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COLLEGE OF ARTS AND SCIENCES
DYNAMICS (SEASONAL AND INTRA-DIURNAL) OF AIR-BORNE
FUNGAL SPORE POPULATION OF DOHA AREA, QATAR

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Abstract

Airborne fungi are considered to cause adverse health impacts on humans, animals and plants. Fungal spores and due to their volume in the atmosphere and small size have an important contribution to the respiratory dysfunctions, allergies and to cause various symptoms range from asthma, allergic rhinitis to bronchitis. The availability and dynamics of airborne fungal spores in the atmosphere are strongly influenced by the meteorological parameters and by other factors such as air pollutants. From 106 settle plate exposures (on alternative days) throughout the period April 2015-March 2016, a total of 1197 mould- and 283 yeast colony-forming units (CFU), twenty one genera and 62 species were retrieved. The highest fungal spore's concentration was recorded in February 2016, whereas the lowest concentration occurred in August 2015. The main constituents of the fungal airspora were attributed to *Cladosporium* (60.2 %), *Aspergillus* (10.4 %), *Fusarium* (9.4%), *Alternaria* (8.5 %), and *Ganoderma* spp. (2.3 %). *Cladosporium* showed two peaks in April and February, while *Fusarium* and *Alteranria* peaked in July. *Aspirgillus* had one peak in August. The prevalence of *Ganoderma* spp. were exclusively detected in February and March.

Temperature was significantly and negatively correlated with the total colony count and fungal species, however no significant correlation was found between relative humidity and both the total colony count and fungal species. Wind speed was significantly and positively correlated with the total colony count and fungal species. However, no significant correlation was detected between wind direction and the incidence of fungal airspora. The correlation between rainfall and either total colony

count or fungal species was non-significant. However, *Alternaria* was significantly and positively correlated with precipitation. Intra-diurnal fluctuations of fungal spores was investigated during the period of 1st of Feb - 31st of March 2016. The highest dispersal of fungal spores favored 18:00 h, whereas at 00:00 h (midnight) the lowest fungal spores release was recorded. The mean daily colony count was negatively correlated with mean daily relative humidity and positively correlated with mean daily temperature. The potentiality of fungal growth on different media on two different culture media, Potato Dextrose Agar (PDA) and Rose Bengal were examined during 1st of Feb - 31st of March 2016 using gravimetric method. No significant difference was observed in total number of fungal colonies or species collected with the two media. Nevertheless, certain fungal taxa were highly selective and thus their growth rate was on one media much higher than with another.

The impact of atmospheric CO₂ concentration on the abundance and diversity of airborne fungal spores were investigated at two different locations. There were no significant differences in the composition and diversity of the airborne fungal population between the two study sites, though daily concentration of CO₂ was higher at the Industrial area site than at Qatar University Campus. Remarkably, the concentrations of *Alternaria* spp. and *Fusarium* spp. were significantly higher at Industrial area site in Corresponding to CO₂ than at Qatar University site

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Dedication

I dedicate this work to my MOM's soul who passed away just two months ago, and during the final step of my thesis work. To whom her prayers still accompany me at every moment of my life, to whom I still hear her voice making prayers and gently asking Allah to empower me, to support me and to be always on my side. May Almighty Allah's blessings grace and mercy be upon her soul.

CHAPTER ONE

INTRODUCTION

Aeromycology is concerning about airborne fungal spores, their release in the air, concentrations, composition and the parameters affecting their dynamics in the atmosphere. As a branch of aerobiology, aeromycological studies help aerobiologists, plant pathologists and allergist by providing baseline knowledge about fungal populations. Airborne fungi are considered to cause adverse health impacts on humans, animals and plants (Harrison *et al.*, 1992; Bush &Portnoy, 2001; Ren *et al.*, 2001; Shelton *et al.*, 2002). The allergen production by fungi is influenced by their life cycle as the allergens are released into the atmosphere from the spores during spore germination (Reponen, 2011). Fungal spores, due to their volume in the atmosphere and small size, have an important contribution to the respiratory allergies and cause various symptoms range from asthma, allergic rhinitis to bronchitis (Vijay *et al.*1991, D'amato &Spieksma 1995, Hasnain *et al.*, 1998). It has been demonstrated that the environmental factors such as meteorological and seasonal climatic factors (Rossi *et al.*, 2005; Klarić & Pepeljnjak, 2006), the type of vegetation (Pepeljnjak & Šegvić, 2003), air pollution (Lin &Li, 2000), and human activities (Mitakakis *et al.*, 2003), affect the variety of air borne fungi. Numerous works on airborne fungal spores have been published worldwide with almost the same objectives to detect the dynamics of the aeromycota in associations with the biotic and abiotic factors of their surrounding environment, among those: India (Chandara & Chanada, 2000; Jothish & Nayar, 2004); Australia (Mitakakis & Guest, 2001); Chile (Ibanez *et al.*, 2001), Poland

(Stepalska & Wolek, 2005; Grinn-Gofroń & Bosiacka 2015) and In China (Wang *et al.*, 2010). Only a limited number of aeromycological studies were conducted in the Middle East compared to other parts of the world, however the similar objectives were introduced. For example; In Palestine (Barkai & Glazer 1962) in Jordan (Abu-Dieyeh & Barham 2014), in Egypt (Hameed *et al.*, 2009) in Turkey (Erkara *et al.*, 2008), in Iran (Nourian *et al.*, 2007; Shams-Ghahfarokhi *et al.*, 2014), in Saudi Arabia (Abdel-Hafez, 1984; Hasnain *et al.*, 2005), in Kuwait (Halawagy, 1989; Khan *et al.*, 1999). Fungal spores are ubiquitous component of the bioaerosols. The availability and survivability of the aeromycota are known to be influenced by their interactions with biotic and abiotic components of the environment such as meteorological factors, geographical location, air pollutions, vegetation cover, and anthropogenic activities. The effects of meteorological factors on the dynamic of airborne fungal spore's populations were intensively reviewed in the literatures. However; other parameters such as the impact of air pollution are not studied as much as meteorological factors.

In Qatar only one study was carried out by Al-Subai (2002) to reflect the airspora of Doha for the whole year (1997-1998). The author concluded that the mycoflora in the air of Doha exhibited a seasonal and diurnal variation in which *Cladosporium*, *Alternaria* and *Ulocladium* were the most abundant genera in the atmosphere of Doha (Al-Subai, 2002). Qatar is a peninsula located in the Arabian Gulf region. Plant coverage is infrequent and scattered due to the shortage in water supplies, mainly varied between herbaceous plants, shrubs and limited number of tree species (Syed & Abdullah, 2002). The most of Qatari's land are arid desert with a scarce annual precipitation. According to Batanouny (1981), the

landforms system in Qatar is mainly comprised of the following phyto-ecogeomorphologies: rocky and conglomerates hamads which occupy the major land in Qatar, rocky ridges where the vegetation in both systems are poor, depressions, wadies and runnels and sabkhas (salt marshes) where the soil is of high salt content and the water table is near the surface (Batanouny, 1981). Doha is the capital of Qatar and the most popular city. Huge anthropogenic changes including urbanization, industrialization, and establishment of housing compounds, shopping centers and high-way roads were occurred in Doha within the last 15 years. Those anthropogenic changes are expected to affect the dynamics and species composition of air-borne fungal spores.

This research project was mainly initiated to update the knowledge about the seasonal and diurnal variations in airborne fungal spores of Doha area and to correlate these variations with meteorological factors. The present study also tackled with other parameters, atmospheric CO₂ concentrations and different culture media, which might provide more information about the dynamics of fungal spores in relation to their surroundings. More specifically, this study was conducted to achieve the following objectives:

- (1) Provide baseline knowledge about density, diversity and dynamics of airborne fungal spore in the atmosphere of Doha using settle plate exposures. This knowledge is necessary for mycologists, applied ecologists, plant pathologists and other disciplines of biological, environmental and health sciences.
- (2) Investigate variations in seasonal and diurnal distribution of airborne fungal spores, abundance, density and diversity and to correlate these variations with meteorological factors.

- (3) Investigate variations in diurnal distribution of airborne fungal spores, abundance, density and diversity during February and March (2016).
- (4) Correlate the variations in species composition and abundance of air-borne fungi with atmospheric CO₂ concentration in two different areas in Qatar during February and March (2016) in order to investigate the impact of urbanization on air-borne fungal spore population.
- (5) Compare results obtained from two different cultural selective media: Potato Dextrose Agar (PDA) and Czapek's/ Rose-bengal media during February and March 2016.

CHAPTER TWO

LITRTURE REVEIW

2.1. The Science of Aerobiology

The science of aerobiology deals with air-borne agents, the processes of aerosolization, the aerosol itself, and to a lesser extent, the exposure and response (Burge, 2002). Biological materials which exist in the atmosphere, or in other wards bioaerosols, are consisting of airborne particles of biological origin including fungi, bacteria, pollen and viruses, as well as their by-products such as endotoxins and mycotoxin, fragments and other fragments or waste products of living organisms like animal allergens (Reponen, 2011). In the atmosphere, the performance of any particle will be depended mainly on its shape, density and size .The numerous types of particles involving in biological aerosol are of different sizes. While pollens might have the largest diameter, anemophilous plants's pollens are of typical diameters ranges from 17 till 58 μm , and to lesser extent the diameter of fungal spores which ranges from 1to 30 μm (Chatterjee & Hargreave, 1974), viruses are thought to be the smallest in its surrounding, they have a typical diameter of less than 0.3 μm (Taylor, 1988). Bacteria are ubiquitous in the atmosphere representing a considerable portion of the bioaerosols. The size of bacterial cells ranges between 0.6 and 2 μm . Certain infectious bacteria are windblown cells which can affect the man health such as *Mycobacterium tuberculosis* (Bowers *et al.*, 2011).

