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# Impact of Phytoconstituents on Oral Health Practices: A Post COVID-19 Observation

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**ABSTRACT:** Appropriate oral hygiene significantly reduces the possibility of oral infections. However, dental caries and periodontal diseases are major oral health issues causing chronic diseases due to poor oral health. Recently, herbal compounds have gained interest in maintaining oral health. Extracts of burdock root (*Arctium*), noni fruit (*Morinda citrifolia*), and neem leaf (*Azadirachta indica*) are now used as intracanal medicaments in endodontics and periodontics. *Plectranthus amboinicus* species and other plants produces essential oil like  $\beta$ -caryophyllene, p-cymene, and  $\gamma$ -terpinenecan exhibiting antibacterial activity, highlighting phytoconstituents plays a vital role in oral health. The COVID-19 pandemic highlighted the importance of hygiene and sanitization, to curb SARS-CoV-2. Oral cavity is among the gateways for virus entry into saliva. Saliva is a potential reservoir of SARS-CoV-2, and there is an increased risk of infection if there is any fissure in the mouth. This enables entry of virus into the vascular system through gingival or periodontal pocket, possibly reaching lung periphery then to lung vessels by interacting with endothelial surface receptors triggering pulmonary vasoconstriction and lung damage due to endothelial dysfunction. This review aims to draw attention to the possible route of SARS-CoV-2 infection via the oral cavity and the importance of oral hygiene against COVID-19.

Keywords: COVID-19; SARS-CoV-2; oral; hygiene; microbiota; Phytochemicals.

# 1. Introduction

# 1.1 Importance of Oral Hygiene

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Oral well-being is a crucial aspect of overall health. Dental caries (tooth decay), oral cancers, and periodontal disease (gum disease) are the most prevalent chronic oral disorders affecting many of the world'syoung and adult populations. Oral health was recognized as a significant global public health issue at the United Nations summit on preventing non-communicable diseases[1]. Periodontal disease and dental caries are the major chronic diseases, with dental caries ailing people from infancy to old age at any stage. Even though most adults have some gingival inflammation (gingivitis), around 11% of the global adult population has severe progressive periodontal diseases, leading to tooth loss at an early age[2]. According to a recent global burden of disease survey, oral disorders are responsible for 15 million disabilities globally (Figure 1)[2].As perAbbasi-Shavaziand his colleagues, the deciding factors for children's oral health-related quality of life (OHRQoL) involvestrengthening, predisposing, and oral health behaviorand status[3]. On the other hand, Zhang and his teammates found that risk predictors responsible for root caries can be categorized based on age, socioeconomic status, and tobacco usage. Moreover, patients with gingival recession, poor oral hygiene, and prior dental caries are more susceptible to developing new root caries[4]. Based on clinical trials, oral diseases are worsened by high-sugar diets, poor oral hygiene, tobacco consumption, lack of fluoride, and excessive alcohol intake. Oral diseases are gradually recognized asbehavioral threats to other major non-communicable diseases[5, 6].



Figure 1. The impact of oral diseases[1]

Oral diseases have a socioeconomic impact on human beings; compromised oral health may impact general health and other disorders associated with oral hygiene; it may have psychological effects like anxiety and fear; it may influence an individual's economic growth. This visual representation provides a compact glimpse of different aspects of oral diseases that impact human well-being.

Lately, the diversity and applications of herbal compounds havesignificantly increased through oral care crops and their conjunction with conventional treatment methodsdue to their physical and medicinal value[7]. These herbal compounds in dentistry are primarily used to relieve tooth pain, canker sores, and gum irritation[8]. Antiseptics and analgesics are two primaryclasses of compounds derived from plantsextensively used in dentistry for various problems[9]. Burdock root, noni fruit, and neem leaf, are now being extensively used as intracanal medicine in endodontics and periodontics, which have enabled the usage of these herbal

agents in dentistry around the world.For instance, clove oil is primarily used to reduce toothache[10]. Interestingly, clove oil relieves toothache and prevents tooth decay via different bacterial species owing to their antibacterial potential.

#### 1.2 Oral Health and COVID-19

Several studies have recently shed light on our understanding of SARS-CoV-2 transmission from the mouth to the lungs and the development of COVID-19(Figure 2)[11]. SARS-CoV-2 uses ACE2 receptors, furin, and trans-membrane protease serine 2 (TMPRSS2) to infect host cells. For instance, TMPRSS2 induces endocytosis by binding viral spike proteins to ACE2 receptors on the host 'cell's surface, whereas furin is involved in the release of viral particles to the extracellular compartment[12]. These infection-fighting mediators are commonly found in the oral cavity and nasal airways, including the tongue, gingival tissues, and minor salivary glands [13, 14]. Moreover, although the three mediators of viral entry are not distributed in all oral tissues, the sulcular epithelium cells express ACE2, furin, and TMPRSS2, suggesting that SARS-CoV-2 can spread via the gingival sulcus. As a result, the virus can infect many niches in the oral cavity, besides the gingival sulcus[14, 15]. Per a recent analysis of autopsy tissues from deceased COVID-19 patients, oral mucosa and salivary glands were infected with SARS-CoV-2[13]. Five of seven COVID-19 patients reported dead in a post-mortem analysis showed viral RNA in the periodontal tissues[16]. Following these observations, the abundance of SARS-CoV-2 in saliva may lead to the infection of gingival cells, salivary gland tissues, and oral mucosal cells. According to Huang et al., SARS-CoV-2 can survive in saliva or the nasopharynx for over two months. After 0.5-3.5 weeks, viral clearance was observed in asymptomatic individuals[17]. The salivary viral load has been related to mortality, loss of smell and taste, and disease severity. These factors are better predictors of poor outcomes than'patients' age or nasopharyngeal viral load[17, 18], and 'patients' age as a potential risk factor is a significant finding[19].

# 1.3 Dentistry and COVID-19 management

COVID-19 have shoot up the risk among the health professionals, since the main hub of this disease initiates from the nasopharynx to mouth and then to the other parts of the body. The plausible guidelines which can be followed by the health professionals which are at higher risk like the dentists and others needs to follow the guidelines which can curb the health risks are: the devices with CAD (Computer Aided Design) and CAM (Computer Aided Manufacturing) technology can be used by the dentists for the production of breathing devices which will reduce the time of the patient and the staff and will transmit a safety message among the patients, this will also reduce the time of invasiveness of the virus [20].

# 1.4 Dental implantations and their maintenance

Airborne transmission of SARS-CoV-2 has regulated the dental health care and implantations, as the procedures followed for it are aerosol generating which aid in risk of producing and with spread of contaminated droplets [21]. It is necessary to have improved ventilation systems in the operating rooms where implants and other regular checkups are being carried to minimize the potential risk through the aerosols. The usage of air purifiers which will reduce the suspended particles present in the room using Highly Efficient

Particulate Air (HEPA) filters. It has been studied in a clinical trial that the use of HEPA filters in the operating rooms reduces the risk of transmission by aerosols to the greater extent[22].

The dental implants have increased tremendously due to less complications and high success rates [23]. In a study functional implant prosthodontics score was calculated over 30 individuals who have undergone soft tissue implants in posterior sites and were followed up for an year indicated great potential of both conical and hexagon connections and their good performances with one year of clinical services [24]. Although implants can also be ruined by poor oral health which subsequently results in periodontal inflammation leading to failure of secondary implant caused due to peri-implantitis. These failures are mainly due to plaque index, bleeding on probing and pocket of depth which leads to implant failure [25]. These implants can be either prosthetic rehabilitation with title, which is an efficient implantation technique for oral cavities such individuals also might get affected if a patient suffers from COVID-19 duration of implant alterations in the ACE2 pathway, inflammatory cytokine storm or due to microvascular dysfunction [26]. The intricacy increases if an individual is diabetic due to large wounds, delay in the healing process, and greater force is applied during the implantation process [27, 28]. Since, Covid-19 patients are more susceptible to plaque, gingivitis and bleeding and poor oral hygiene might have aided in difficulties in maintaining the dental implantation during the pandemic. One of the other major factors is to prevent microeakage from the implant so that infection can be controlled for which the implant-abutment connection should be resistant against bacterial microleakage resulting in stopping infections and success of implantation [29]. If hygiene would have been maintained along with periodic checkups might help reducing the chances of implant failures and other complications caused.

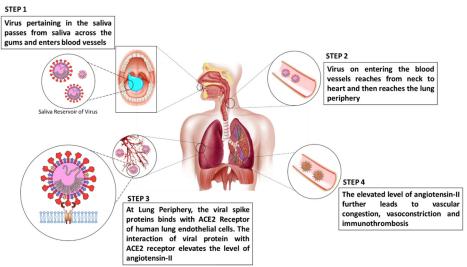


Figure 2.COVID-19 pathway: a hypothetical model for the oral-vascular-pulmonary route of infection [Adopted from[11]]

Diagrammatic representation shows the putative oral passage of SARS CoV infection via saliva and its entry into blood vessels leading its way to the lung periphery. The virus may elevate angiotensin-II levels as ACE2 receptors facilitate viral proteins (spike proteins). This elevation may further lead to the progression of congestion, vasoconstriction, and immunothrombosis.

In earlier findings, Epstein-Barr virus (EBV), Zika virus, human immunodeficiency virus (HIV), herpes simplex virus (HSV), cytomegalovirus (CMV), and some other human viruses have been foundin gingival crevicular fluid (GCF), gingival tissues, saliva, and subgingivalplaque[30-34]. Recently, viral RNA has been

found in the GCF of 64% of COVID-19 patients[35]. In the oral cavity, viral particles migrate to the gingival sulcus/periodontal pockets, which are more suitable for survival. In patients with periodontal disease, the periodontal pocket epithelium develops micro-ulcerations that enable microorganisms and viral particles to move through connective tissue and gingival capillary complex, eventually reaching the systemic circulation[15, 16]. In moderate periodontitis, the region of exposed connective tissue and related blood vessels directly contacting the sub-gingival biofilm is  $5 \text{ cm}^2$ , while in severe periodontitis, it is more than 20 cm<sup>2</sup>[36].Peripheral blood neutrophils in patients with periodontal disease have been shown to have type-1 interferon gene expression signatures, consistent with intravascular exposure to periodontal bacteria, fungi, and viruses[37]. As a result, periodontal pockets can provide favourable conditions for viral replication, infection, and dissemination to gingival capillaries. Studies on bacteriemiaindicated the presence of oral bacteria in the systemic circulation and infective endocarditis of oral origin. These studies suggest that oral bacteria can harm the body, witha higher risk in individuals with poor oral hygiene and periodontal inflammation[38]. Since bacteria can enter the systemic circulation through a mouth lesion, this pathway is used by the SARS-CoV-2 viruses enabled by periodontal disease. In this context, this review highlights the potential route of viral infection, focusing on COVID-19 via the oral cavity and the importance of oral hygiene against such infections. The application of simple, inexpensive measures, such as toothpaste and mouthwashes, is underlined. Furthermore, usingvarious phytoconstituents to oral hygiene products to restrain salivary viral load and mitigate viral infection is also stressed.

# 2. Oral Microbes and Their Effects

#### 2.1. Oral Microbiota

Miller first suggested the "chemical bacteria hypothesis" for dental caries in his book "Microbes in the Human Mouth" in 1890, implying that the dental biofilm is made up of microorganisms[39]. Oral microorganisms, like viruses, bacteria, mycoplasmas, yeasts, archaea, and protozoa, form an ecological environment in the mouth called the oral microbiota[40, 41]. The oral cavity offers a warm and healthy habitat for the oral microbiota while controlling bacterial colonization to prevent invasion by pathogenic microbes. Moreover, oral microbiota is essential for maintaining good oral health[42]. On the other hand, invading microorganisms may induce an imbalance in the commensal microbial community of the host, thereby resulting in dental diseases.

