts of Salvia desoleana on two human endothelial cell models. Food Res Int. 46(1):354–359.
- Said SA, Fernandez C, Greff S, Torre F, Derridj A, Gauquelin T, Mevy J-P. 2011. Inter-population variability of terpenoid composition in leaves of *Pistacia lentiscus* L. from Algeria: a chemoecological approach. Molecules 16(3):2646–2657.
- Santos DT, Albarelli JQ, Beppu MM, Meireles MAA. 2013. Stabilization of anthocyanin extract from jabuticaba skins by encapsulation using supercritical CO₂ as solvent. Food Res Int. 50(2): 617–624.
- Shaito A, Aramouni K, Assaf R, Parenti A, Orekhov A, El Yazbi A, Pintus G, Eid AH. 2022. Oxidative stress-induced endothelial dysfunction in cardiovascular diseases. Front Biosci (Landmark Ed). 27(3):105.
- Shaito A, Posadino AM, Younes N, Hasan H, Halabi S, Alhababi D, Al-Mohannadi A, Abdel-Rahman WM, Eid AH, Nasrallah GK, et al. 2020. Potential adverse effects of resveratrol: A literature review. IJMS 21(6):2084.
- Shaito A, Thuan DTB, Phu HT, Nguyen THD, Hasan H, Halabi S, Abdelhady S, Nasrallah GK, Eid AH, Pintus G. 2020. Herbal medicine for cardiovascular diseases: efficacy, mechanisms, and safety. Front Pharmacol. 11:422.
- Starmans DA, Nijhuis HH. 1996. Extraction of secondary metabolites from plant material: a review. Trends Food Sci Technol. 7(6):191–197.
- Thuan DTB, Zayed H, Eid AH, Abou-Saleh H, Nasrallah GK, Mangoni AA, Pintus G. 2018. A potential link between oxidative stress and endothelial-to-mesenchymal transition in systemic sclerosis. Front Immunol. 9:1985.
- Vono R, Fuoco C, Testa S, Pirrò S, Maselli D, Ferland McCollough D, Sangalli E, Pintus G, Giordo R, Finzi G, et al. 2016. Activation of the pro-oxidant PKCβll-p66Shc signaling pathway contributes to pericyte dysfunction in skeletal muscles of patients with diabetes with critical limb ischemia. Diabetes 65(12):3691–3704.
- Xanthis V, Fitsiou E, Voulgaridou G-P, Bogadakis A, Chlichlia K, Galanis A, Pappa A. 2021. Antioxidant and cytoprotective potential of the essential oil *Pistacia lentiscus* var. chia and its major components myrcene and α-pinene. Antioxidants 10(1):127.
- Zinellu A, Sotgia S, Scanu B, Pintus G, Posadino AM, Cossu A, Deiana L, Sengupta S, Carru C. 2009. S-homocysteinylated LDL apolipoprotein B adversely affects human endothelial cells in vitro. Atherosclerosis 206(1):40–46.