In addition to naturally occurring sources of bioaerosols such as, plants, animals and biological decomposition of organic matters, various human activities leads to enriching the ambient atmosphere with biological aerosols . Waste disposal facilities, agricultural

and food production activities are examples of potential facilities that represent bioaerosol sources of emission (Kummer & Thiel, 2008). Pollen grains are such an important consistent of outdoor bioaerosol which found to exacerbate allergenic reactions. Pollen grains are produced by many trees, weeds and grasses. Dispersal by wind is the main mechanism of pollens distribution in the air, they absorb moisture from the atmosphere until they are swollen enough to breach and release pollen allergens out to the air (Reponen, 2011). Their concentration in the air is highly dependent on temperature and humidity; higher temperature and lower humidity drive higher pollen grain concentrations. The effect of wind velocity depends on pollen aerodynamic diameter (Jones & Harrison, 2004). Because of their relative large size, the pollen allergens which are associated with particles of less than 5 μm in diameter are able to penetrate lower in the lungs and subsequently causing asthma (Burge, 2002).

Among the most common bioaerosol particles either in outdoor or indoor are fungal spores. Aeromycological studies are mainly dealing with airborne fungal spores providing baseline information to plant pathologists, allergists and aerobiologists. Air-borne fungal spores are of primary importance for the induction of plant diseases and public health hazards in term of allergic reactions to fungal spores. The main source of outdoor fungal aerosols is plant materials (Moubasher, 1993). Fungal diversity has been found to be greatly influenced by the existence of plants and much less by bare sediment (Mohamed & Martiny, 2011). However, the significant changes with ecology of plants can lead to considerable changes in fungal air spores (Burge, 2002). Soil is another considerable source of fungal spores (Rousk & Baath, 2011). Physical characteristics of the soil such as pH and temperature are

factors shaping the fungal communities and activities which subsequently affecting the release rate of airborne fungal spores (Rousk & Baath, 2011; Yuste *et al.*, 2011). Human activities can induce changes in airborne fungal spore's sources; global warming and increasing atmospheric CO₂ level are examples. The usage of fungi as bio-control agents is also contributing to human-induced sources which might disturb fungal spore's distribution. As an example, spreading *Epicoccum nigrum* conidia onto sunflowers can control pathogens affecting this specific plant but it might rise the distribution of this significant allergenic fungus in the atmosphere (Pieckenstain *et al.*, 2001). The exposure to fungi is associated with public health hazards because they can cause infections, irritations, allergies and toxic effects (Agrios, 2005; Mohammadi & Krumbein, 2008).

2.2. Airborne fungal spores and allergy

During the past decades, the changes in environment have associated with an increase in prevalence of allergies (Koppelman, 2007). Fungi are abundant in the environment and they have a great ability to grow either as saprophytes colonizing nonliving organic substrate or as pathogens penetrating living tissues, from where they become airborne (Green *et al.*, 2005). The personal exposure to airborne fungal spores and their components is strongly linked to allergic diseases and dysfunction of respiratory

system including allergic rhinitis, bronchopulmonary, and hypersensitivity diseases (Koppelman, 2007) pneumonitis and bronchial asthma (Masoli *et al.*, 2004; Bousquet *et*

al., 2008). In general, the admission rate to emergency department because of allergy to airborne fungal spores increases and hospitalization period for respiratory diseases extends as well (Dales *et al.*, 2004; Atkinson *et al.*, 2006).

The common mechanism of fungal airospora distribution in the atmosphere is through asexual conidia or sexual spores which considered significant aerosol particles. Quantities of fungal conidia are generally dependent on the environments of indoor and outdoor, different seasons, weather and climatic conditions, and topographical features. One hundred genera of fungal conidia and more are recently documented as a source of inhalant allergens (Green *et al.*, 2005). Airborne fungal spores can range from zero to higher than one hundred thousand colony forming units (CFU) per cubic meter of air. According to Horner *et al.*, (1995) among the 69,000 fungal species defined, around 80 species have been connected to allergic respiratory illnesses which usually intermediated by IgE hypersensitivity. Most molds are capable of producing highly allergenic proteins or glycoproteins which can exacerbate the allergic symptoms in atopic hosts (Reponen, 2011). The productions of fungal allergens are widely dependent on fungal species and on the media where they develop and grow. Moreover, life cycle of fungi can affect the allergens production; commonly spores release their allergens into the surrounding environment during germinations (Green *et al.*, 2005)

Spore germination of *Alternaria* and *Aspergillus* is found to increase the number of allergens (Green *et al.*, 2005). Green *et al.*, (2003) concluded that allergens were released from 11 fungal species, *Alternaria alternata*, *Cladosporium herbarum*, *Aspergillus fumigatus*, *Botrytis cinerea*, *Epicoccum nigrum*, *Exserohilum rostratum*, *Penicillium*

chrysogenum, *Stemphylium botryosum*, *Curvularia lunata*, *Trichoderma viride*, and *Bipolaris spicifera*. 5.7% - 92% of spores of 9 out of 11 fungal species mentioned above liberate their allergens before germination (Green *et al.*, 2003). However, all the germinated spores released allergens from their hyphae after germination and the total quantity of allergens was recognized to be greater after germination than before it, and thus germination is a principal contributor in allergen release mechanism for many allergenic fungi species even though many of non-germinated spores of certain species contributed to allergens release as well, but in small amounts (Green *et al.*, 2003). Mitakakis *et al.* (2001) conducted a study to examine if the spore germination of *Alternaria* increases the allergen release, they revealed that a greater allergen elution from spores occurs by germination, yet not all spores release allergens.

As it was demonstrated by skin test, 10% to 60% of vulnerable hosts (atopic) represent hypersensitivity to molds (Burge, 2002). The most commonly known allergenic fungi include *Penicillium*, *Aspergillus*, *Cladosporium*, *Alternaria*, *Curvularia*, *Bipolaris*, *Epicoccum*, and *Candida* (Vermani *et al.*, 2011). Most of them can penetrate downwards to lower part of pulmonary airways and cause hypersensitivity reactions. In Qatar, an observational study was carried out to estimate the size of fungal infections at public level (Taj-Aldeen *et al.*, 2015). The data analysis revealed that, 1486 people were affected by severe asthma with fungal sensitization, 1126 patients were diagnosed with allergic bronchopulmonary and 176 individuals complained from chronic pulmonary aspergillosis (Taj-Aldeen *et al.*, 2015). The majority of fungal spores have a size range of 2-10 μm though a significant quantity of fungal components including allergens, glucan and

mycotoxins occur in a size less than 1 μm . The size of conidia and fungal spores detect the deposition location in lungs and the severity of allergenic reactions. Conidia and spore size of 5 μm and above is linked to direct type of hypersensitivity but with smaller size, about 1 μm , the hypersensitivity is said to be of delayed type where the conidia and spores are able to penetrate further downward of respiratory tract (Kurup & Fink, 1993). Additionally, the mode of spore's entrance into respiratory system determines the deposition site in lungs (Burge, 1989; Kurup, 1989; Kurup & Fink, 1993). The conidial clusters of *Aspergillus fumigatus* are commonly settled down in the upper airways tract, but with individual conidia, the deposition site are usually in lower airways (Kurup *et al.*, 2000). Approximately 60 % of *Clavatia* species spores have been found to penetrate lower respiratory tracts into alveoli, due to their relative smaller sizes (Burge, 2002). According to Green *et al.*, (2005), fungal hyphae fragmented conidia and new fungal genera belonging to Ascomycetes, Deuteromycetes, and Myxomycetes are considered as aeroallergen sources and subsequently contribute to inhalant allergens.

In addition to allergens, molds contain other biologically active constituents including (1-3)-B-D-glucan and mycotoxins. (1-3)-B-D-Glucan is a principle cell wall structure of fungi and other microorganisms, comprising about 60% of fungal cell wall (Sharpe *et al.*, 2015). (1-3)-B-D-Glucan exposure is related to some of pulmonary diseases such as airways inflammation by which more common with infants and schoolchildren (Jaakkola *et al.*, 2010). Mycotoxins are secondary fungal metabolites in which their production depends on the environmental conditions like temperature and the availability of moisture and nutrients (Reponen, 2011). According to the later author, mycotoxins are common aerosol in

significantly contaminated agricultural environments. In Saudi Arabia, an aerobiological study was conducted in three sites (Jizan, Hail and Taif) revealed the presence and prevalence of *Ganoderma* basidiospores which cause respiratory allergic diseases. Three study sites were included in the investigation. One site (Jizan) had the highest concentration of *Ganoderma* basidiospores which interestingly correlated to the highest level of asthma prevalence among children through the three study sites (Hasnain *et al.*, 2004).

2.3. Factors affecting availability and survivability of airborne fungi:

The air pollutants are well known causes for adverse effects on both the health of human and environment. Biological and non-biological contaminants are together associated with the increase of allergenic symptoms, recurrent visit to emergency room and hospitalization period or even motility. Several studies have examined the potential consequences of non-biological pollutants, physical components of the environment and meteorological parameters on the dynamics and concentrations of airborne fungal spores (Al-Subai, 2002; Sousa *et al.*, 2008; Abu-Dieyeh *et al.*, 2010; Hameed *et al.*, 2012; Abu-Dieyeh & Barham, 2014).