# 2.2. Dental Biofilms

Dental biofilms are a polymicrobial community formed by oral microbiota on the surface of teeth[43]. The extracellular polymeric substances (EPS) matrix provides pathogenic habitat for dental caries, causing microorganisms. Dental caries ismainly a biofilm-induced disease, not an infectious disease, as it begins with the biofilm surrounding the tooth's surface[44, 45]. Due to a highly active and complex ecosystem with abundant EPS, the biofilm can stimulate caries. Indeed, a biofilm begins to develop when a salivary glycoprotein film, called a dental pellicle, surrounds the tooth surface[46]. Gram-positive bacteria, such as

*Streptococcus mutans* and *Streptococcus mitis* species (known as the biofilm's initial colonizers), produce the EPS, improving the adherence of other microorganisms[47, 48]. Acid-producing bacterial species from the genera *Lactobacillus*, *Scardovia*, *Propionibacterium*, and *Veillonella* can trigger cariogenic conditions in the mouth and colonize the dental biofilm[49-52]. However, other acid-producing microorganisms use the EPS to create new binding sites and increase virulence[47].

The microbial composition of cariogenic biofilms has been the subject of intense research. Still, it is now generally acknowledged that the biochemical and structural properties of the EPS play a role in the etiology of dental caries[53]. The EPS matrix protects the biofilm and provides mechanical stability, rendering it resistant to antimicrobials. The microbes are lodged in an EPS substrate and continuously produce acids, which are physically shielded from saliva's rapid buffering[54]. Specifically, research on dental caries has been focused on microbial behavior in biofilm communities employing experimental biofilm models, which can imitate metabolic processes through carbohydrate absorption in the mouth and determine the dose-response sensitivity of anti-caries agents[44]. This approach aids in investigatingthe cariogenicity of dietary sugars and evaluates their anti-caries effects[55].

Biofilm of *S. mutans* is believed to have cariogenic potential based on three main characteristics: (i) acid production, (ii) capability to produce EPS, which has a growth-promoting mechanism that protects cells and allows them to thrive in harsh environments, and (iii) acid resistance, which allows conversion of carbohydrates into organic acids while still thriving in low pH environments[56-58]. Moreover, the matrix-producing glucosyltransferases (GtfBCD) in *S. mutans* involved in cariogenic biofilm formation[59]. Although much research has covered the treatment and prevention of dental caries, it is apparent that merely limiting sugar consumption and avoiding *S. mutans* and fructans formed by *Streptococcus gordonii, Streptococcus salivarius*, and *Actinomyces* species are vital EPS components in cariogenic biofilms[50, 53].

Recently, molecular studies have detected the presence of pathogenic flora, which include bacteria like *Streptococcus, Actinomyces* and *Scardoviaspp.*, and fungi, like *Candida albicans*, in dental biofilms[60, 61]. In contrast to *S. mutans*, dental caries-associated colonizers include *Scardovia*, *Lactobacillus*, and *Bifidobacterium*species[50, 51, 62]. Earlier studies have shown that biofilm resistance to anti-adhesion compounds, antibiotics, and preservatives isdirectly linked to microbial diversity[63-65]. Microbial abundance declines during the maturation of cariogenic biofilms due to the competition between cariogenic microorganisms[53]. Hence, the supremacy of cariogenic microorganisms over health-associated commensal species is thought to be the cause of dental caries. Indeed, both the complexity of the biofilm matrix and the proliferation of microorganisms pose challenges in preventing and managingdental caries.

# 2.3. Microbial Ecology and Dental Caries

The long-term association between fermentable carbohydrates and acid-producing microorganisms is the key reason for dental caries[40]. The oral microbiome plays a role in developing dental caries and some host

factors like saliva and the physical properties of oral hard tissues. Dental biofilm plays a significant role in the etiology of dental caries. Caries are triggered by the matrix's complexity, the transmission of resistance genes, and the physical security offered by the EPS matrix. Based on the studies, the main associated issues include the lack of a single apparent therapeutic target and poor retention of locally administered therapies. Hence, controlling dental biofilm is best to avoid tooth decay[47, 66].

# 2.4. Microbiota in Periodontal Diseases

In 1960, research on the relationship between bacterial strains and periodontal diseases began by isolating certain bacterial morphotypes[67]. The discovery of *Aggregatibacteractinomycetemcomitans* as a pathogen associated with localized aggressive periodontitis (LAP) supports the specificity of periodontal microflora[68, 69]. Subsequent cross-sectional and long-term studies bacterial species to health and disease. These studies have established a connection between periodontitis and a small group of periodontal pathogens: *Tannerella forsythia*, *Porphyromonasgingivalis*, and *A. actinomycetemcomitans*. Indeed, they are closely linked to the onset of periodontal disease, progression of periodontal disease, and failure of periodontal therapy. Other bacteria isolated from the subgingivalmicrobiota, like *Prevotellaintermedia*, *Peptostreptococcus micros*, *Eubacteriumnodatum*, *Fusobacteriumnucleatum*, *Campylobacter rectus*, *Treponemadenticola*, and *Prevotellanigrescens*, have also been identified. Still, their etiologic roles are less specific. As a result, periodontal disease may be a polymicrobial infection[67].

# 3. Oral Hygiene Practices

Tooth brushing and interdental cleaning are two of the most popular oral hygiene recommendations[70, 71]. India is a large country with a population of around 1.27 billion. Most people rely on traditional oral hygiene practices, like mango bark or neem tree stick, tobacco leaves-based toothpowder, customized charcoal, or salt recommended in Vedas [72, 73]. Due totheir benefits, numerous healthcare industries have started adding the above constituents to their products and marketing them for their health benefits. The sacred plant, Ocimum sanctum, also known as tulsi, is widely known for its medicinal value as it exhibits antimicrobial and antifungal activity against diverse oral pathogens responsible for dental problems. Nowadays, tulsi extracts are incorporated into toothpaste and mouthwash via pharmaceutical industries to treat pulpitis and toothache[74]. The essential oil produced by Salvadorapersica (miswak) has been reported to possess anti-gingivitis, anti-cariogenic, anti-plaque, promotion of gingival wound healing, orthodontic chain preservation and whitening properties [75]. Moreover, the eucalyptus oil synthesized by Eucalyptus globulushas been reported to encompass a bioactive compound named eucalyptol, used as a mouthwash and endodontic solvent in dental preparation in dentistry. Studies conducted on Azadirachtaindica (Neem) oil and bark have been reported to treat different dental issues via different methods to cure cavities, gum disease, and dental plaque[76, 77]. Different mouthwash formulations consist of neem extract to prevent bleeding and soring of gums and treat tooth decay and oral infection[77]. Varioustoothpaste and mouthwashes available in the world market and their composition are summarized in table1 and 2.

Brand Name	Manufacturer/ Marketed By	Composition	Oral Hygiene Claim
Colgate Total Whole Mouth Health	č	Sorbitol, CI77891, Water, Sodium hydroxide, CI74160, Hydrated silica, Sodium fluoride, PVM/MA Copolymer, Triclosan, Sodium lauryl sulfate, Flavor, Sodium saccharin, Carrageenan	Anti-gingivitis, Anti-plaque, Anti-cavity
Close-Up Anti-cavity Fluoride	Hindustan Unilever Ltd	Hydrated Silica, Water, Sorbitol, PEG-8, Cellulose gum, SD Alcohol 38-B, Sodium lauryl sulfate, Flavor, Red 33, Red 40, Sodium saccharin;Sodium fluoride (0.24%)	Anti-cavity, Fresh breath
PepsodentGermi Check	Hindustan Unilever Ltd	Sorbitol, Water, Calcium carbonate, Hydrated silica, Benzyl alcohol, Sodium lauryl sulfate, Cetylpyridinium chloride, Kaolin, Flavor, Limonene, Sodium monofluorophosphate, CI45430, Sodium saccharin, Sodium silicate, Cellulose gum, Potassium nitrate	Remove plaque, Fresh breath
Sensodyne Rapid Relief	GlaxoSmithKline plc	Strontium acetate, Sodium fluoride, Purified water, Silica, Sorbitol, Xanthan gum, AC1131 Flavour, Glycerol, Titanium dioxide, Sodium methyl cocoyltaurate, Sodium methyl hydroxybenzoate, Saccharin sodium, Precipitated silica, Propyl hydroxybenzoate	Sensitivity relief
Oral-B Pro-Expert	Procter & Gamble	Stannous chloride, Polychelation complex, Sodium fluoride	Anti-cavity, Anti-plaque, Fresh breath
Dabur <i>Meswak</i>	Dabur India Ltd	Carrageenan, Sorbitol, Water, CI 77891, Silica, Sodium silicate, Sodium lauryl sulfate, Calcium carbonate, Flavour, Meswak extract, Cellulose gum, PVM/MA copolymer, p-Thymol, Zinc gluconate, Sodium saccharin, Benzyl alcohol, Sodium benzoate	Antibacterial, Anti-inflammatory, Astringent
PatanjaliDantKanti	PatanjaliAyurved Ltd	Haldi ( <i>Curcuma longa</i> ), Akarkara root extract ( <i>Anacyclus pyrethrum</i> ), Laung oil ( <i>Syzygiumaromaticum</i> ), Vidang fruit extract ( <i>Embeliaribes</i> ), Babool ( <i>Acacia arabica</i> ), Tomar seed oil ( <i>Zanthoxylumalatum</i> ), Bakul ( <i>Mimusopselengi</i> ), Pudina ( <i>Menthapiperita</i> ), Majuphal ( <i>Quercusinfectoria</i> ), Neem bark extract ( <i>Azadirachtaindica</i> ), Vajradanti ( <i>Barleriaprionitis</i> ), Pilu ( <i>Salvadorapersica</i> ), Pippalichhoti ( <i>Piper sylvaticum</i> )	Prevent gingival bleeding and periodontal diseases
ViccoVajradantiAyurvedic	Vicco Laboratories	Vajradanti, JeshthamadhAjwain, Babhul, Jambhul, Laung, Manjishtha, Bor, Akhrot, Dalchini, Khair, Patang, Bakul, Harada, Anantmul, Behada, Amla, Maifal, AkkalKadha, Kavab	Anti-plaque, Remedy for pyorrhea
Dabur Red	Dabur India Ltd	PudinaSatva, Clove oil, Ginger, TomarBeej (Zanthoxylumalatum)	Anti-plaque, Fresh breath
Himalaya Complete Care Lacalut Extra Sensitive®	Himalaya Global Holdings Ltd TheissNaturwaren, 66424 Homburg, Germany	Neem, Indian gum Arabic, Triphala, Bishop's Weed, Pomegranate fruit rind, Pepper, False Black Sodium fluoride, Aluminium salts, Chlorhexidine, KCl, silicium dioxide, Sodium fluoride Amine	Germ-free mouth Improvement of nerve cells, Caries prevention
Biomed Sensitive®	Splat Oral Care, 121099 Moscow, Russia	L-Arginine, Hydroxyapatite, Natural component (Plantain extract, birch leaf polyphenols and red grape seeds)	Enamel strengthening and eliminating the causes of tooth sensitivity
Aslamed for Sensitive Teeth®	Farmec SA, 400616 ClujNapoca, Romania	Sodium fluoride, special clay, potassium nitrate, sodium lauryl sulfate (SLS) free	Remineralization of teeth and strengthens their enamel, astringent effect

Table 1 Toothpaste that is available in the world market and their composition[78].

The table is intended to summarize the different kinds of toothpaste available in the global markets and their compositions and claims regarding oral hygiene decreed by the brands.