2.3.1. Abiotic factors:

The diversity or composition of fungi is thought to be determined by CO₂ concentration in the atmosphere (Klamer *et al.*, 2002), soil depth (O'Brien *et al.*, 2005), nitrogen availability (Allison *et al.*, 2007) and salinity (Mohamed & Martiny, 2011). It was concluded that, fungal biomass in the soil increased due to the treatment with elevated atmospheric CO₂; however, the treatment with elevated atmospheric CO₂ didn't affect either the fungal community composition or their species richness (Klamer *et al.*, 2002). It is believed that, plants and through the richness in the quality and quantity of resource supply, to significantly affect soil micro biota (Mohamed & Martiny, 2011). Plant diversity adds to the fungal richness and diversity by providing more organic substrate diversity, increasing habitat complexity, decreasing surface temperature, reducing surface salinity and increasing O₂ availability (Zak *et al.*, 2003; van der Heijden *et al.*, 2008; Mohamed & Martiny, 2011;). In their study, O'Brien *et al.*, 2005 demonstrated that, different soil samples showed compositional variances, while mycorrhizal species were the most abundant fungi in deeper soil profile, saprophytic fungi predominated the upper litter layer (O'Brien *et al.*, 2005). Nitrogen is a basic nutrient required to promote healthy growth of any living organisms. Yet, increased nitrogen deposition due to N fertilization was reported to decline the fungal taxonomic richness and alters community structure (Allison *et al.*, 2007). Other abiotic factors can influence the composition and diversity of fungal populations. In Jordan, mycorrhizal abundance and distribution was not influenced by soil pH, soil phosphorus, or soil texture, but it was strongly and positively correlated to organic matter and CaCO₃ percentages (Mohammad *et al.*, 2003). Salinity is another factor that can

significantly affect the availability of fungal communities. Sediment fungal composition was found to be influenced by salinity gradient more than plant appearance. In contrast, greater fungal diversity was detected by the presence of plants and much less by salinity, only with intermediate salinity, greater fungal diversity was detected (Mohamed & Martiny, 2011). In Kuwait, the saline soil yielded 14 species of *Aspergillus* revealing that the saline soil may provide a suitable habitat for considerable number of fungi and subsequently contribute to the aerial occurrence of the related fungi (Moustafa, 1975).

2.3.2. Meteorological parameters:

Bioerosols contribute significantly to atmospheric pollutants in which fungal spores are an important portion of them. Environmental variables such as temperature, relative humidity and wind speed affect fungal spores' concentration in the atmosphere, their growth and reproduction as well. The concentration of airborne fungal spores in the atmosphere is varying daily and seasonally depending on many environmental variables, among them air pollution, meteorological parameters, human activities and seasonal climatic factors (Lin & Li, 2000; Mitakakis *et al.*, 2003; Rossi *et al.*, 2005; Abu-Dieyeh & Barham, 2014). The most important indicators of fungal airspora population in the air are meteorological variables. Temperature and humidity are of highly significant positive effects on the total concentration of fungal population (Aydogdu & Asan, 2008; Oliveira *et al.*, 2009).

Temperate and tropical regions were found to accommodate higher concentration of airborne spores, while in desert region the number is much lesser (Lacey, 1981). As an example of the concentration of air-borne fungal spores collected from Amman area (humid Mediterranean climate) was 461 779 spores (Shaheen, 1992) compared to 76 396 fungal spores collected from Zarqa area, Jordan (Arid Irrano-terranean climate) (Abu-Dieyh & Barham, 2014). The most significant meteorological factor which positively correlated to fungal airspora is the temperature. A positive correlation was found between temperature and airborne fungal spores (Aydogdu & Asan, 2008), including certain fungal genera *Aspergillus* (Adhikari *et al.*, 2006), *Cladosporium* (Quintero *et al.*, 2010) and *Alternaria* (Reyes *et al.*, 2009). Grinn-Gofroń & Bosiacka, (2015) demonstrated that and through a 4-years study, *Alternaria*, *Drechslera* type and *Cladosporium* spores were displayed their maximum abundance at higher mean temperature values. Al-Subai, (2002) deduced that, the greatest density of fungal spores was recorded in July and December. Abu-Dieyh *et al.*, (2010) revealed that, the highest frequency of fungal spores in Zarqa area in Jordan was found to occur during winter and rainy months of the year. According to Oliveira *et al.*, (2009) the maximum concentration of fungal spores was observed during summer and autumn. Supporting previous findings indicated that, during the hot and dry days, the population of the following airborne fungal spores are prevailed: *Alternaria*, *Torula*, *Cladosporium*, *Drechslera*, while *Epicoccum* spp. were the highest when the moisture is relatively high (Grinn-Gofroń & Bosiacka, 2015).

Wind speed showed inconsistent effects on the distribution of airborne fungal spores. Some publications demonstrated that wind velocity can enhance microorganisms' dispersion in

the atmosphere by detaching them from their attached surfaces (Jones and Harrison, 2004). On other hand, other studies found the wind speed might dilute fungal spore's concentrations in the air (Sabariego *et al.*, 2000; Stennett & Beggs, 2004). Highest wind speed might bring to higher fungal spores concentration (Al Subai 2002; Jones & Harrison, 2004). When the wind speed is less than 5m/s, the fungal spore's concentration decreased, but when the wind velocity is more than 5m/s the concentration of fungal spores increased (Lin & Li, 2000). On the other hand, wind can act as a dispersal and diluting factor for spores when the wind is high and so decrease the concentration of spores significantly (Quintero *et al.*, 2010). Fluctuations in the wind velocity are significantly associated with changes in spore's concentration and particularly when other meteorological factors are optimum (Sakiyan & Inceoglu, 2003). The study of Grinn-Gofroń & Bosiacka (2015) showed that the highest release of *Alternaria*, *Cladosporium* and *Drechslera* type spores occurred when the wind speed is at the lowest value of the mean, while the concentration of *Leptosphaeria* and *Didymella* spores increased when the mean of wind speed is the highest. Abu-Dieyeh & Barham (2014) found the *Puccinia* spores are positively and significantly correlated with wind speed, while *Alternaria*, *Cladosporium*, *Drechslera* and *Ustilago* didn't show any significant correlation with the wind velocity.

The study of Pakpour *et al.*, (2015), which conducted for over 20 years period in two North American cities (New York and Toronto) of humid continental climate type, demonstrated that airborne fungal spores are determined by annual variations in climate especially for precipitation and to a lesser extent for temperature. During drier years, a higher fungal spores in the air was reported (Pakpour *et al.*, 2015)

Rainfall might act as a vacuum cleaner which cleans the air by wet deposition mechanism and thus fungal spores are forced to settle down on the ground or other surfaces (Katial *et al.*, 1997; Polymenakou, 2012). Contrary results were noted by Abu-Dieyeh *et al.* (2010) who recorded the highest colonies count events post the heaviest precipitation and moderate temperature conditions, in Zarqa area, Jordan (Arid Irano-terranean climate). Heo *et al.* (2014) agreed with Abu-Dieyeh *et al.*, (2010), they found the concentration of fungal bioaerosols was much higher during rainy season than in non-rainy days. In such studies, the positive correlation between fungal bioaerosol's concentrations and rain was due to the fact that rain helps creating an environment of required moisture level for better fungal vegetation process, and due to the absence or reduction in solar radiation as well. After all, the effects of meteorological factors on the diversity and concentrations of fungal airspora is a complex relationship between biological and environmental factors. Thus, it is difficult to assess the importance of each meteorological factor separately due to the instantaneous response of fungi to a combination of parameters (Abu-Dieyeh *et al.*, 2010).

2.3.3. Air pollution:

The interrelationship between atmospheric pollutants, meteorological factors and airborne fungi are thoroughly investigated in the literature (Adhikari *et al.*, 2006; Sousa *et al.*, 2008). Although the impact of meteorological factors on air quality is considerable and well established, studies concerning the relationship between fungal counts and air pollutants

are still few (Adhikari *et al.*, 2006). Among those pollutants are: ozone (O₃), nitrogen dioxide (NO₂), sulphur dioxide (SO₂) and particulate matter (PM10). Tropospheric ozone levels have been significantly increasing in the last decades because of uncontrolled anthropogenic emissions (Sousa *et al.*, 2008). Ozone in the troposphere resulted from photochemical reactions involving solar radiation and emitted pollutants from earth surface (Marenco *et al.*, 1994). In contrast to stratospheric ozone, tropospheric ozone has been associated with adverse impact on human health, climate and even on atmospheric composition. A significant positive correlation between O₃ and spore concentration of *Alternaria* and *Cladosporium* were reported (Adhikari *et al.*, 2006; Grinn-Gofron *et al.*, 2011). The concentration of tropospheric O₃ was found to be affected by relative humidity and moreover the highest average level of O₃ favors relative humidity of 40% or lower (Van Weele & Ma 2000; Elminir, 2005; Grinn-Gofron *et al.*, 2011). Also, higher O₃ production was recorded during warmer years when more UV radiation and lower wind velocity are the weather trend (WHO, 2000; Sousa *et al.*, 2008). On the other hand the concentration of fungal spores such as *Cladosporium* sp. can be significantly affected by high O₃ concentration in which high O₃ level is capable of killing microorganism by oxidizing their cellular component and cell walls as well (Das *et al.*, 2006; Whangchai *et al.*, 2006). A negative or no significant correlation between O₃ and fungal spores in the air was demonstrated by Sousa *et al.* (2008). Similar results were obtained by Ho *et al.* (2005), who deduced from a 3 years study period (1994-1996) a negative correlation between O₃ and general fungal spores, total fungal spores and in particular with *Ganoderma* spp.

SO₂, NO₂ and PM₁₀ concentrations in the atmosphere were thought to affect the fungal spore survivability. The concentration of fungal airospora decreased when SO₂ increased (Asan *et al.*, 2002). The influence of SO₂ and NO₂ on *Cladosporium* and *Alternaria* spores was rather low or weak (Grinn-Gofron *et al.*, 2011). NO₂ and SO₂ in the atmosphere are going under chemical reactions and their chemical products are found to be toxic to microorganisms (Won & Ross, 1969; Harrison and Perry, 1986; Ho *et al.*, 2005; Adhikari *et al.*, 2006; Grinn-Gofron *et al.*, 2011). PM₁₀ presented a positive trend relationship especially with *Aspergillus* and *Alternaria* (Adhikari *et al.*, 2006). PM₁₀ can affect the morphology, allergenicity and aerodynamic attributes of bioaerosols (Adhikari *et al.*, 2006, Ormstad *et al.*, 1998). However, Lin & Li (2000) and Sousa *et al.*, (2008) concluded that there is a non-significant positive correlation between PM₁₀ and *Aspergillus* or *Cladosporium*. Because of their chemical composition, PM₁₀ might severely affect fungi's metabolism processes or even adsorb them on their surfaces (Matthias-Maser *et al.*, 1998). It is demonstrated that, the concentration of both airborne bacteria and fungi were lower in heavy haze days than during non- haze days (Gao *et al.*, 2015). With respect to increasing CO₂ level, when accompanied with drought it will have a positive effect on soil fungal communities particularly on pathogenic strains such as *Fusarium* and thus indirect effect on their airborne spores (Curlevski *et al.*, 2014). Klironomos *et al.* (1997) concluded that the increasing level of atmospheric CO₂ can have a recognized effect on airborne fungal vegetation and subsequently on their dispersal rate in the atmosphere and air quality.

In general, many studies demonstrated that, there were no significant correlation between air pollutants and fungal survivability in the air (Ho *et al.*, 2005; Wu *et al.*, 2007; Lin & Li,

2000; Sousa *et al.*, 2008). However, the lack of proves is not prove of lack important effect of air pollutants on fungal spores viability in the air.

2.3.4. Diurnal variations:

The number of airborne fungal spores varies from season to another, from day to day and from hour to hour. Accordingly, diurnal variations were frequently studied to describe the pattern of fungal airspora throughout the day. Different airspora types showed different diurnal periodicity trends. The following studies are just few examples of what is found in the literature. According to Sreeramulu (1959) who studied the seasonal and diurnal variations affecting the airborne fungal spores in UK, when *Ustilago*, *Helminthosporium*, *Uredospores*, *Botrytis*, *Alternaria*, *Stemphylium* and *Periconia* spores were at their maximum concentration in the afternoon, their minimum concentrations were used to be in the early morning. Before noon, *Erysiphe*, *Polythrincium* and *Epicoccum* reached their maximum. Between midnight and dawn, *Ganoderma*, *Coniophora* and other dark colored basidiospores were the most abundance (Sreeramulu, 1959). In Finland, the circadian periodicity of certain airborne spores was investigated. *Cladosporium* spores released at their maximum around noon, the concentration of *Suillus* spores had a minimum concentration at 16:00 h-18:00 h (Helander & Pessi, 1991). In Ontario-Canada an aerobiological study was conducted to monitor the outdoor air mycoflora throughout 1992. The diurnal pattern of fungal spores varied. The maximum concentration of *Cladosporium* and *Epicoccum* showed a midday peak pattern. The maximum concentration of

Cladosporium occurred at 14:00h while the minimum value occurred at 6:00 h. *Epicoccum* peaked at around 10:00 and 4:00 and dropped at midnight. No clear diurnal pattern was shown by *Aspergillus* and *Penicillium* (Li & Kendrick, 1995).

Seasonal and diurnal fluctuations of airborne basidiomycete spores group in Mexico City were observed for one year. Basidiospores occurred at their greatest concentration in the early morning, while their lowest concentration was recorded in the afternoon (Calderon *et al.*, 1995). In Qatar, Al-Subai (2002) demonstrated that the diurnal pattern of total fungal spores showed one peak at noon time and the lowest fungal spores distribution was at midnight and *Cladosporium* showed a regular peak at noon time. The positive correlation between mid-day and fungal spores' concentration was reflected by high wind speed, high temperature and low relative humidity while at midnight, lower temperature, lower wind velocity and higher moisture command the trough phenomenon (Al-Subai, 2002). In Zarqa area, Jordan, Abu-Dieyeh *et al.*, (2010) studied diurnal variations of fungal airspora in two different periods of the year, from February to April and from November to January. They found that the total fungal spores presented their maximum distribution pattern at 15:00 hour versus 10:00 hour of collection samples during November and January. *Cladosporium*, *Fusarium* and *Alternaria* spp. favored their spores release at 15:00 hour which might be due to lower relative humidity at early afternoon (Abu-Dieyeh *et al.*, 2010). Unlike those cold months, February and April had no significant differences in terms of diurnal variations (Abu-Dieyeh *et al.*, 2010). Abdel Hameed *et al.* (2009) conducted a study to evaluate diurnal variations of airborne fungi and bacteria in Helwan, Egypt. They

revealed that the maximum distribution of fungal airspora was at 20:00 h during three seasons, except for winter season in which the highest concentration was detected 2 h earlier than other patterns. *Aspergillus* showed an outline of doublepeaks, *Penicillium* and *Cladosporium* achieved their peak at 20:00 while *Alternaria* was favoring a midday spore release (Hameed *et al.*, 2009).

Weather conditions throughout the day controlled the fungal spore's distribution pattern, but also the location and human activities can contribute effectively. In Dublin, Ireland, O'Gorman & Fuller (2008) deduced that there was no significant variations between morning, afternoon and evening counts of total fungal airspora; however they realized that the highest fungal spores concentrations to happen in the summer mornings (June-August), yet the variation was not statistically significant.

As indicated by above mentioned literature intra-diurnal variation in abundance of airspora is not systematic and variable with region, season and weather conditions which mean that the local environment is the most limiting factor in determining the trend of diurnal variation of airspora.

2.3.5. Sampling methodologies:

The sampling of airborne fungal spores are commonly conducted using either the gravimetric method (settle plate exposures) or the volumetric method by sucked a known amount of air using spore traps such as Burkard spore trap (Hirst, 1952).

Gravimetric method is a simple, non-volumetric, passive sampling method collecting spores by settling on the surface of agar-containing plate. The collected samples are simply incubated in an incubator for definite time and temperature degree, and then the recovered fungi are classified based on their macro morphology and micromorphology features. Nonetheless, it is associated with some drawbacks which might affect its validity; for instance: any clustered colonies will not be distinguishable and counted as one, not time discrimination i.e. it is difficult to repeat it several times through the day, the spores which recover on the laboratory substrata are only the viable ones, and thus it cause a bias in the fungal types grow on the agar media (Mostafa & Kamel, 1976). Spore trap is an active method that use artificial force to control and help spore collection. The vacuum pumps a volume of air of 10 L/min. rate, airborne particles adhere to a tape covered with a sticky substance; this tape is then removed and divided into 7 segments that are placed on microscopic slides and stained for further investigation (Sabariego *et al.*, 2007). Yet this sampler is an expensive and labour extensive (slides preparation, tape replacement) method. No strong evidence that favors either of the two methods, gravimetric method or spore traps. Abu-Dieyeh & Barham (2014) mentioned a detailed discussion about the potentiality of the two methodologies after two different studies (Abu-Dieyeh *et al.* 2010, Abu-Dieyeh & Barham, 2014) based on two different methodologies on the same area. According to the above authors, *Cladosporium* was found to be the main constituent of airborne fungal spores using both settle exposure and spore trapping methodologies. *Alternaria*, *Ustilago* and *Drechslera*, in addition to *Cladosporium* comprise the main percentage of the total tapped fungal spores in the air using spore trapping method. Even

though; *Alternaria* quantity, which detected using spore trapping method, showed similarity to which revealed by settle exposure method in Zarqa, Jordan (Abu-Dieyeh *et al.* 2010, Abu-Dieyeh & Barham, 2014), the genera *Puccinia*, *Ustilago* and *Drechslera* are biotrophic fungi, which do not grow on culture media and, consequently, were not reported using the settle exposure method (Abu-Dieyeh & Barham, 2014). A genus like *Fusarium*, in comparison to *Puccinia*, *Ustilago* and *Drechslera*, is recognized to be better detected on settle exposure rather than spore trapping (Mitakakis & Guest 2001), and thus it was introduced as a rare constituent of the total tapped fungal spores in the air of the Zarqa area (Abu-Dieyeh & Barham, 2014). It was demonstrated that, *Aspergillus/Penicillium* group represented a small percentage of the total airospora reported by spore trapping method (Abu-Dieyeh & Barham, 2014). In contrast, they showed higher percentage using the settled exposure method (Abu-Dieyeh *et al.*, 2010). Because of the relative smaller diameter of their spores (1–2 μm) (Mitakakis & Guest 2001) and the disability of Burkard trap to accurately collect particles less than 5 μm in aerodynamic diameter (Hirst, 1952), *Aspergillus* and *Penicillium* spores are under- estimated using spore trapping method. Unlike, the easily identifiable spores of *Alternaria* and *Epicoccum*, for example, it is difficult to visually distinguish between *Aspergillus* and *Penicillium* conidia collected using a spore trap (Eduard, 1996). Nevertheless, to conclude better information about aeromycology of an area, both settle exposure and spore trap methodologies are recommended and they are complementary to each other (Abu-Dieyeh & Barham, 2014). To a lesser extent, Anderson impactor is used as an aeroallergen sampler as well. This sampler is a combination between volumetric and culture methods. Both, the concentration

and the size of airborne particles are measured. The six stages type consist of filters combined with petri dishes in which the air flows and so the biological aerosol of different diameters settle on the respected stage; later, the estimation of airborne particles is done based on the count of grown colonies (Gillespie *et al.* 1981; Jensen *et al.*, 1992; Stein, 1999; Wu, 2000; Nesa *et al.*, 2001).