	Table 2 Mouthwashformulations in the world market and their composition				
Brand Name	Manufacturer/ Marketed By	Composition	Oral Hygiene Claim		
Listerine Cool Mint	Johnson & Johnson Pvt. Ltd	Thymol (0.064%), Menthol (0.042%), Methyl Salicylate (0.060%), Eucalyptol (0.092%); FD&C Green 3 CI 42053, Water, Sodium Benzoate, Alcohol (21.6%), Sorbitol Solution, Sodium Saccharin, Poloxamer 407, Benzoic Acid	Kills 99.9% of germs		
Colgate Plax	Colgate- Palmolive	Propylene Glycol, Glycerine, Polysorbate 20, Sodium saccharin, Sodium benzoate, Sodium fluoride, Sorbitol	Kills 99% of germs		
Closeup Red Hot	Aero Pharma/ Hindustan Unilever Ltd	Sodium saccharin, Sorbitol, Water, PEG-40 Hydrogenated, Potassium citrate, glycine, Sodium lauryl sulfate, Benzyl alcohol, Phenoxyethanol, C116035, Zinc sulfate, Eugenol, <i>Eugenia caryophyllus</i> leaf oil (Clove), Sodium fluoride, Castor Oil	Kills 100% of germs		
Dabur Red Pulling Oil	Dabur India Ltd	Sesame oil, Coconut oil, Thyme, Cinnamon, Mint, Tulsi, Clove	Kills 99.9% of germs		
Amflor	Group Pharmaceutical Ltd	Purified water, Sodium benzoate, Sorbitol, Polyoxyl 40 hydrogenated castor oil, Ponceau 4R, Propylene glycol, Flavours, Amine fluoride, Poloxamer, Sodium saccharin	ND		
Bioayurveda Basics	ArganshePvt Ltd	Turmeric, Tea tree oil, Basil, Lemon, Mint, Clove, Ginger, Mint	ND		
Bio Resurge	Bio Resurge Life Coaching Health Services Pvt. Ltd	Aqua, Clove oil Peppermint, Alum, <i>Rubiacordifolia</i> (Indian madder), <i>Glycyrrhizaglabra</i> (Mulethi), <i>Terminaliabellirica</i> (Bahera), <i>Commiphorawightii</i> (Guggal)	ND		
Cur Q Fresh	Onika Organics	Curcumin, Thymol oil, Tea tree oil, Mint, Clove oil, Tulsi, and Eucalyptus oil	ND		
Organic Aloe Vera	Dr. Organic Ltd	<i>Aloe vera</i> , Icelandic moss extract, Peppermint oil, Tea tree leaf oil and Arnica extract, Apple fruit extract, Indian pennywort extract	ND		
Himalaya HiOra-K	The Himalaya Drug Company	Clove, naturally-derived Potassium nitrate (Suryakshara)	ND		
Listerine- Vanilla Mint	Pfizer Inc Lilitz, Pennsylvania, USA	Thymol, menthol, eucalyptol, methyl salicylate	ND		
Listerine Tartar Control	Johnson & Johnson Industrial Ltda., São Paulo, Brazil	Thymol, menthol, eucalyptol, methyl salicylate	ND		
Listerine Fresh Burst	Johnson & Johnson Industrial Ltda., São Paulo, Brazil	Thymol, menthol, eucalyptol, methyl salicylate	ND		

ND: Not Defined, Table is intended to summarize the different mouthwash available in the global markets and their compositions and claims regarding the oral hygiene decreed by the brands. Interestingly, most of the products have phytochemicals in them.

# 4. Bioactive Compounds in Dentistry: Focus on Antibacterial and Antifungal activities

Dental plaque and gingivitis can be managed by brushing with anti-plaque agents containing dentifrice and inter-dental cleaning with dental floss and toothpicks. A proximal brush is usually prescribed to cleaninterdental spaces[71].Competent plaque management by a dental hygienist has been shown to help maintain a safe periodontium[79]. Mouth rinses can also helpprevent plaque formation by reducing the rate at which bacteria reproduce in plaques and by adhering to dental surfaces,thereby enhancing oral health[80].

Various bioactive compounds have been widely exploited in mouthwashes and toothpaste. These bioactive compounds are volatile and abundant in essential oils (EOs). Briefly, EOs are a rich pool of naturally-occurring bioactive agents with antimicrobial effects, and many of them are used to treat various diseases[81, 82]. Most EOsare secondary metabolites produced by plants to protect against pests, microorganisms, and weather adversity[83, 84]. EOs account for over 3,000 of 100,000 recognized secondary metabolites, with around 300 having commercial importance and extensively used in the pharmaceutical, food, and cosmetic industries[81]. Based on their chemical structures, EOs are divided into two groups: terpenes (sesquiterpenes and monoterpenes) and terpenoids (isoprenoids); both have distinct biosynthetic origins, aromatic and aliphatic compounds (phenols and aldehydes), and low molecular weights[83, 85]. Monoterpenes account for most EOs, and their antibacterial activities against dental caries-causing microorganisms have been documented [82, 86]. Yet, only a few studies on EOs explored their wide dental-care applications. Since only a few compounds from these phytochemical groups have been used in anti-plaque and anti-gingivitis mouthwash formulations, further research iswarranted to explore utilizing EOs in anti-caries chemotherapy [87-89]. We would also like to highlight, using constant mouth wash may alter the microbiome of the mouth but the evidence-based work on SARS-CoV infected individuals showed reduced viral load[90]. Even though, solid evidence and relationship has still been determined but we speculate using mouth wash during the infection period may vary the micro flora but not in that extent where mouthwash is used as a routine practice.

# 4.1 Eugenol

Eugenol is a versatile natural compound with powerful health-promoting properties. It is found in cinnamon, cardamom, cloves, nutmeg, and variousother plants[91]. It was first isolated from the leaves and buds of clove-like *Eugenia caryophyllata*[91].Moreover, eugenol is synthesized by allylation of guaiacol with allyl chloride, which exhibits the same functional properties as eugenol. The presence of eugenol in the extracts of medicinal herbs has triggered the interest of many researchers and opened the door for further studies to assess its therapeutic potentialagainst many diseases.Eugenolhasanaestheticproperties, neuro-protective properties, antimicrobial activity, hypolipidemicaction, antioxidant activity, anti-carcinogenic effects, anti-diabetic, and anti-inflammatory properties[91].

According to the World Health Organization (WHO), eugenol is non-mutagenic and generally recognized as a safe (GRAS) compound. Clove buds, tulsi leaves, cinnamon bark and berries, pepper, turmeric, ginger, thyme, and oregano are other sources of eugenol[91]. However, several other aromatic herbs, such as nutmeg, marjoram, basil, mace, and bay, have been shown to contain substantial amounts of eugenol. Cinnamon (20-50%) and clove (45-90%) are the most abundant plant sources of eugenol. However, commercial eugenol extraction and higher production costs are the significant issues associated with these sources[91]. On the contrary, bay, pepper, tulsi, and ginger are inexpensive and abundant sources that can be utilized instead of cloves and cinnamon. For example, in Gram-negative and Gram-positive bacteria, eugenol damages cell membranes and cell walls, causing cell lysis and leakage of intracellular fluid and lipid and protein contents[92, 93]. *In vivo* and *in vitro* studies on biofilms have shown that eugenol has a significant eradicative and inhibitory effect against many oral microorganisms (**Table 3**).

# 4.2. Thymol

Thymol, known as 5-methyl-2-isopropylphenol and 2-isopropyl-5-methylphenol, is a crystalline, colourlessmonoterpenoid phenol with a distinct odour. It is also an isomer of carvacrol and is the main active ingredient in thyme oil, obtained from *Thymus vulgaris* species. Thymol has been isolated from a variety of plants, including *Origanum* L., *Trachyspermumammi* (L.), *Ocimumgratissimum* L., genus *Monarda* L., *Satureja* L. (Lamiacaeae), *Oliveriadecumbens* Vent, *Anemopsiscalifornica* (Saururaceae), *Carumcopticum* L. (Apiaceae), species of *Ranunculaceae*, *Verbenaceae*, and *Scrophulariaceae* [88]. Different communities in many countries use these extracts as medicinal plants. *T. vulgaris* EOs and others containing thymol have been shown to have expectorant, antitussive, digestive, stomachic, antispasmodic, and carminative action against oral diseases like dental caries[94]. These pharmacological applications, modes of action, and pharmacokinetic studies suggest that thymolmay be a potent, natural therapeutic agent with a high medicinal value[95, 96].

Commercial pharmaceuticals are available with Shirazi thyme (*Zatariamultiflora*) EO as the main ingredient widely used in treating respiratory diseases[97]. Listerine is another well-known thymol-containing medication named after Joseph Lister, who identified the medicinal value of thymolin treating oral and throat infections, including gingivitis[95]. Thymol-containing EOs exert antibacterial activity against Gram-negative (*Escherichia coli, Proteus mirabilis, Pseudomonas aeruginosa, Salmonella typhimuium* Ty2) and Gram-positive (*Bacillus subtilis, Bacillus cereus, Streptococcus faecalis,* and *Staphylococcus aureus*) bacteria[98-100]. The antimicrobial action of thymol is mainly due to its ability to incorporate itself within the lipid bilayer of the host cell, increasing its surface curvature (**Table 3**). Briefly, the hydrophilic parts of thymol molecules interact with the polar region of the membrane. At the same time, the aliphatic side chains and hydrophobic benzene rings fall into the biological membrane's inner region. The destabilization of the lipid bilayer increases fluidity and decreases elasticity, leading to significant changes in the membrane structure. This process causes increased permeability to hydrogen and potassium ions and affects internal membrane proteins, including membrane receptors and enzymes. Following incorporation into the cell membrane,

thymol interacts with the embedded proteins through non-specific interactions, causing changes in the conformation and activity of internal and membrane proteins. As a result, the presence of thymol can trigger cell membrane tension and destabilization[101].

# 4.3 Terpinen-4-ol

Terpinen-4-ol [3-cyclohexen-1-ol,4-methyl-1-(1-methylethyl)-,(R)-] is a terpenethat is considered to be the primary component of *Melaleucaalternifolia*EO, also known as tea tree oil (TTO), and can be found in other plants such as *Alpiniazerumbet* and *Eucalyptus* species from HajebLayoun arboreta (Tunisia)[102, 103]. TTO comprises 100 different compounds, including the monoterpenesterpinolene, 1,8-cineole, and terpinen-4-ol, which account for at least 30% of the oil. TTO has anti-inflammatory, antimicrobial, and anticancer properties and was first reported to exert medicinal effects in the 18<sup>th</sup>century[104]. TTO works against Gram-positive,and Gram-negative bacteria,exhibits broad-spectrum activity, avoids antimicrobial resistance, and is effective against multi-resistant microorganisms[105].The antimicrobial properties of terpinen-4-ol are due to itslipophilicity, allowing it to pass through cytoplasmic membranes and cell walls. Internal osmotic pressure causes the cell wall to weaken, rupture the membrane, and lose the cytoplasmic content[106, 107].

In fungi, terpinen-4-ol has hydroxyl groups, making them moderately water-soluble. This helps them pass through water, penetrate cells, and disrupt cell membranes, resulting in osmotic shock[108]. The addition of TTO and terpinen-4-ol in oral care products, such as toothpaste and mouthwashes, has been demonstrated to exert antiseptic effects due to their ability to adhere to dental biofilms and inhibit bacterial growth[109-112].

# 4.4 Curcumin

Curcumin (diferuloyl methane) is a polyphenolic compound isolated from the rhizomes of *Curcuma* longa L. It is the natural yellow pigment present in the roots of turmeric (family Zingiberaceae). It accounts for roughly 4% of thedry weight[113],*i.e.*, [1,7-bis (hydroxyl-3-methoxyphenyl)-1,6-heptadiene-3,5-dione](C<sub>21</sub>H<sub>20</sub>O<sub>6</sub>) being one of the essential active ingredients. Turmeric has been a culinary spice in curry for centuries in many Asian countries to give its distinct colour and flavour[114]. Turmeric is a food additive and acolouring agent in yellow mustards, cosmetics, pharmaceuticals, and hair and fur dyes[115].