The slide exposure method is a kind of gravity sampling technique where the slides are smeared with a sticky material, and then the slides are exposed to the atmosphere for certain period of time. Later the slides are collected and then carefully stained with a dye and a cover slip is placed, then the slide will be studied to quantify fungal spores under the microscope. This method can help identifying the viable but not culturable fungal spores (Frey & Durie, 1962; Agarwal *et al.*, 1969)

Frey & Durie (1962) used the slide exposure method and culture plates to study the availability of fungal spores in the atmosphere of Sydney. The number of colonies isolated with culture plates was higher than the total number of fungal spores counted on slides. The most dominant fungal spores counted on the slides were of *Alternaria*, *Cladosporium* and rusts and smuts. While the predominant fungal colonies recovered with culture plate method were of *Cladosporium*, *Alternaria*, *Epicoccum*, *Pleospora*, *Penicillium*, *Pullularia* genera.

In Delhi, India an aeromycological study was conducted by Agarwal *et al.* (1969) to assess the prevalence of fungal spores in Metropolitan area and their relationship with allergic cases incidence using two different methods; plate exposure method and slide exposure method. Results detected that *Alternaria* spp., *Cladosporium* spp. and spherical spores

were the predominant using slide exposure method, whereas *Cladosporium* spp., *Alternaria* spp., *Aspergillus* spp., and *Fusarium* spp. spores were the prevalent when Petri dishes were used (Agarwal *et al.*, 1969).

The Durham sampler is another gravimetric method which technically has the same concept as slide exposure method. Slides, before air exposure, are covered with a sticky jell then placed on the Durham sampler for a certain period of time, then they are collected to identify the fungal spores and pollen grains on the microscope. This method can be used for both culturable and non-culturable fungal spores; however it is most frequently used to study the prevalence of pollen grains in the atmosphere (Crispen *et al.*, 2010; Erkara *et al.*, 2009). Eskisehir City, Turkey, the relationship between *Alternaria* and *Cladosporium* with meteorological conditions was investigated using Durham gravimetric sampler. *Cladosporium* spores count was as two third as *Alternaria* spores and their availability was influenced significantly by relative humidity (Erkara *et al.*, 2009).

2.3.6. Culture media:

The identifiable growth of viable airborne fungal species on laboratory media is basically controlled by contained nutrients. Thus, the germination and growth of these spores are strongly dependent on the employed media. Burge *et al.* (1977), used eight different culture media (Modified Mehrlich's medium (MM) , Sabouraud dextrose agar (SDA), Malt extract agar (MALT), V8 juice agar (V8), Rose Bengal-streptomycin agar, Casein hydrolysate agar (CH), and Potato dextrose agar (PDA) (with four different Rose Bengal

concentrations) to compare the fungal spores recoveries. They concluded that, the total growth were similar on SDA, MALT, V8, and PDA, whereas MALT and SDA exhibited the highest occurrences of growth pattern, mostly of all colony types. *Epicoccum* was excluded by CH and MM; however, *Cladosporium* presented high recovery on those media. PDA containing Rose Bengal produced lower total recoveries in comparison with other media (Burge *et al.*, 1977). In Saudi Arabia, Abdel-Hafez (1984) used two different media, glucose- and cellulose-Czapek's agar to study the prevalence of air mycoflora at Taif. The recovery of *Aspergillus* spores were higher on glucose media than it occurred on cellulose culture, while *Alternaria*, *Cladosporium*, *Drechslera*, *Scopulariopsis* and *Phoma* were greatly recovered on both media. *Penicillium* and *Ulocladium* were highly selective, while *Penicillium* showed high growth pattern on glucose, *Ulocladium* were recovered mainly on cellulose (Abdel-Hafez , 1984).According to Moubasher, (1993) 50% sucrose Czapek's agar provides the highest total count of fungal recoveries and the widest species spectrum in comparison with glucose and cellulose-Czapek's agar. Mcquilken *et al.*, (1997) proved that not only temperature, pH and light affect the development on fungal growth, but also the laboratory substrata can drive the developmental process. Both, Potato-dextrose agar (PDA) and malt extract agar (MEA) produced the highest rate of conidial germination, pycnidial production and hyphal extension of *Coniothyrium minitans*. Hyphal extension rate on molasses-yeast agar was more dawdling, other developmental processes rate were similar to those on PDA and MEA (Mcquilken *et al.*, 1997). A single stage / N6 Andersen impactor which was attached with agar plates of different media, MEA and Dichloran Glycerol (DG18) was used to compare the fungal concentration and genera

recovered on the different culture media. In comparison with MEA, DG18 media supported higher fungal concentrations. Furthermore, the genera of *Aspergillus*, *Penicillium*, *Fusarium*, yeast, and nonsporulating fungi yielded significantly higher concentration on DG18 than with MEA (Wu *et al.*, 2000). For the quantification of culturable fungi purpose and assessing the relative efficiencies of different culture media, three culture media were used (CIP10-M, malt extract agar (ME) and dichloran glycerol-18 (DG18). Higher colonies of *Aspergillus* spp were acquired on DG18 and ME at 25 °C, whereas greater CFU values of *Scopulariopsis* spp. with ME at 37°C. Unknown species were obtained with higher values with DG18 at 25 °C than on ME at 25 °C or at 37 °C. *Alternaria* spp. was isolated at higher rate with ME and CIP10-M. Only ME media supported the recoveries of Mucorales (Niegutsila *et al.*, 2011).

2.4. Worldwide relevant aeromycological studies

Numerous works on airborne fungal spores and their responses to their surrounding environment and their correlation with respiratory diseases have been established worldwide. In Chicago, USA Feinberg and his co-authors (1936) surveyed the atmosphere content of air borne fungal spores to determine the diversity and dynamic of fungal population to correlate them with the allergic incidence reacting to mold exposure. They used gravimetric methods to collect their samples for one year (1934-1935). *Alternaria*, *Hormodendrum* (*Cladosporium*), *Penicillium*, *Aspergillus*, *Rhizopus*, *Mucor*,

Chaetomium, and *Monilia* were the predominant spores through the year, yet they were more prevalent in the warmer months. *Alternaria* and *Hormodendrum* showed a seasonal trend, in July *Alternaria* presented much higher prevalence than it showed in May. Moreover, the seasonal variation of *Alternaria* and *Hormodendrum* was correlated positively with allergic cases reacting to these molds (Feinberg *et al.*, 1936). Deamer & Graham (1947) screened the airborne fungal spores in the air of San Francisco for one year (1942-1943) in order to determine the frequency and diversity of mold spores in the atmosphere and to correlate findings with seasonal incidences of respiratory complaints. Both culture and slide methods were used. *Hormiodendroni*, *Penicillium*, *Macrosporium*, *Botrytis*, *Saccharomyces*, *Candida*, *Alternaria*, *Cephalosporium*, *Monilia*, *Mucor*, *Spirotrichium*, *Spicaria* and *Rhizopus* were the predominant spores during the 12 month in San Francisco, but they didn't show any seasonal trend which upset the correlation of mold incidence related to respiratory diseases .

In Britain, Richards (1952) studied the census of airborne fungal spores in the air over Britain using gravimetric methodology. *Cladosporium* was the predominant in overall areas, and displayed a seasonal pattern between June to September. Other genera such as *Penicillium* and *Aspergillus* were prevalent in the center of London, while *Cladosporium*, *Epicoccum*, *Botrytis*, *Alternaria* and *Pullularia*, *Pullularia* were more abundant in the rural places. Di Menna (1955) conducted a quantitative study between March 1953- April 1954 to evaluate the population of airborne fungal spores in Dunedin- New Zealand; using plate exposure method. Results showed the predominance of *Cladosporium* with 42.9 %, followed by *Penicillium* with occurrence pattern of 34.2 %. Yeast also displayed high

frequency. In terms of colonies yield, the maximum was during summer, whereas the minimum was during winter. In Hong Kong, Turner, (1966) carried out a study to screen the fungal spores in the atmosphere using gravimetric method. He demonstrated that *Cladosporium* was the overwhelming predominant genera, followed by *Penicillium*, *Aureobasidium*, and *Aspergillus* respectively. During winter and drier seasons, higher number of colonies was counted. In contrast, in summer and more humid months fewer colonies were obtained (Turner, 1966). In USA and by using volumetric sampler, the influence of weather conditions on the concentrations of atmospheric airspora was examined. It was demonstrated that, wind speed had significant effects on the dispersal rate of spores. Changes in relative humidity levels were found to affect fungal spore release of different fungal phyla. Rainfall affected positively the release and dispersal of Basidiomycetes (Lyon *et al.*, 1984).

In Melbourne, Australia, Mitakakis & Guest (2001) established a fungal spore calendar. Using volumetric means, they identified twenty nine genera and five spore groups. The most abundant fungal spores were noticed for *Cladosporium* with 41.7 % of overall fungal spore distribution. In Poland, Grinn-Gofroń & Bosiacka (2015) conducted a study of four years period to assess the effect of meteorological factors on the airspora composition in the atmosphere. They concluded that, temperature had the greatest impact on fungal airspora composition. The second most significant meteorological factor was the dew point, followed by relative humidity and wind speed respectively (Grinn-Gofroń & Bosiacka 2015).

A study was conducted for one year, using volumetric sampler, to screen the population of airborne fungi in the atmosphere of Beijing, China and to detect the urbanization effect on their distribution and concentration. Air borne fungal spores varied seasonally; during summer and autumn their concentrations were significantly high, whereas in spring and winter they become lower. *Cladosporium* spores were the most dominant fungal group, followed by *Alternaria*, *Pencillium* and *Asperigillus*. The area with more vegetation coverage had significantly higher fungal concentrations compared to densely urban and high traffic sites (Fang *et al.*, 2005).

Also in China, the seasonal dynamics of fungal airspora was detected in different open caves using volumetric sampler. Airborne fungal spores showed a seasonal distribution pattern in which greater concentrations were reported in summer and autumn. *Cladosporium* was the most prevalent spores, followed by *Penicillium*, *Alternaria* and *Aspergillus*. Temperature had the highest impact on the concentrations of ambient fungal spores, but the relative humidity and rain were inversely correlated to them (Wang *et al.*, 2010).

The above mentioned research studies represent just part of heavy literatures conducted to achieve main objectives like: to study the dynamics of fungal populations, to detect correlations between airborne fungal spores and the ambient environment and to construct fungal spore calendars in several countries all over the world.

The significance of almost all aeromycological studies is mainly to provide baseline data about fungal profiles and their fluctuations due to abiotic interactions in order to support preventive measures of respiratory diseases due to fungal allergy.

2.5. Middle East Aeromycological Studies

Aeromycological studies in the Middle East are not established as strong as in other regions; they are still few and dispersed. Abdel-Hafez (1984) studied the prevalence of air mycoflora at Taif region, Saudi Arabia, for one year period, August 1981 to July 1982, using gravimetric method. Two different media, glucose- and cellulose-Czapek's agar, were used in the investigation as well. A total of 25 genera, 58 species and one variety were identified. The colonies number represented great seasonal variations with maximum concentration during winter and minimum in summer season. Moreover; the recovery of *Aspergillus Alternaria*, *Cladosporium*, *Drechslera*, *Scopulariopsis* and *Phoma* spores on the two different media was greatly fluctuated (Abdel-Hafez, 1984). More specifically, Hasnain *et al.*, (2005) monitored the seasonal and diurnal fluctuations of airborne basidiospores throughout three coastal discrete sites in Saudi Arabia using volumetric method. One site out of the three represented the maximum concentration of basidiomycetous spores with taking into consideration that smuts spores were the most significant constitute. The findings didn't show any seasonal pattern related to smuts and rusts spore dispersion. Nevertheless, diurnal variations of basidiomycetous spores exhibited a nocturnal pattern in one sit only. Accordingly, not only seasonal and diurnal factors affected the dynamics of air borne basidiospores, but also regional parameter had significant impact (Hasnain *et al.*, 2005).

In Kuwait, Halawagy (1989) monitored seasonal variations of fungal airspora in three different areas in Kuwait during the five years period 1977-1982 using volumetric samplers. 37 genera were introduced with the most common fungal spores of

Cladosporium Ustilago Alternaria, Drechslera and Chaetomium. Cladosporium was the most abundant among the others. Results showed two peaks of concentrations in spring and autumn while the concentrations decreased significantly twice during summer and winter in all sites. The highest seasonal distribution of airborne molds was observed in 1978, which followed by precipitation and subsequently vegetation growth (Halawagy, 1989). Another aeromycological study was carried out by Khan *et al.* (1999) in Kuwait using volumetric method for one year to screen allergenic fungi in the outdoor and indoor environment. It was demonstrated that, *Asperigillus* spp. were the predominant in outdoor environment, while *Cladosporium* spp. constituted the majority of indoor airspora. In the study, 25 genera were identified as both outdoors and indoors. By comparison, *Aspergillus*, *Alternaria* and *Fusarium* were greatly higher in outdoor environment, whereas *Cladosporium*, *Penicillium*, and *Bipolar* is represented higher prevalence in indoor environment (Khan *et al.*, 1999).

In Qatar, only a single aeromycological study was conducted by Al-Subai (2002) in the period between March 1997 and March 1998 to intensively investigate the effect of seasonal and diurnal variations, and wind velocity on the dynamics of airborne fungal spore's population in the air of Doha. Gravimetric methodology was used for sampling. 35 genera and 73 species were identified. *Cladosporium*, *Alternaria* and *Ulocladium* were the most abundant genera in the atmosphere of Doha. Summer months, May and July, revealed the highest incidence of fungal airspora. Wind was positively correlated to the number of fungal colonies. Once more, *Cladosporinm* and *Alternaria* were the predominant concerning diurnal variations effect (Al-Subai, 2002).

In Palestine, Barkai-Golan (1957) studied the distribution of airborne fungal spores in the atmosphere using plate's exposure method. In Tel-Aviv area, fungal airospora represented seasonal pattern in which their concentration reached the maximum in summer and autumn; while in winter, it was the minimum. In addition to that the author recognized a strong relationship between colonies number and wind velocity, the higher the wind speed, the greater the colonies number. The most frequently fungal spores were *Hormodendrum*, *Penicillium*, *Aspergillus* and *Alternaria* (Barkai-Golan, 1957). In 1962, Barkai-Golan & Glazer studied airborne fungal spore concentration at two different areas in Palestine, Tel-Hashomer and Eilat. By comparison, the concentration of fungal airospora was much higher in Tel-Hashomer than in Eilat. However, the fungal distribution in both areas represented seasonal variations in which the lower colonies count was recorded during winter and markedly increased during other seasons; moreover, five fungal genera represented the main airospora in both sites: *Cladosporium*, *Alternaria*, *Prnicillium*, *Aspergillus*, and *Stemphylium* with *Cladosporium* as the most dominant genera, followed by *Alternaria* (Barkai-Golan & Glazer, 1962). During the years 1993-1995 and in three cities along the Palestine coastal line, the concentration of airborne fungal spores was monitored by Waisel *et al.*, (1997) using volumetric sampler method. They identified fourteen different genera by which *Cladosporium* and *Alternaria* were the predominant fungal airospora. Minor monthly variation in atmospheric mold spores was realized among the three cities (Waisel *et al.*, 1997).

In Jordan, Shaheen (1992) carried out an aeromycological study during the years 1987-1988, for the first time in Amman, to monitor the concentration and distribution of fungal

aerosol and to initiate a fungal spore calendar for the first time in Jordan. Using volumetric method, the author was able to quantify thirty eight genera related to three classes. Between January and May, ascomycetes were most frequent; basidiospores were more common during April and August, while deuteromycetes were highly dominant in October. In terms of fungal genera fluctuation, the maximum incidence of *Botrytis* airspora was recorded in January, *Cladosporium* and *Alternaria* were greatly abundant in October. Furthermore, *Cladosporium* was found to be the main constitute of airspora in the atmosphere (Shaheen, 1992). About 16 years later, another aeromycological study was conducted by Abu-Dieyeh *et al.*, (2010) from January 2008 to January 2009 to screen the concentration of airborne fungal spore in relation to seasonal variation of Zarqa area, Jordan. They deduced from 170 settle plate exposure that, *Cladosporium* was the most prevalent airspora, followed by *Fusarium*, *Alternaria*, *Ulocladium*, *Penicillium* and then *Aspergillus*. Various airborne fungi peaked in different months. For instance; *Cladosporium* peaked in October whereas *Aspergillus* and *Penicillium* showed a peak in September. Also, diurnal variations were examined during two separated seasons. *Cladosporium*, *Fusarium* and *Alternaria*, favored their release and dispersion into the atmosphere at 15:00 h during winter, however there were no differences in their release time during spring months (Abu-Dieyeh *et al.*, 2010). Another research study was carried out using volumetric methods (spore trapping) by Abu-Dieyeh & Barham (2014) to study the seasonal and intradermal variations affecting the concentration of atmospheric mould spora in Zarqa area, Jordan. 41 genera were recorded; yet again *Cladosporium* was the most abundant fungus among others. The dispersal of fungal airspora was positively correlated with air temperature and rain except for

Alternaria, and negatively with relative humidity. Regarding the intradermal variations the release and dispersal of fungal spores favored period 20.00– 04.00 h period. Even though, the two previous studies were conducted at the same area and almost during the same time, the authors concluded that the findings of the two methodologies might be complementary rather than comparable (Abu-Dieyeh & Barham 2014).