Curcumin has been shown to alter gene expression and prevent bacterial DNA replication (**Table 3**). Moreover, it destroys bacterial cell membranes and decreases the motility of several microorganisms[116]. *In vitro* studies have shown that curcuminpreventsFtsZprotofilament polymerization and GTPase activity disruption in the cytoskeleton of *E. coli*, *S. aureus*, and *B. subtilis*[117, 118]. Moreover, it can affect bacterial cell division and proliferation through this mechanism. According to some studies, curcumin induces an apoptosis-like response in *E. coli*[119]. The downregulation of  $\Delta^{5,6}$ -desaturase (ERG3) was discovered to be a potential mechanism underlying the antifungal activity of curcumin, resulting in a substantial reduction in ergosterol in fungal cells. Reduced ergosterol levels lead to the accumulation of biosynthetic precursors of ergosterol, which causes cell death by releasing reactive oxygen species (ROS)[120]. Other potential effects of the antifungal activity of curcumin include decreased proteinase secretion and a change in membrane-associated properties of ATPase activity[121].

Bacteria (Nature)	MIC	Refer
Dacteria (Nature)	WIIC	nce
Porphyromonasgingivalis ATCC33277 (RS)	31.25 μM	[122]
Streptococcus salivarius (C)	>1000 µg/mL	[123]
Streptococcus mutans (C)	$>1000 \mu g/mL$	L ]
Streptococcus sanguinis (C)	$>1000 \mu g/mL$	
Streptococcus sobrinus ATCC33478 (RS)	600-800 µg/mL	
Staphylococcus epidemidis (C)	$>1000 \ \mu g/mL$	
Corynebacteriumpseudodiphtheriticum (C)	$>1000 \mu g/mL$	
Corynebacteriumxerosis (C)	$>1000 \ \mu g/mL$	
Lactobacillus salivarius ATCC11741 (RS)	$>1000 \ \mu g/mL$	
Rothiadentocariosa ATCC17931 (RS) Neisseria	$>1000 \ \mu g/mL$	
bflava ATCC49275 (RS)	$>1000 \ \mu g/mL$	
Streptococcus mutans MTCC497	0.625 µg/mL	[124]
Staphylococcus aureus B234 (C)	256 µg/mL	[124]
Staphylococcus aureus B294 (C)	128 μg/mL	
Staphylococcus aureus B598 (C) Staphylococcus aureus B147 (C)	512 μg/mL	
Staphylococcus aureus B193 (C)	128 μg/mL	
Staphylococcus aureus, B295 (C)	32 μg/mL	
Staphylococcus aureus, B295 (C) Staphylococcus aureus B285 (C)	64 μg/mL	
Staphylococcus aureus B285 (C) Staphylococcus aureus B456 (C)	256 μg/mL	
Streptococcus mutans B200 (C)	64 μg/mL	
Streptococcus mutans B200 (C)	$64 \ \mu g/mL$	
1		
Streptococcus constellatus B629 (C)	$64 \ \mu g/mL$	
Enterococcus faecium P1 (C)	256 μg/mL	
Enterococcus feacalis P2 (C)	512 μg/mL	D. (
Fungi (Nature)	MIC	Refer nce
Candida dubliniensis 131 (C)	750 μg/mL	[126]
Candida dubliniensis 219 (C)	375 µg/mL	L 3
Candida dubliniensis 248 (C)	375 µg/mL	
<i>Candida tropicalis</i> 23 (C)	375 µg/mL	
<i>Candida tropicalis</i> 150 (C)	750 μg/mL	
<i>Candida tropicalis</i> 176 (C)	375 μg/mL	
b) Thymol	eve yg mil	
Bacteria (Nature)	MIC	Refei
		nce
Aggregatibacteractinomycetemcomitans	100 μg/mL	[127]
TCC33384 (RS)		
Streptococcus mutans ATCC25175 (RS)	200 μg/mL	
Methicillin-resistant Staphylococcus	200 µg/mL	
ureus(MRSA) ATCC33591 (RS)		
EscherichiacoliATCC10798 (RS)	200 μg/mL	
Staphylococcus aureus B234 (C)	64 μg/mL	[125]
Staphylococcus aureus B398 (C)	64 μg/mL	[1=0]
Staphylococcus aureus B147 (C)	256 μg/mL	
	64  µg/mL	
Staphylococcus aureus B147 (C) Staphylococcus aureus, B193 (C) Staphylococcus aureus, B295 (C)	64 μg/mL 32 μg/mL	

Table 3 Antimicrobial activity of EOs against various oral bacteria and fungi

Staphylococcus aureus B456 (C)	256 μg/mL	
Streptococcus mutans B200 (C)	32 µg/mL	
Streptococcus mutans B509 (C)	32 µg/mL	
Streptococcus constellatus B629 (C)	64 µg/mL	
Enterococcus faecium P1 (C)	128 µg/mL	
Enterococcus feacalis P2 (C)	64 µg/mL	
Streptococcus mitis (C)	5 µg/mL	[128]
S. sanguis (C)	5 µg/mL	
S. salivarius (C)	5 μg/mL	
Fungi (Nature)	MIC	Refere
		nce
Candida albicans CBS562 (C)	39 µg/mL	[129]
Candida tropicalis(C)	78 μg/mL	
Candida krusei(C)	3 µg/mL	
c) Terpinen-494		
Bacteria (Nature)	MIC	Refere
		nce
Streptococcus mutans ATCC25175 (RS)	0.24%	[130]
Lactobacillus acidophilus ATCC4356 (RS	) 0.27%	
Streptococcus mutans ATCC25175 (RS)	44000 μg/mL	[131]
Streptococcus salivarius ATCC13419 (RS)	) $2750 \mu g/mL$	
Porphyromonasgingivalis ATCC33277 (R	S) 44000 µg/mL	
Pseudomonas aeruginosaATCC 27853 (RS		
Staphylococcus aureusATCC 25923 (RS)	44000 µg/mL	
Escherichia coli ATCC 25922 (RS)	11000 µg/mL	
Fungi (Nature)	MIC	Refere
		nce
Candida albicans ATCC1031 (RS)	11000 µg/mL	[131]
Candida albicans (C)	0.0625-0.5%	[132]
d) Curcuntin		
HO	V YOH	
Bacteria (Nature)	MIC	Ref
Porphyromonasgingivalis OMZ314 (C)	10 µg/mL	[133]
Streptococcus mutans ATCC35668 (RS)	333.33 µg/mL	[134]
Actinomyces viscosusATCC10048 (RS)	167.67 µg/mL	
Lactobacillus caseiATCC334 (RS)	125 μg/mL	
Porphyromonasgingivalis ATCC32277 (R		
Prevotellaintermedia ATCC25611 (RS)	208.33 µg/mL	
Streptococcus mutans ATCC25175 (RS)	128 μg/mL	[135]
· · · · ()		[]

RS: Referred Strain; C: Clinical; NA: Not Applicable.

# 5. Possible use of eugenol, thymol, terpinen-4-ol, and curcumin against SARS-CoV-2

Eugenolcan inhibit viral infection and replication in herpes simplex viruses in a specific way (**Table 4**). Eugenol prevents the replication and autophagy of the influenza A virus by inhibiting the initiation of IKK/NF-κB, p38MAPK, and ERK signalling pathways[136]. For example, Silva and colleagues used molecular docking techniques to test the potential anti-SARS-CoV-2 efficacy of eugenolagainst various SARC-CoV-2 targets. According to docking ratings, eugenol has a binding affinity for main protease (M<sup>pro</sup>), SARC-CoV-2 spike (S) protein, human ACE-2 protein, and RNA-dependent RNA polymerase[137]. SARS-CoV-2 pulse glycoprotein crystal structure was also determined in detail[138]. The S protein is made up of two sections, S1 and S2. S1 is a receptor-binding domain (RBD), which binds to the peptidase domain of angiotensin-converting enzyme 2 (ACE2) and allows the virus to adhere to the host cell's surface.On the other

hand, S2 is responsible for virus-host membrane fusion[139]. S1 subunit is crucial according to clinical aspects as inhibiting RBD can trigger S protein conformational changes, which is the first step in preventing viral infections.

Based on recent studies, thymol has a good affinity for hydrophobic residues surrounding the ligands in the binding pocket, contributing to the stability of the protein-ligand complex[138, 140]. Thymol binds to the SARS-CoV-2 spike receptor-binding domain (PDB ID: 2AJF) and SARS-CoV-2 spike ectodomain structure (PDB ID: 6VYB), preventing the virus from binding to the host receptor ACE2[141]. Terpinen-4-ol binds to the 3CL protease enzyme (**Table 5**)[142].

Curcumin is a potent antiviral compound withvarious antiviral activities against multiple viruses. Because of its rate-limiting involvement in the *de novo* synthesis of guanine nucleotides, the inosine monophosphate dehydrogenase (IMPDH) enzyme has been proposed as a therapeutic target for antiviral and anticancer compounds[143].Curcumin has been shown to exert inhibitory activity against IMPDH function, either in a competitive or a non-competitive manner[144].

Long terminal repeat (LTR) of type 1 human immunodeficiency virus (HIV-1) provirus plays a vital role in transcription. Antiviral drug candidates may block HIV-1 replication by inhibiting LTR activity[145, 146]. Curcumin was an effective inhibitor of HIV-1 LTR-directed gene expression with no significant side effects on cell viability[147]. Curcumin and its derivatives, such as allyl-curcumin, tocopheryl-curcumin, and reduced curcumin, inhibited Tat protein transactivation of HIV-1 LTR by 70-85%, as measured by  $\beta$ -galactosidase activity in HeLa cells[148]. Moreover, curcumin displays anti-influenza efficacy against influenza viruses H1N1, H6N1, and PR8[149]. Results showed that using 30 µM of curcumin reduced virus yield by more than 90% in cell culture. The inhibition of hemagglutinin interaction in H1N1 and H6N1 subtypes illustrated the direct effect of curcumin on viral particle infectivity, as shown by an opioid abuse experiment[149].In addition, in cell culture assays, curcumin significantly reduced HSV-1 immediate-early (IE) gene expression and infectivity[150]. Compared to other natural products(gallic acid, luteolin, naringenin, quercetin, resveratrol, and zingiberene) and controls, docking analysis revealed that curcumin has the most potent interaction with spike glycoprotein and ACE2 receptor[151]. In contrast to the two anti-malarial drugs (Chloroquine and Hydrochloroquine), Gonzalez-Paz and colleagues reported that curcumin strongly binds to 3CL-protease of SARS-CoV-2 and promotes significant structural changes and folding of the viral protease[142].

Compound	Target Viruses*	Mechanism Type	IC <sub>50</sub>	Reference
Eugenol	HSV-1	Intercellular	35µg/mL	[152]
Eugenol	IFV-A (H1N1)	ND	<3.1 µg/mL	[153]
Thymol	HSV-1	Intercellular	0.002%	[150]
Thymol	HSV-1	Intercellular	7μΜ	[154]
Thymol	BVDV	Intercellular	248.56µg/mL	[155]
Terpinen-4-ol	Influenza	Intercellular	0.0025%	[156]
Terpinen-4-ol	HSV-1	Intercellular	0.05%	[156]

Table 4. Anti-viral activity of eugenol, thymol, terpinen-4-ol, and curcumin

Terpi	nen-4-ol	HSV-2	Intercellular	0.05%	[156]
Terpi	nen-4-ol	ECHO 9	Intercellular	0.05%	[156]
Terpi	nen-4-ol	Cox B1	Intercellular	0.05%	[156]
Terpi	nen-4-ol	Polio 1	Intercellular	0.05%	[156]
Terpi	nen-4-ol	Adeno 2	Intercellular	0.05%	[156]
Curcu	ımin	IFV (H1N1), (H6N1)	Intercellular	0.47µM	[149]
Curcu	ımin	ZIKV	Intercellular	1.90µM	[157]
Curcu	imin	CHIKV	Intercellular	3.89µM	[157]

\*HSV, human herpes viruses; IFV, influenza virus; BVDV, bovine viral diarrhoea virus; ECHO, enteric cytopathic human orphan virus; Cox, coxsackievirus; ZIKV,zika virus; CHIKV, chikungunya virus; ND, not determined

 Table 5. Molecular docking analysis of the anti-SARS-CoV-2 activity of eugenol, thymol, terpinen-4-ol, and curcumin

Compound	Target Component	Interacting Amino Acid Residues	Ref
Eugenol	Spike glycoprotein of SARS-CoV-2 (6VXX)	Gln314, Thr315, Asn317, Asn764, Arg765	[151]
Eugenol	Angiotensin-converting enzyme 2 (1R42)	His401, Glu402, Arg514	[151]
Thymol	Spike receptor binding domain (2AJF)	HIS 401, HIS 378, ASP 382, ALA348, ASP 350, TRP 349, PHE 40	[141]
Thymol	Spike ectodomain structure (6VYB)	LEU 865, PRO863, PHE 782, ILE 870, ALA1056, GLY 1059	[141]
Terpinen-4-ol	3C-like protease (3CL <sup>pro</sup> )	Lys102, Asp153	[142]
Curcumin	Spike glycoprotein of SARS-CoV-2 (6VXX)	Thr302, Lys304, Gln314, Thr315, Asn317, Asn764, Arg765, Thr768	[151]
Curcumin	Angiotensin-converting enzyme 2 (1R42)	Ala348, His378, Asn394, Tyr385, His401, Glu402	[151]
Curcumin	3C-like protease (3CL <sup>pro</sup> )	Lys5, Val125, Asp289, Lys137	[142]

6. Unprecedented Challenges associated with COVID-19

With the surge of the SARS-Cov-2 virus rapidly in different parts of the world, it has become essential to look at its architecting and structural identity to decipher the reasons for the 'virus's evolution to form a new variant with mutations in its morphic structure. The development of these variations hasled to the prevalence of new variants. Notably, the arisen variants include Alpha (B.1.1.7), Beta (B.1.351), Gamma (P.1), Delta (B.1.617.2), Lambda (C.37), Mu (B.1621), Eta (B.1.525), Iota (B.1.526), Kappa (B.1.617.1) and currently Omicron (B.1.1.529)[158, 159]. Thus, to understand the mutations of Coronavirus, there is a need to understand the genomic structure of the virus. The SARS-Cov-2 consist of four protein-encoding genes, which are:

1.S Protein-This is a transmembrane protein, also called S-Glycoprotein or the Spike, that is present on the outskirts of the virus; its role is to establish the pathogenicity and range of the host organism. It acts as a medium of entry for the virus by arbitrating the attachment of the virus to the host 'cell's plasma membrane, thereby interacting with the receptor known as Angiotensin-converting enzyme 2 (ACE2) and in some conditions uses CD209L as an alternate receptor[160].

2.M Protein- A membrane protein that dictates the shape of the virus; it also functions as a small transmembrane protein that binds to various proteins[160].

3.N protein-Nucleocapsid is the structural component of the Corona Virus and is bound/attached to the RNA of SARS-CoV-2. It regulates signalling, replication cycle, and immune response toward SARS-CoV-2 infection[160].

4.E protein- Envelope protein interacts with the cell membrane of the host organism to regulate itself in the production and maturation of the virus[160].

By looking into the COVID-19 vaccines genomes, we can ascertain that almost all vaccines except the SARS-CoV-2 viral vaccine are produced based on the genetic makeup of the Spike (S) gene of the SARS-CoV-2 virus and the S gene is concentrated by researchers and pharma companies as the S gene along with its receptor-binding domain (RBD) encodes 1,273 amino acids; therefore the availability of Spike protein is the key element for the pathogenesis of Coronavirus[161]. It also regulates T-cell response, immunity, neutralizing antibody (NAb), and mediating the binding of the 'host's ACE2 receptors with its RBD, which is crucial for infection into the host organism. This is why the vaccines are targeted in this region to neutralize the antibody to safeguard an organism from SARS-CoV-2 infection. The presence of mutations in this site paves the way for the development of variants[162].

The SARS-CoV-2 virus is an RNA virus, and they have an evolutionary change of 10-4 nucleotides per site/year. They mutate over time due to either factor such as genetic drift, natural selection, or environmental influence, therefore playing a pivotal role in strengthening transmissibility, the severity of the virus, its infectivity rate, pathogenicity, transmission, evasion of the immune system and resisting vaccine. The SARS-CoV-2 virus, the mutation in the S protein area includes its RBD region. The ACE2 receptor is the main focus due to its high mutability rate, which is very severe and impacts the vaccination effects [163-165]. Due to the changes in Spike protein, changes in the genome occur, leading to the production of a new kind of virus with changes in its genome known as variants where changes to spike protein exist[166]. The changes to the S protein and RBD region differ among every arisen variant, such as the SARS-CoV-2- $\alpha$  variant exhibits its primary mutations in RBD sites of E484K, S494P, and N501Y;the  $\beta$  variant of this virus makes the mutation in K417N is unique to this variant. The  $\delta$  variant of this virus makes the mutationsidentical to all other lineages. Still, they illustrate three distinct mutations of E156del, R158G and T478K unique to the  $\delta$  variant, making it possible to be one of the deadliest variants[167-169].

The arisen variants have produced a unique challenge for oral health maintenance.Keeping hygienic oral health is become a necessity, as it is known that the SARS-Cov-2 virus is present in the nasal and oral cavity.This allows variants with specific mutations to worsen it.The periodontal tissue is an area that niches the SARS-CoV-2 virus due to an unconditionally favourable environment, with changes to the spike protein.The emergence of variants has raised concerns regarding their biological interactions and implicationsfor oral health[170]. The hypothesis is that variants with better invasive capabilities and changes to their genome can overexpress themselves in the oral region, thereby migrating to the bloodstream and organs and may cause a higher rate of infection to cells such as T-lymphocytes, B-lymphocytes, macrophages, etc. as new variants possess the ability to evade the immune system[38]. The potential arrival of new

SARS-CoV-2 variants raises many questions regarding their pattern of infection and severity. With only the available previous data, we can only contemplate the actions of the variants prepared for them; the need to investigate each strain in-depth and their association with various receptors, body organs, and functional pathways is required.

# 7. Phytochemicals against Coronaviruses

After the outbreak of SARS-CoV-2, many researchers primarily focused on using natural entities to treat the complication associated with COVID-19[171]. Thus, the primary in-vitro and in-vivo studies were targeted to screen different phytochemicals effective against coronaviruses (especially SARS-CoV-2). Bio-informaticians were performing computer docking using other models to predict the anti-CoVs of the various phytochemicals against the members of the coronavirus family like MERS-CoV, SARS-CoV and SARS-CoV-2[172, 173]. This investigation unveiled that natural polyphenol compounds like apigenin,kaempferol, myricetin, quercetin and resveratrol have significant activities against different coronaviruses[171].Cho and his teammates isolated geranylated flavonoids (tomentin A-E) from Paulownia tomentosa (Thunb.) Steud. (Paulowniaceae), which inhibit the activity of papain-like protease involved in SARS-CoV propagation[174]. Another study reported three flavonoids, *i.e.*, apigenin-7-O-rhamnoglucoside, pectolinarin and herbacetin, which at the concentration of 20 µM ceases the activity of 3C-like protease, a crucial enzyme involved in SARS-CoVreplication[175]. Additionally, the ten polyphenols isolated from Broussonetiapapyrifera (L.) L'Hér.ex Vent. (Moraceae), also reported the inhibition of 3C-like protease, exclusively with papyriflavonol A at 3.7 µM[176].On the other hand, the molecular docking assessment was done on the theaflavin, a flavonoid compound isolated from black tea, Camellia sinensis (L.) Kuntze (Theaceae) (Traditional Chinese Medicinal Plant) inhibits the normal functioning of SARS-CoV-2 RNA-dependent RNA polymerase and exhibits the antiviral potential of these compounds, especially SARS-CoV-2[177]. Furthermore, the compound hesperidin, predominantly found in citrus fruits, has also been proposed to show an inhibitory effect on ACE2; owing to this, it has been proposed as a good candidate against SARS-CoV-2 for clinical trials[178]. Interestingly, Alam and his colleagues reviewed the use of an oral spray ArtemiC (containing a mixture of artemisinin, Boswelliaserrata, curcumin and vitamin C) in 50 covid patients. The oral administration of ArtemiC twice a day for two consecutive days during treatment showed negative COVID-19 PCR after 14 days[179].

# 8. Conclusion and Future Prospects

The importance of oralhygiene in the current COVID-19 pandemic is very evident. It is assumed that virus transmission can occur through the oral cavity to the lungs, but it elucidates the disease's clinical variability and radiological aspects. Viral transmission can also occur through the oral mucosa and nasal route in healthy individuals, whereby those with poor oral health impose themselves infected by SARS-CoV-2. On the other side, individuals with a weakened immune system are more likely to develop severe symptomsrequiring intensive care with mechanical ventilation, which might lead to death in many cases.

Moreover, new variants of COVID-19 are imposing health challenges and risks. Thus, considering the risk, following proper oral hygiene and other measures for controlling plaque needs to be prioritized. These precautions will improve oral health and well-being and reduce the risk of developing COVID-19. The pandemic has acted as an opportunity for the oral health plan to be renewed. The health care policymakers should promote oral health by giving training programs to the health workers and educating the local communities about oral health care. Through this practice, the health workers and locals can be prepared for the causalities like the pandemic of COVID-19 beforehand.

Considering the Indian scenario, where most of the population resides in the villages, the concept of oral health is almost negligible. As a result, toothpaste and mouthwashes are not readily available or, in some places, unaffordable. Insuch cases, awareness of simple measures, such as gargling with saltwater and rinsingwith boiled water, helps reduce gingival inflammation. Following these simple measures will aid in reducing the viral load in saliva. Thus, healthcare industries need to take the initiative to spread awareness and incorporate effective phyto compounds in their products, making them affordable for people worldwide. This step will substantially help curb the progression of lung diseases and mitigate the chances of severe COVID-19, ultimately reducing the mortality rate. The health care workers should be trained; in their training programs, interdepartmental professionals should be involved like the dentists, clinicians, and researchers, where the ground information about oral health care can be discussed, and effects parameters can be designed which can work in Indian picture at the ground level in villages and towns. By following this cumulative approach, a bridge between the departments can be made, which will benefit swaying the oral health statistics in India.

#### **Declaration of competing interest**

The authors declared that there was no conflict of interest.Consent for publication

# **Competing interests**

The authors declare that they have no competing interests.

# References

- [1] Burton-Jeangros, C., et al., A life course perspective on health trajectories and transitions. 2015.
- [2] Marcenes, W., et al., Global burden of oral conditions in 1990-2010: a systematic analysis. Journal of dental research, 2013.
   92(7): p. 592-597.
- [3] Abbasi-Shavazi, M., et al., Predictors of oral health-related quality of life in 2-5 year-old children in the South of Iran. Health Qual Life Outcomes, 2020. 18(1): p. 384.
- [4] Zhang, J., et al., Risk predictors of dental root caries: A systematic review. J Dent, 2019. 89: p. 103166.
- [5] Sheiham, A. and R.G. Watt, The common risk factor approach: a rational basis for promoting oral health. Community Dentistry and Oral Epidemiology: Commentary, 2000. 28(6): p. 399-406.
- [6] Watt, R.G. and A. Sheiham, Integrating the common risk factor approach into a social determinants framework. Community dentistry and oral epidemiology, 2012. 40(4): p. 289-296.
- [7] Dalir Abdolahinia, E., et al., Potential applications of medicinal herbs and phytochemicals in oral and dental health: Status quo and future perspectives. Oral Dis, 2022.