Aeromycological studies were more frequently conducted in Egypt rather than other countries, especially during the last century and to a lesser extent in current century. Abdel-Hafez & El-Said, (1989) studied the quantity and quality of airborne fungal spores in the air of Wadi Qena, Egypt. To identify 31 genera, 83 species and 2 varieties of fungal airspora, they utilized gravimetric method. The most prevalent fungal genera were *Alternaria*, *Aspergillus*, *Cladosporium*, *Curvularia*, *Drechslera*, *Epicoccum*, *Penicillium*, *Stemphylium* and *Ulocladium*. The total counts of previous airborne fungi showed fluctuated pattern along different seasons. Also they examined the survival ability (osmophilic and halophilic ability) of certain genera based on different media types and nutrients concentrations. *Aspergillirs* and *Eurotium* members presented great ability to grow and survive on media of high osmotic potential (Abdel-Hafez & El-Said, 1989). In Assiut, the seasonal fluctuations of fungal airspora in the atmosphere and sediment dust were investigated for two years (January 1985 -December 1986) (Abdel-Hafez *et al.*, 1993). 32 genera and 89 species were recorded from the air using settle plate-method, whereas 33 genera and 92 species were detected using the dilution plate-method related to sediment dust sample collecting method. The maximum counts of airborne fungi were observed during April and December 1985, while the highest concentrations of fungal

spores revealed from sediment dust was recorded in February and November 1986. *Alternaria*, *Aspergillus*, *Cochliobolus*, *Penicillium*, *Pleospora* and *Ulocladitrrn* were the most common mycoflora genera (Abdel-Hafez *et al.*, 1993). Hameed *et al.* (2009) aimed from their study to investigate diurnal distribution of air microflora in Helwan, Egypt during the period from March 2006 to February 2007. They used plate exposure method; they revealed that *Aspergillus*, *Alternaria*, *Cladosporium* and *Penicillium* were the most predominant genera. During winter season, the highest concentration of fungal airspora took place 2 hours earlier than other seasons. At 10:00 hours and 20:00 hours, *Asperigillus* showed two peaks. The overall concentration peaks of total fungal airspora regularly occurred at 20:00 hours (Hameed *et al.*, 2009). An aeromycological study was conducted over a period of one year (2008-2009) to evaluate the airborne fungi concentrations and their seasonal fluctuations in indoor and outdoor environment of urban and rural homes in Egypt. Two-stage viable impactor samplers were used to collect samples. The total count of fungi was significantly greater in the rural site than in the urban site. In both, outdoor and indoor the fungal population presented seasonal variations. The highest fungal spore's prevalence was observed during autumn and spring. Similarly, in indoor and outdoor, *Alternaria*, *Aspergillus*, *Cladosporium*, *Penicillium* and yeasts were the most dominant genera (Awad *et al.*, 2013). Very close to Arab countries, in Turkey, Çolakoglu (1996) monitored the concentration of air borne fungal spores at the Anatolia quarter of Istanbul using volumetric method during the period from January 1994 - December 1994. The most common fungal airspora were *Cladosporium*, *Penicillium*, *Ustilago*, *Aspergillus*, *Alternaria*, *Botrytis*, *Leptosphaeria*, *Rhizopus*, *Helminthosporium* and

basidiospores. *Cladosporium* was the most abundant genera with maximum prevalence during windy weathers, then followed by *Penicillium*. They presented seasonal variations in which their maximum occurrence in summer season (Çolakoglu, 1996). In Eskisehir (Turkey), a study was conducted to identify airborne fungi in three different urban sites using culture method. The most frequent and predominant fungal genera were *Alternaria alternata*, *Cladosporium cladosporioides* and *Scopulariopsis brevicaulis*. The highest density of fungal spores was recorded in September, May and November, while the lowest observed concentration occurred in April, March and June (Asan *et al.*, 2004).

The relationships between *Alternaria* and *Cladosporium* with meteorological conditions were investigated in Eskisehir City, Turkey throughout 2005 to 2006. Samples were collected using Durham gravimetric sampler throughout 3 different sites. *Cladosporium* spores count was as two third as *Alternaria* spores. For both genera, their ultimate distribution occurred in summer while the lowest was recorded in winter. Precipitation, temperature and wind velocity didn't represent significant impact on the fungal spore's concentration according to the sites; however, relative humidity had a significant effect on the dispersal rate of *Cladosporium* and *Alternaria* spores together (Erkara *et al.*, 2009).

In Iran, Nourian *et al.* (2007) carried out a study to monitor fungal airspora in the atmosphere of Zanzan-Iran. To achieve their objective, gravimetric method was used. The most predominant fungal spores in the air were yeast species, *Cladosporium*, *Penicillium*, *Aspergillus* and *Alternaria*. They presented seasonal variations in their distributions. Colonies count was the greatest in winter while the lowest was during summer season; yet, *Cladosporium* was the most abundant during all seasons. Negative correlation was detected

between airborne fungal spores and temperature while they positively linked to relative humidity (Nourian *et al.*, 2007). Also in Iran, the mycoflora in the air of Tehran was screened using gravitational settle plate method. The overall findings determined that the most common fungal spores in the air were of *Aspergillus*, *Cladosporium*, *Penicillium* and *Alternaria* species. *Asperigillus* was the predominant airborne fungal spores (Shams-Ghahfarokhi *et al.*, 2014).

Obviously, the variations in the concentration, composition and distribution of aeromycoflora of different countries in the Middle East were significantly influenced by the meteorological parameters, geographical and physical features. However, some fungal genera are common between those countries such *Cladosporium*, *Alternaria* *Aspergillus* and *Penicillium*. In Gulf region, only few aeromycological studies were conducted and many have been done since a long time and they may not reflect the present changes in the airborne fungal populations related to their biotic and abiotic surrounding features in those countries. Qatar is not an exception; the only study about fungal airospora in the air of Doha was conducted since 15 years that make the need for this present study necessary to update the knowledge about the seasonal and diurnal variations in airborne fungal spores of Doha area.

The main objectives of the present study are:

- 1- Provide baseline knowledge about density, diversity and dynamics of airborne fungal spore in the atmosphere of Doha using settle plate exposures. This knowledge is necessary for mycologists, applied ecologists, plant pathologists and other disciplines of biological, environmental and health sciences.

- 2- Investigate variations in seasonal and diurnal distribution of airborne fungal spores, abundance, density and diversity and to correlate these variations with meteorological factors.
- 3- Investigate variations in diurnal distribution of airborne fungal spores, abundance, density and diversity during February and March (2016).
- 4- Correlate the variations in species composition and abundance of air-borne fungi with atmospheric CO₂ concentration in two different areas in Qatar during February and March (2016) in order to investigate the impact of urbanization on air-borne fungal spore population.
- 5- Compare results obtained from two different cultural selective media: Potato Dextrose Agar (PDA) and Czapek's/ Rose-bengal media during February and March 2016.

CHAPTER THREE

MATERIALS AND METHODS

3.1. Study site

Qatar is a peninsula occupies an area of more than 11,000 Km² and a coastline of 900 km in length. It lies between latitude 24°27' and 26°10' and longitude 50°45' and 51°40'. of the Arabian Peninsula and connected to Saudi Arabia in the south, bordered by a very shallow, semi-enclosed sea characterized by hyper-salinity, salinity is ranging between 39 practical salinity unit (psu) and 41 practical salinity unit (psu) at the surface, and is 1-2psu higher at the bottom (Richer,2009). Geographically, it is a flat, rocky and arid desert and in the south sand dunes are the predominant features. As a subtropical desert, Qatar is hot and has a dry weather, the annual rainfall is about 81 mm, and the average highest air temperature is 31°C although the maximum temperature can be beyond 47 °C (Brook *et al.*, 2006). Doha is the capital of the state, located on the eastern Qatari coastal line at 25° 17' 12" N, 51° 32' 0" E. It is the most urbanized and populous city in the state, the number of population within the country until February-2015 is 2,116,400 (Ministry of Development Planning and Statistic's, Qatar 2015). Doha has been flourishing rapidly; it is the commercial and economic center of the state as well.

The present study was carried out at Qatar University which is located on the northern side of the capital Doha at 25.3747° N, 51.4903° E. The main habitat and associated vegetation in northern part of Qatar is characterized by 'rodat' areas in which compact soils are common. This type of soils are usually support more moisture and organic matters than

other desert parts. Vegetation type is usually ranges from trees, shrubs to grasses and herbs, commonly *Acacia* spp., *Prosopis juliflora*, *Ziziphus nummularia* and *Lycium shawii*. Because this area accommodates larger rodents, they have been utilized as farms for growing crops (Norton *et al.*, 2009). Within the campus of University of Qatar, naturally occurring plants are widespread. In addition, many species associated with man-made and man-influenced sites such as gardens, green houses and roadside are very common.

3.2. Seasonal variations in air-borne fungal populations in Doha city

A gravimetric method (settle plate exposure) using a Petri dish was carried out directly on the roof of the building of Qatar University (about 12 m above ground level). One Petri dish (9 cm diameter) containing Potato Dextrose Agar (PDA) (HiMedia Laboratories Pvt.Ltd, India) was exposed on the roof of the above mentioned building for 15 minutes, which result in more accurate colony counts, at 3:00 pm on each of 3 days a week (alternate days) from the beginning of April 2015 to the end of March 2016. A total exposure of (106) were collected during the year of study. After sampling, the plates were incubated at 25 °C for 3-5 days. The fungal grown colonies for each species were counted and recorded. For classification, fungal colonies were isolated and purified on Potato Dextrose Agar(PDA) and other selective agars like Malt Extract Agar (MEA) (HiMedia Laboratories Pvt.Ltd, India), then they were incubated for 3-5 days. Identification was based on the macro- and microscopic features following the keys and description given by many authors like: Booth

(1971), Domsch *et al.* (1980), Kozakiewicz (1989), Moubasher (1993), Pitt & Hocking (1997), Barnett & Hunter (1999), Samson *et al.* (2002) and Lacey & West (2007).

Daily meteorological data from Qatar University weather station were supplied by the Weather Record Department, Doha, Qatar. The meteorological data considered are: minimum and maximum daily temperature, minimum and maximum relative humidity, daily rainfall, wind direction and wind speed.