- [8] Kumar, G., et al., Emerging trends of herbal care in dentistry. J Clin Diagn Res, 2013. 7(8): p. 1827-9.
- [9] Sinha, D.J. and A.A. Sinha, Natural medicaments in dentistry. Ayu, 2014. 35(2): p. 113-8.
- [10] Uju, D.E. and N.P. Obioma, Anticariogenic potentials of clove, tobacco and bitter kola. Asian Pac J Trop Med, 2011. 4(10): p. 814-8.
- [11] Lloyd-Jones, G., et al., The COVID-19 pathway: a proposed oral-vascular-pulmonary route of SARS-CoV-2 infection and the importance of oral healthcare measures. J Oral Med Dent Res, 2021. 2(1): p. 1-25.
- [12] Murgolo, N., et al., SARS-CoV-2 tropism, entry, replication, and propagation: Considerations for drug discovery and development. PLoS pathogens, 2021. 17(2): p. e1009225.
- [13] Huang, N., et al., Integrated single-cell atlases reveal an oral SARS-CoV-2 infection and transmission axis. MedRxiv, 2020.
- [14] Sakaguchi, W., et al., Existence of SARS-CoV-2 entry molecules in the oral cavity. International journal of molecular sciences, 2020. 21(17): p. 6000.
- [15] Badran, Z., et al., Periodontal pockets: A potential reservoir for SARS-CoV-2? Medical Hypotheses, 2020. 143: p. 109907.
- [16] Fernandes Matuck, B., et al., Periodontal tissues are targets for Sars-Cov-2: a post-mortem study. Journal of oral microbiology, 2021. 13(1): p. 1848135.
- [17] Huang, N., et al., SARS-CoV-2 infection of the oral cavity and saliva. Nature medicine, 2021. 27(5): p. 892-903.
- [18] Silva, J., et al., Saliva viral load is a dynamic unifying correlate of COVID-19 severity and mortality. MedRxiv, 2021.
- [19] Control, C.f.D. and Prevention, Covid-19 risks and vaccine information for older adults. Retrieved on Sept, 2021. 9: p. 2021.
- [20] Gherlone, E., et al., Dentistry and Covid-19 pandemic: operative indications post-lockdown. New Microbiol, 2021. 44(1): p. 1-11.
- [21] Feher, B., et al., The Effect of the COVID-19 Pandemic on Patient Selection, Surgical Procedures, and Postoperative Complications in a Specialized Dental Implant Clinic. J Clin Med, 2022. 11(3).
- [22] Cappare, P., et al., The Usage of an Air Purifier Device with HEPA 14 Filter during Dental Procedures in COVID-19 Pandemic: A Randomized Clinical Trial. Int J Environ Res Public Health, 2022. 19(9).
- [23] Hong, D.G.K. and J.H. Oh, Recent advances in dental implants. Maxillofac Plast Reconstr Surg, 2017. 39(1): p. 33.
- [24] Cagidiaco, E.F., et al., Functional Implant Prosthodontic Score of a one-year prospective study on three different connections for single-implant restorations. Journal of Osseointegration, 2018. 10(4): p. 130-135.
- [25] Tecco, S., et al., The association between three attitude-related indexes of oral hygiene and secondary implant failures: A retrospective longitudinal study. Int J Dent Hyg, 2018. 16(3): p. 372-379.
- [26] Block, M.S., Coronavirus Disease 2019 May Affect Dental Implant Integration. J Oral Maxillofac Surg, 2021. 79(6): p. 1197-1198.
- [27] D'Orto, B., et al., Full Arch Implant-Prosthetic Rehabilitation in Patients with Type I Diabetes Mellitus: Retrospective Clinical Study with 10 Year Follow-Up. Int J Environ Res Public Health, 2022. 19(18).
- [28] Dubey, R.K., D.K. Gupta, and A.K. Singh, Dental implant survival in diabetic patients; review and recommendations. Natl J Maxillofac Surg, 2013. 4(2): p. 142-50.
- [29] Gherlone, E.F., et al., Evaluation of resistance against bacterial microleakage of a new conical implant-abutment connection versus conventional connections: an in vitro study. New Microbiol, 2016. 39(1): p. 49-56.
- [30] Pallos, D., et al., Periodontal disease and detection of human herpesviruses in saliva and gingival crevicular fluid of chronic kidney disease patients. Journal of Periodontology, 2020. 91(9): p. 1139-1147.
- [31] Contreras, A., H. Nowzari, and J. Slots, Herpesviruses in periodontal pocket and gingival tissue specimens. Oral microbiology and immunology, 2000. 15(1): p. 15-18.
- [32] Musso, D., et al., Detection of Zika virus in saliva. Journal of Clinical Virology, 2015. 68: p. 53-55.
- [33] Matičić, M., et al., Proviral HIV-1 DNA in gingival crevicular fluid of HIV-1-infected patients in various stages of HIV disease. Journal of dental research, 2000. 79(7): p. 1496-1501.
- [34] Parra, B. and J. Slots, Detection of human viruses in periodontal pockets using polymerase chain reaction. Oral microbiology and immunology, 1996. 11(5): p. 289-293.
- [35] Gupta, S., et al., SARS-CoV-2 detection in gingival crevicular fluid. Journal of dental research, 2021. 100(2): p. 187-193.

- [36] Hujoel, P., et al., The dentogingival epithelial surface area revisited. Journal of periodontal research, 2001. 36(1): p. 48-55.
- [37] Wright, H.J., et al., Periodontitis associates with a type 1 IFN signature in peripheral blood neutrophils. The Journal of Immunology, 2008. 181(8): p. 5775-5784.
- [38] Ito, H.-O., Infective endocarditis and dental procedures: evidence, pathogenesis, and prevention. The Journal of Medical Investigation, 2006. 53(3, 4): p. 189-198.
- [39] Chen, X., et al., Microbial etiology and prevention of dental caries: exploiting natural products to inhibit cariogenic biofilms. Pathogens, 2020. 9(7): p. 569.
- [40] Pitts, N.B., et al., Dental caries. Nature reviews Disease primers, 2017. 3(1): p. 1-16.
- [41] Lu, M., S. Xuan, and Z. Wang, Oral microbiota: A new view of body health. Food Science and Human Wellness, 2019. 8(1): p. 8-15.
- [42] Razi, M.A., et al., Role of natural salivary defenses in the maintenance of healthy oral microbiota in children and adolescents. Journal of Family Medicine and Primary Care, 2020. 9(3): p. 1603.
- [43] Marsh, P. and E. Zaura, Dental biofilm: ecological interactions in health and disease. Journal of clinical periodontology, 2017.
   44: p. S12-S22.
- [44] 44. Sim, C.P., S.G. Dashper, and E.C. Reynolds, Oral microbial biofilm models and their application to the testing of anticariogenic agents. Journal of Dentistry, 2016. 50: p. 1-11.
- [45] Yadav, K. and S. Prakash, Dental caries: A microbiological approach. J Clin Infect Dis Pract, 2017. 2(1): p. 1-15.
- [46] Van der Mei, H., et al., Effects of amine fluoride on biofilm growth and salivary pellicles. Caries research, 2008. 42(1): p. 19-27.
- [47] Lamont, R.J., H. Koo, and G. Hajishengallis, The oral microbiota: dynamic communities and host interactions. Nature reviews microbiology, 2018. 16(12): p. 745-759.
- [48] Seminario, A., Z. Broukal, and R. Ivancakova, Mutans streptococci and the development of dental plaque. Prague medical report, 2005. 106(4): p. 349-358.
- [49] Widyarman, A.S. and C.F. Theodorea, Effect of reuterin on dual-species biofilm in vitro of Streptococcus mutans and Veillonella parvula. Journal of International Dental and Medical Research, 2019. 12(1): p. 77-83.
- [50] Fakhruddin, K.S., H.C. Ngo, and L.P. Samaranayake, Cariogenic microbiome and microbiota of the early primary dentition: A contemporary overview. Oral diseases, 2019. 25(4): p. 982-995.
- [51] Liu, J.-f., C.-L. Hsu, and L.-R. Chen, Correlation between salivary mutans streptococci, lactobacilli and the severity of early childhood caries. Journal of dental sciences, 2019. 14(4): p. 389-394.
- [52] Obata, J., et al., Pathogenic mechanisms of cariogenic Propionibacterium acidifaciens. Archives of Oral Biology, 2019. 105: p. 46-51.
- [53] Bowen, W.H., et al., Oral biofilms: pathogens, matrix, and polymicrobial interactions in microenvironments. Trends in microbiology, 2018. 26(3): p. 229-242.
- [54] Amaechi, B.T., et al., Protocols to study dental caries in vitro: Microbial caries models, in Odontogenesis. 2019, Springer. p. 357-368.
- [55] Ccahuana-Vásquez, R.A. and J.A. Cury, S. mutans biofilm model to evaluate antimicrobial substances and enamel demineralization. Brazilian oral research, 2010. 24(2): p. 135-141.
- [56] Alshahrani, A.M. and R.L. Gregory, In vitro Cariostatic effects of cinnamon water extract on nicotine-induced Streptococcus mutans biofilm. BMC complementary medicine and therapies, 2020. 20(1): p. 1-9.
- [57] Zhang, Q., et al., Inhibitory and preventive effects of Lactobacillus plantarum FB-T9 on dental caries in rats. Journal of Oral Microbiology, 2020. 12(1): p. 1703883.
- [58] Palmer, S.R., et al., Streptococcus mutans yidC1 and yidC2 impact cell envelope biogenesis, the biofilm matrix, and biofilm biophysical properties. Journal of bacteriology, 2019. 201(1): p. e00396-18.
- [59] Zhang, Q., et al., Structure-based discovery of small molecule inhibitors of cariogenic virulence. Scientific reports, 2017. 7(1):
   p. 1-10.

- [60] Elgamily, H., R. Safy, and R. Makharita, Influence of medicinal plant extracts on the growth of oral pathogens Streptococcus mutans and Lactobacillus acidophilus: an in-vitro study. Open access Macedonian journal of medical sciences, 2019. 7(14): p. 2328.
- [61] Philip, N., et al., Polyphenol-rich cranberry extracts modulate virulence of Streptococcus mutans-Candida albicans biofilms implicated in the pathogenesis of early childhood caries. Pediatric dentistry, 2019. 41(1): p. 56-62.
- [62] Manome, A., et al., Acidogenic potential of oral bifidobacterium and its high fluoride tolerance. Frontiers in Microbiology, 2019. 10: p. 1099.
- [63] do Rosário Palma, A.L., et al., Influence of Streptococcus mitis and Streptococcus sanguinis on virulence of Candida albicans: in vitro and in vivo studies. Folia microbiologica, 2019. 64(2): p. 215-222.
- [64] Mira, A., et al., Development of an in vitro system to study oral biofilms in real time through impedance technology: validation and potential applications. Journal of oral microbiology, 2019. 11(1): p. 1609838.
- [65] Shu, M., et al., Development of multi-species consortia biofilms of oral bacteria as an enamel and root caries model system. Archives of oral biology, 2000. 45(1): p. 27-40.
- [66] Balhaddad, A.A., et al., Toward dental caries: Exploring nanoparticle-based platforms and calcium phosphate compounds for dental restorative materials. Bioactive materials, 2019. 4: p. 43-55.
- [67] Popova, C., V. Dosseva-Panova, and V. Panov, Microbiology of periodontal diseases. A review. Biotechnology & Biotechnological Equipment, 2013. 27(3): p. 3754-3759.
- [68] Newman, M. and S. Socransky, Predominant cultivable microbiota in periodontosis. Journal of periodontal research, 1977. 12(2): p. 120-128.
- [69] Slots, J., Subgingival microflora and periodontal disease. Journal of clinical periodontology, 1979. 6(5): p. 351-382.
- [70] Choo, A., D.M. Delac, and L.B. Messer, Oral hygiene measures and promotion: review and considerations. Australian dental journal, 2001. 46(3): p. 166-173.
- [71] Claydon, N.C., Current concepts in toothbrushing and interdental cleaning. Periodontology 2000, 2008. 48(1): p. 10-22.
- [72] Shah, N., et al., Association between traditional oral hygiene methods with tooth wear, gingival bleeding, and recession: A descriptive cross-sectional study. Indian Journal of Dental Research, 2018. 29(2): p. 150.
- [73] Swati. Top 10 Best Toothpaste Brands in India. 2022 [cited 2022 June 6]; Available from: https://www.grabon.in/indulge/health/best-toothpaste-brands-india/.
- [74] Kumar, M., et al., Beneficial Role of Antioxidant Secondary Metabolites from Medicinal Plants in Maintaining Oral Health. Antioxidants (Basel), 2021. 10(7).
- [75] Nordin, A., et al., Miswak and oral health: An evidence-based review. Saudi J Biol Sci, 2020. 27(7): p. 1801-1810.
- [76] Malik, A.S., et al., Comparative effectiveness of chewing stick and toothbrush: a randomized clinical trial. N Am J Med Sci, 2014. 6(7): p. 333-7.
- [77] Elavarasu, S., et al., Evaluation of anti-plaque microbial activity of Azadirachta indica (neem oil) in vitro: A pilot study. J Pharm Bioallied Sci, 2012. 4(Suppl 2): p. S394-6.
- [78] Naik, D., et al., Commercially Available Toothpaste in India. Indian Journal of Forensic Medicine & Toxicology, 2020. 14(4).
- [79] Axelsson, P., B. Nyström, and J. Lindhe, The long-term effect of a plaque control program on tooth mortality, caries and periodontal disease in adults: results after 30 years of maintenance. Journal of clinical periodontology, 2004. 31(9): p. 749-757.
- [80] Netuschil, L., et al., Plaque bacteria counts and vitality during chlorhexidine, meridol and listerine mouthrinses. European Journal of Oral Sciences, 1995. 103(6): p. 355-361.
- [81] Bassolé, I.H.N. and H.R. Juliani, Essential oils in combination and their antimicrobial properties. Molecules, 2012. 17(4): p. 3989-4006.
- [82] Galvão, L.C.d.C., et al., Antimicrobial activity of essential oils against Streptococcus mutans and their antiproliferative effects. Evidence-Based Complementary and Alternative Medicine, 2012. 2012.
- [83] Bakkali, F., et al., Biological effects of essential oils-a review. Food and chemical toxicology, 2008. 46(2): p. 446-475.
- [84] de Cássia da Silveira e Sá, R., L.N. Andrade, and D.P. de Sousa, A review on anti-inflammatory activity of monoterpenes. Molecules, 2013. 18(1): p. 1227-1254.

- [85] Pichersky, E., J.P. Noel, and N. Dudareva, Biosynthesis of plant volatiles: nature's diversity and ingenuity. Science, 2006. 311(5762): p. 808-811.
- [86] Bernardes, W.A., et al., Antibacterial activity of the essential oil from Rosmarinus offi cinalis and its major components against oral pathogens. Zeitschrift f
  ür Naturforschung C, 2010. 65(9-10): p. 588-593.
- [87] Botelho, M.A., et al., Efficacy of a mouthrinse based on leaves of the neem tree (Azadirachta indica) in the treatment of patients with chronic gingivitis: A double-blind, randomized, controlled trial. J Med Plants Res, 2008. 2(11): p. 341-346.
- [88] Preus, H.R., et al., The plaque-and gingivitis-inhibiting capacity of a commercially available essential oil product. A parallel, split-mouth, single blind, randomized, placebo-controlled clinical study. Acta Odontologica Scandinavica, 2013. 71(6): p. 1613-1619.
- [89] Van Leeuwen, M., D. Slot, and G. Van der Weijden, The effect of an essential-oils mouthrinse as compared to a vehicle solution on plaque and gingival inflammation: a systematic review and meta-analysis. International Journal of Dental Hygiene, 2014. 12(3): p. 160-167.
- [90] Eduardo, F.P., et al., Salivary SARS-CoV-2 load reduction with mouthwash use: A randomized pilot clinical trial. Heliyon, 2021. 7(6): p. e07346.
- [91] Khalil, A.A., et al., Essential oil eugenol: sources, extraction techniques and nutraceutical perspectives. RSC advances, 2017. 7(52): p. 32669-32681.
- [92] Hemaiswarya, S. and M. Doble, Synergistic interaction of eugenol with antibiotics against Gram negative bacteria. Phytomedicine, 2009. 16(11): p. 997-1005.
- [93] Oyedemi, S., et al., The proposed mechanism of bactericidal action of eugenol, ∝-terpineol and g-terpinene against Listeria monocytogenes, Streptococcus pyogenes, Proteus vulgaris and Escherichia coli. African Journal of Biotechnology, 2009. 8(7).
- [94] Escobar, A., et al., Thymol bioactivity: A review focusing on practical applications. Arabian Journal of Chemistry, 2020. 13(12): p. 9243-9269.
- [95] Milovanovic, S., et al., Supercritical CO2-assisted production of PLA and PLGA foams for controlled thymol release. Materials Science and Engineering: C, 2019. 99: p. 394-404.
- [96] Nagoor Meeran, M.F., et al., Pharmacological properties and molecular mechanisms of thymol: prospects for its therapeutic potential and pharmaceutical development. Frontiers in pharmacology, 2017. 8: p. 380.
- [97] Ebrahimzadeh, H., et al., Chemical composition of the essential oil and supercritical CO2 extracts of Zataria multiflora Boiss. Food chemistry, 2003. 83(3): p. 357-361.
- [98] De Feo, V., et al., Chemical composition and antibacterial activity of essential oils from Thymus spinulosus Ten.(Lamiaceae). Journal of agricultural and food chemistry, 2003. 51(13): p. 3849-3853.
- [99] Nabavi, S.M., et al., Plants belonging to the genus Thymus as antibacterial agents: From farm to pharmacy. Food chemistry, 2015. 173: p. 339-347.
- [100] Tohidpour, A., et al., Antibacterial effect of essential oils from two medicinal plants against Methicillin-resistant Staphylococcus aureus (MRSA). Phytomedicine, 2010. 17(2): p. 142-145.
- [101] Kowalczyk, A., et al., Thymol and thyme essential oil—new insights into selected therapeutic applications. Molecules, 2020. 25(18): p. 4125.
- [102] Elaissi, A., et al., Variation in volatile leaf oils of twelve Eucalyptus species harvested from Hajeb Layoun Arboreta (Tunisia). Chemistry & biodiversity, 2010. 7(3): p. 705-716.
- [103] Souza, T.d.A.d., et al., Alpinia essential oils and their major components against Rhodnius nasutus, a vector of chagas disease. The Scientific World Journal, 2018. 2018.
- [104] Nogueira, M., et al., Terpinen-4-ol and alpha-terpineol (tea tree oil components) inhibit the production of IL-1β, IL-6 and IL-10 on human macrophages. Inflammation research, 2014. 63(9): p. 769-778.
- [105] Thomsen, N.A., et al., Effect of habituation to tea tree (Melaleuca alternifolia) oil on the subsequent susceptibility of Staphylococcus spp. to antimicrobials, triclosan, tea tree oil, terpinen-4-ol and carvacrol. International journal of antimicrobial agents, 2013. 41(4): p. 343-351.

- [106] Carson, C.F., B.J. Mee, and T.V. Riley, Mechanism of action of Melaleuca alternifolia (tea tree) oil on Staphylococcus aureus determined by time-kill, lysis, leakage, and salt tolerance assays and electron microscopy. Antimicrobial agents and chemotherapy, 2002. 46(6): p. 1914-1920.
- [107] Li, W.-R., et al., The dynamics and mechanism of the antimicrobial activity of tea tree oil against bacteria and fungi. Applied microbiology and biotechnology, 2016. 100(20): p. 8865-8875.
- [108] Straede, A., et al., The effect of tea tree oil and antifungal agents on a reporter for yeast cell integrity signalling. Yeast, 2007. 24(4): p. 321-334.
- [109] Saxer, U.P., et al., Effect of mouthwashing with tea tree oil on plaque and inflammation. Schweizer Monatsschrift fur Zahnmedizin= Revue Mensuelle Suisse D'odonto-stomatologie= Rivista Mensile Svizzera di Odontologia e Stomatologia, 2003. 113(9): p. 985-996.
- [110] Maggi, F., et al., Composition and biological activity of essential oil of Achillea ligustica All.(Asteraceae) naturalized in central Italy: Ideal candidate for anti-cariogenic formulations. Fitoterapia, 2009. 80(6): p. 313-319.
- [111] Rahman, B., et al., Comparative antiplaque and antigingivitis effectiveness of tea tree oil mouthwash and a cetylpyridinium chloride mouthwash: A randomized controlled crossover study. Contemporary clinical dentistry, 2014. 5(4): p. 466.
- [112] Varma, S.R., et al., The antiplaque efficacy of two herbal-based toothpastes: A clinical intervention. Journal of International Society of Preventive & Community Dentistry, 2018. 8(1): p. 21.
- [113] Ali, B.H., et al., Some biological properties of curcumin: A review. Natural Product Communications, 2006. 1(6): p. 1934578X0600100613.
- [114] Eigner, D. and D. Scholz, Ferula asa-foetida and Curcuma longa in traditional medical treatment and diet in Nepal. Journal of ethnopharmacology, 1999. 67(1): p. 1-6.
- [115] Grant, K.L. and C.D. Schneider, Alternative therapies. American Journal of Health-System Pharmacy, 2000. 57(12): p. 1121-1122.
- [116] Tyagi, P., et al., Bactericidal activity of curcumin I is associated with damaging of bacterial membrane. PloS one, 2015. 10(3): p. e0121313.
- [117] Kaur, S., et al., Probing the binding site of curcumin in Escherichia coli and Bacillus subtilis FtsZ–a structural insight to unveil antibacterial activity of curcumin. European journal of medicinal chemistry, 2010. 45(9): p. 4209-4214.
- [118] Teow, S.-Y., et al., Antibacterial action of curcumin against Staphylococcus aureus: a brief review. Journal of tropical medicine, 2016. 2016.
- [119] Yun, D.G. and D.G. Lee, Antibacterial activity of curcumin via apoptosis-like response in Escherichia coli. Applied microbiology and biotechnology, 2016. 100(12): p. 5505-5514.
- [120] Sharma, M., et al., Antifungal curcumin induces reactive oxygen species and triggers an early apoptosis but prevents hyphae development by targeting the global repressor TUP1 in Candida albicans. Bioscience reports, 2010. 30(6): p. 391-404.
- [121] Neelofar, K., et al., Curcumin as a promising anticandidal of clinical interest. Canadian Journal of Microbiology, 2011.57(3): p. 204-210.
- [122] Zhang, Y., et al., Antibacterial and antibiofilm activities of eugenol from essential oil of Syzygium aromaticum (L.) Merr.
   & LM Perry (clove) leaf against periodontal pathogen Porphyromonas gingivalis. Microbial pathogenesis, 2017. 113: p. 396-402.
- [123] Lopez, M., M. Hadisurya, and R. Cornwall, Antimicrobial Investigation and Structure activity analysis of natural eugenol derivatives against several oral bacteria. J. Pharm. Biol, 2019. 5: p. 1.
- [124] Adil, M., et al., Eugenol-induced suppression of biofilm-forming genes in Streptococcus mutans: An approach to inhibit biofilms. Journal of global antimicrobial resistance, 2014. 2(4): p. 286-292.
- [125] Miladi, H., et al., Synergistic effect of eugenol, carvacrol, thymol, p-cymene and  $\gamma$ -terpinene on inhibition of drug resistance and biofilm formation of oral bacteria. Microbial pathogenesis, 2017. 112: p. 156-163.

- [126] de Paula, S.B., et al., Effect of eugenol on cell surface hydrophobicity, adhesion, and biofilm of Candida tropicalis and Candida dubliniensis isolated from oral cavity of HIV-infected patients. Evidence-Based Complementary and Alternative Medicine, 2014. 2014.
- [127] Wang, T.-H., et al., Evaluation of the antibacterial potential of liquid and vapor phase phenolic essential oil compounds against oral microorganisms. PLoS One, 2016. 11(9): p. e0163147.
- [128] Botelho, M., et al., Antimicrobial activity of the essential oil from Lippia sidoides, carvacrol and thymol against oral pathogens. Brazilian Journal of Medical and Biological Research, 2007. 40: p. 349-356.
- [129] De Castro, R.D., et al., Antifungal activity and mode of action of thymol and its synergism with nystatin against Candida species involved with infections in the oral cavity: an in vitro study. BMC complementary and alternative medicine, 2015. 15(1): p. 1-7.
- [130] Bordini, E.A.F., et al., Antimicrobial effects of terpinen-4-ol against oral pathogens and its capacity for the modulation of gene expression. Biofouling, 2018. 34(7): p. 815-825.
- [131] Bucci, A.R., et al., The antimicrobial and antiadhesion activities of micellar solutions of surfactin, CTAB and CPCl with terpinen-4-ol: applications to control oral pathogens. World Journal of Microbiology and Biotechnology, 2018. 34(6): p. 1-9.
- [132] Ramage, G., et al., Antifungal, cytotoxic, and immunomodulatory properties of tea tree oil and its derivative components: potential role in management of oral candidosis in cancer patients. Frontiers in microbiology, 2012. 3: p. 220.
- [133] Izui, S., et al., Antibacterial activity of curcumin against periodontopathic bacteria. Journal of periodontology, 2016.87(1): p. 83-90.
- [134] Mandroli, P.S. and K. Bhat, An in-vitro evaluation of antibacterial activity of curcumin against common endodontic bacteria. Journal of Applied Pharmaceutical Science, 2013. 3(10): p. 106-108.
- [135] Song, J., et al., Curcumin suppresses Streptococcus mutans adherence to human tooth surfaces and extracellular matrix proteins. European journal of clinical microbiology & infectious diseases, 2012. 31(7): p. 1347-1352.
- [136] Raja, M.C., et al., Versatile and synergistic potential of eugenol: a review. Pharm. Anal. Acta, 2015. 6(5): p. 1-6.
- [137] da Silva, J.K.R., et al., Essential oils as antiviral agents, potential of essential oils to treat SARS-CoV-2 infection: An in-silico investigation. International journal of molecular sciences, 2020. 21(10): p. 3426.
- [138] Zhang, Y. and T.G. Kutateladze, Molecular structure analyses suggest strategies to therapeutically target SARS-CoV-2. Nature communications, 2020. 11(1): p. 1-4.
- [139] Walls, A.C., et al., Structure, function, and antigenicity of the SARS-CoV-2 spike glycoprotein. Cell, 2020. 181(2): p. 281-292. e6.
- [140] Kulkarni, S.A., et al., Computational evaluation of major components from plant essential oils as potent inhibitors of SARS-CoV-2 spike protein. Journal of Molecular Structure, 2020. 1221: p. 128823.
- [141] Rolta, R., et al., Phytocompounds of Rheum emodi, Thymus serpyllum, and Artemisia annua inhibit spike protein of SARS-CoV-2 binding to ACE2 receptor: in silico approach. Current pharmacology reports, 2021. 7(4): p. 135-149.
- [142] Gonzalez-Paz, L.A., et al., Theoretical molecular docking study of the structural disruption of the viral 3CL-protease of COVID19 induced by binding of capsaicin, piperine and curcumin part 1: A comparative study with chloroquine and hydrochloroquine two antimalaric drugs. 2020.
- [143] Zorofchian Moghadamtousi, S., et al., A review on antibacterial, antiviral, and antifungal activity of curcumin. BioMed research international, 2014. 2014.
- [144] Dairaku, I., et al., Inhibitory effect of curcumin on IMP dehydrogenase, the target for anticancer and antiviral chemotherapy agents. Bioscience, biotechnology, and biochemistry, 2010: p. 0912011751-0912011751.
- [145] Nabel, G.J., et al., Alternative mechanisms for activation of human immunodeficiency virus enhancer in T cells. Science, 1988. 239(4845): p. 1299-1302.
- [146] Cullen, B.R. and W.C. Greene, Regulatory pathways governing HIV-1 replication. Cell, 1989. 58(3): p. 423-426.
- [147] Li, C.J., et al., Three inhibitors of type 1 human immunodeficiency virus long terminal repeat-directed gene expression and virus replication. Proceedings of the National Academy of Sciences, 1993. 90(5): p. 1839-1842.

- [148] Barthelemy, S., et al., Curcumin and curcumin derivatives inhibit Tat-mediated transactivation of type 1 human immunodeficiency virus long terminal repeat. Research in virology, 1998. 149(1): p. 43-52.
- [149] Chen, D.-Y., et al., Curcumin inhibits influenza virus infection and haemagglutination activity. Food Chemistry, 2010. 119(4): p. 1346-1351.
- [150] Sharifi-Rad, J., et al., Susceptibility of herpes simplex virus type 1 to monoterpenes thymol, carvacrol, p-cymene and essential oils of Sinapis arvensis L., Lallemantia royleana Benth. and Pulicaria vulgaris Gaertn. Cellular and Molecular Biology, 2017. 63(8): p. 42-47.
- [151] Maurya, V.K., et al., Structure-based drug designing for potential antiviral activity of selected natural products from Ayurveda against SARS-CoV-2 spike glycoprotein and its cellular receptor. Virusdisease, 2020. 31(2): p. 179-193.
- [152] Astani, A., J. Reichling, and P. Schnitzler, Screening for antiviral activities of isolated compounds from essential oils. Evidence-based complementary and alternative medicine, 2011. 2011.
- [153] Vimalanathan, S. and J. Hudson, Anti-influenza virus activity of essential oils and vapors. American Journal of Essential Oils and Natural Products, 2014. 2(1): p. 47-53.
- [154] Wani, A.R., et al., An updated and comprehensive review of the antiviral potential of essential oils and their chemical constituents with special focus on their mechanism of action against various influenza and coronaviruses. Microbial Pathogenesis, 2021. 152: p. 104620.
- [155] Kubiça, T.F., et al., In vitro inhibition of the bovine viral diarrhoea virus by the essential oil of Ocimum basilicum (basil) and monoterpenes. Brazilian Journal of Microbiology, 2014. 45: p. 209-214.
- [156] Garozzo, A., et al., In vitro antiviral activity of Melaleuca alternifolia essential oil. Letters in applied microbiology, 2009.
   49(6): p. 806-808.
- [157] Mounce, B.C., et al., Curcumin inhibits Zika and chikungunya virus infection by inhibiting cell binding. Antiviral research, 2017. 142: p. 148-157.
- [158] Jhun, H., et al., SARS-CoV-2 Delta (B. 1.617. 2) variant: a unique T478K mutation in receptor binding motif (RBM) of spike gene. Immune Network, 2021. 21(5).
- [159] Tracking SARS-CoV-2 variants. 2022 [cited 2022 May,16]; Available from: https://www.who.int/en/activities/tracking-SARS-CoV-2-variants/.
- [160] Bakhshandeh, B., et al., Mutations in SARS-CoV-2; Consequences in structure, function, and pathogenicity of the virus. Microbial Pathogenesis, 2021. 154: p. 104831.
- [161] Ratre, Y.K., et al., Molecular mechanism, diagnosis, and potential treatment for novel coronavirus (COVID-19): a current literature review and perspective. 3 Biotech, 2021. 11(2): p. 1-24.
- [162] Huang, S.-W. and S.-F. Wang, SARS-CoV-2 entry related viral and host genetic variations: Implications on COVID-19 severity, immune escape, and infectivity. International Journal of Molecular Sciences, 2021. 22(6): p. 3060.
- [163] Lu, R., et al., Genomic characterisation and epidemiology of 2019 novel coronavirus: implications for virus origins and receptor binding. The lancet, 2020. 395(10224): p. 565-574.
- Bian, L., et al., Impact of the Delta variant on vaccine efficacy and response strategies. Expert review of vaccines, 2021.
   20(10): p. 1201-1209.
- [165] Alam, I., et al., CovMT: an interactive SARS-CoV-2 mutation tracker, with a focus on critical variants. The Lancet Infectious Diseases, 2021. 21(5): p. 602.
- [166] Choudhary, S., et al., Role of genetic variants and gene expression in the susceptibility and severity of COVID-19. Annals of laboratory medicine, 2021. 41(2): p. 129-138.
- [167] Salleh, M.Z., J.P. Derrick, and Z.Z. Deris, Structural evaluation of the spike glycoprotein variants on SARS-CoV-2 transmission and immune evasion. International Journal of Molecular Sciences, 2021. 22(14): p. 7425.
- [168] Moghaddar, M., R. Radman, and I. Macreadie, Severity, pathogenicity and transmissibility of delta and lambda variants of SARS-CoV-2, toxicity of spike protein and possibilities for future prevention of COVID-19. Microorganisms, 2021. 9(10): p. 2167.

- [169] Baral, P., et al., Mutation-induced changes in the receptor-binding interface of the SARS-CoV-2 Delta variant B. 1.617. 2 and implications for immune evasion. Biochemical and biophysical research communications, 2021. 574: p. 14-19.
- [170] Coke, C.J., et al., SARS-CoV-2 infection and oral health: Therapeutic opportunities and challenges. Journal of Clinical Medicine, 2021. 10(1): p. 156.
- [171] Majnooni, M.B., et al., Phytochemicals: Potential Therapeutic Interventions Against Coronavirus-Associated Lung Injury. Front Pharmacol, 2020. 11: p. 588467.
- [172] Mani, J.S., et al., Natural product-derived phytochemicals as potential agents against coronaviruses: A review. Virus Res, 2020. 284: p. 197989.
- [173] Zhang, D.H., et al., In silico screening of Chinese herbal medicines with the potential to directly inhibit 2019 novel coronavirus. J Integr Med, 2020. 18(2): p. 152-158.
- [174] Cho, J.K., et al., Geranylated flavonoids displaying SARS-CoV papain-like protease inhibition from the fruits of Paulownia tomentosa. Bioorg Med Chem, 2013. 21(11): p. 3051-7.
- [175] Jo, S., et al., Inhibition of SARS-CoV 3CL protease by flavonoids. J Enzyme Inhib Med Chem, 2020. 35(1): p. 145-151.
- [176] Park, J.Y., et al., Evaluation of polyphenols from Broussonetia papyrifera as coronavirus protease inhibitors. J Enzyme Inhib Med Chem, 2017. 32(1): p. 504-515.
- [177] Lung, J., et al., The potential chemical structure of anti-SARS-CoV-2 RNA-dependent RNA polymerase. J Med Virol, 2020. 92(6): p. 693-697.
- [178] Haggag, Y.A., N.E. El-Ashmawy, and K.M. Okasha, Is hesperidin essential for prophylaxis and treatment of COVID-19 Infection? Med Hypotheses, 2020. 144: p. 109957.
- [179] Alam, S., et al., Traditional Herbal Medicines, Bioactive Metabolites, and Plant Products Against COVID-19: Update on Clinical Trials and Mechanism of Actions. Front Pharmacol, 2021. 12: p. 671498.