3.3. Intra-diurnal variations in air-borne fungal populations in Doha city

February and March months (2016) were chosen to investigate intra-diurnal variations in fungal populations of Doha mainly because these are the months that expect to have the greatest diversity in species composition. One Petri dish (9 cm diameter) containing (PDA) was exposed for 15 min at 6 hours interval (at 6:00 am; 12:00 pm; 6:00 pm and 12:00 am) during the period of 1st of February and 31 of March 2016. After sampling, the plates were incubated at 25 °C for 3-5 days. The fungal grown colonies for each species were counted and the total colony counts were recorded. For classification, fungal colonies were isolated and purified on (PDA) and other selective agars like (MEA), and then they were incubated for 3-5 days. Then, the above mentioned books and publications (section 3.2) were used for identification of fungal species.

During the whole period a data logger (OMEGA Engineering, INC., USA) was installed at the site of collection to monitor temperature and relative humidity in hourly time basis.

3.4. Effect of culture media used in sampling on species composition and abundance of fungal spore populations

To differentiate between the potentiality of fungal growth on two different cultural media, two Petri dishes (9 cm diameter) containing (PDA) of the following compositing (Infusion from potato 200 g/l, dextrose 20 g/l and Agar 15 g/l) and Czapek's/Rose-Bengal agar with the following ingredients(Mycological peptone 5 g/l, Dextrose 10 g/l, Monopotassium phosphate 1 g/l, Magnesium sulphate 0.5 g/l, Chloramphenicol 0.1 g/l, Rose Bengal 0.05 g/l and Agar 15 g/l) (HiMedia Laboratories Pvt.Ltd, India) were exposed to the air for 15 minutes for three days weekly (alternate days) during the period of 1st of February and 31 of March 2016. After sampling, the plates were incubated at 25 °C for 3-5 days. The fungal grown colonies for each species were counted and the total colony counts were recorded. For classification, fungal colonies were isolated and purified on (PDA) and other selective agars like (MEA), and then they were incubated for 3-5 days. Then, the above mentioned books and publications (section 3.2) were used for identification of fungal species.

3.5. Effect of urbanization and CO₂ concentration on species composition and abundance of fungal spore populations

Two locations at Doha city were chosen to investigate the impact of urbanization on species composition and abundance of fungal spore populations. Qatar University is one of these locations as relatively far from industrial areas and the other is close to the central industrial

area of Doha (Industrial Area). Industrial area is a district of Doha, Qatar of coordinate 25°10'3"N 51°26'22"E. It is mainly occupied with a working population as the main factories such as cement factories, auto repair garage and other industrial business are located there, where from a pre-investigation for three continuous days approved to have more CO₂ concentration than Qatar University Campus.

During the period of 1st of February and 31 of March 2016, Petri dishes (9 cm diameter) containing (PDA) were exposed to the air for 15 minutes for three days weekly (alternate days). In respect to the two locations and during sampling the plates were opened at the same time and for the same exposure period (15 min.). After sampling, the plates were incubated at 25 °C for 3-5 days. The fungal grown colonies for each species were counted and the total colony counts were recorded. For classification, fungal colonies were isolated and purified on (PDA) and other selective agars like (MEA), and then they were incubated for 3-5 days. Then, the above mentioned books and publications (section 3.2) were used for identification of fungal species.

During sampling and in each location, a portable CO₂/Temperature/RH data logger (CO2METER.COM, USA) was used to record CO₂ concentration, Temperature, and Relative humidity every minute during the collection period (15 min.). The average of the 15 readings was considered for each collection record.

3.6. Statistical and data analysis

Statistical analyses were performed to correlate the mean daily fungal spore concentrations and species composition with the daily data of the meteorological parameter of the same day and for the whole year using Pearson correlation coefficient. Similar correlation analyses were performed for the two months study to investigate the effect of CO₂ on species composition and abundance.

Results of different measured parameters from the two different cultural media as well as results from the two different locations were compared after being subjected to T-test at $P \leq 0.05$. ANOVA was used to detect the significant differences in species diversity and abundance among the four diurnal periods and then means of variables were separated using Tukey test at $P \leq 0.05$ (SigmaStat 4, Systat Software, Inc).

Jaccard similarity coefficient (Jaccard, 1908) was applied to compare the similarities in species composition (including species identified at specific level and other taxa) among the four diurnal periods as follows: Jaccard similarity coefficient = $(c/(a+b)-c)*100$. C= number of common species between any two time periods (a & b).

CHAPTER FOUR

RESULTS

4.1. Seasonal variation of fungal spore populations in the atmosphere of Doha

From the air of Doha, the total colony count number retrieved from 106 exposure samplings during the year of study (1st of Apr 2015 - 31st of March 2016) using gravimetric method was 283 yeast and 1197 mould colony forming units (CFU). The mould colonies were belong to 21 genera and 62 species (Table 1). Due to difficulties in identification, certain mould colonies were only identified to genus level such as *Acremonium*, while others were referred to as unknown. The maximum concentration of airborne fungal spores was recorded in February (2016), whereas the minimum occurred in September (2015). The main constituents of the airborne fungi population in the atmosphere of Doha were attributed to the genera: *Cladosporium* (60.2%), *Aspergillus* (10.4 %) *Fusarium* (9.4 %), *Alternaria* (8.5 %), *Ganoderma* spp.(2.3%) and *Penicillium* (2.0 %) (Figure 1). *Cladosporium* was the most common fungal taxa in the air of Doha compromising around two thirds of the total colony counts and representing with five species. *C. cladosporioides* was the most dominant species with 70.5 % of the total of the genus *Cladosporium*, followed by *C. macrocarpum* with 9.2%, and the third common one was *C. sphaerosprumum* with 6.4%. *Cladosporium* had double peaks in April (2015) and in February (2016) and a trough in July (2015) (Table 1; Figure 3). *Aspergillus* sp. was the second predominant fungal genera and represented with ten species (Table 1). *Aspergillus* achieved the highest concentration in August (2015) and the lowest in January (2016) (Figure 3) *A. flavus* represented by 46.5 % of the total of the genus *Aspergillus*, then *A.*

sydowii with 25.2 % (Table 1; Figure 1). *Fusarium* was ranked as the third among the most common airborne fungi in Doha. Most of recovered colonies of *Fusarium* were identified to genus level, however *F. oxysporum* and *F. clamydosporum* were the most abundant and frequently recorded (Table 1). *Alternaria* represented the fourth most prevalent fungal taxa with seven identified species. The spore's concentration of *A. alternata* put up with more than the half of the whole genus (61.4%), followed by *A. chlamydospora* (12.8%), then *A. infectoria* (11.9%) (Table 1; Figure 1). Both *Alternaria* and *Fusarium* reached their maximum concentrations in July (2015), the density of *Alternaria* spores declined in June (2015), while *Fusarium* colony counts diminished in January (2016) (Figure 3). Interestingly *Ganoderma* spp. was ranked the fifth among the most predominant fungi in the atmosphere of Doha with 2.4 % abundance, even though they exclusively appeared in February and March (2016) of the whole year (Table 1; Figure 1).

The correlation analyses between weather parameters and the incidence of the main fungal taxa, total daily or monthly colony count and species diversity were represented in Table 2. Both daily and monthly counts of either of *Cladosporium* or *Alternaria* spores showed a significant negative correlation ($P \leq 0.05$) with daily maximum, minimum and mean temperatures. While the correlations of *Cladosporium* colony count with both daily or monthly relative humidity and rainfall were not significant, the monthly rainfall data showed a highly significant correlation ($P \leq 0.01$) with *Alternaria* colony counts. Highly significant ($P \leq 0.01$) positive correlation was appeared between daily counts of *Cladosporium* and *Alternaria* and daily wind speed (Table 2). In contrary, *Aspergillus* concentration showed a non-significant correlations with any of the studied weather

parameters (Table 2). *Fusarium* daily colony counts had only a significant correlation ($P \leq 0.01$) with the daily wind speed data (Table 2). Total daily colony counts of all encountered fungi showed significant negative correlations ($P \leq 0.05$) with any of the daily temperature parameters. A highly significant positive correlation ($P \leq 0.01$) was also obtained between the total daily or monthly colony count and daily or monthly wind velocity (Table-2 & Figure 2) and a non-significant correlation with either daily mean relative humidity or rainfall. However no significant correlations were reported between total colony counts and either of relative humidity or rainfall. Daily temperature parameters possessed highly significant ($P \leq 0.01$) negative correlations with fungal species diversity of Doha atmosphere while daily wind speed data possessed - highly significant positive correlation ($P \leq 0.01$) with species diversity (Table 2 & Figure 2). No statistically significant correlations on air-borne fungi had been reported due to wind direction (Table 2). Figure (4) represents the fungal spore calendar for air-borne fungi in the atmosphere of Doha and their seasonal patterns. The highest concentration of fungal colony counts was reported in February (2016), while the lowest was recorded in August (2015). The highest number of fungal species was reported in February (2016), while the lowest was obtained in September (2015). *C. cladosporoides* prevailed in the atmosphere of Doha in relatively very high concentrations (4.5-16 colonies per exposure) in all months. *Ganoderma* occurred only in two months of the whole year with concentration (0.8-1.8 colonies per exposure).

Table 1. Total colony count and occurrence of airborne fungi recovered from the air of Doha during the study period (1st of Apr 2015 - 31st of March 2016) using gravimetric method (a total of 106 exposure samplings).

Fungal Taxa	No. of colonies	^(a)Colony count (%)	^(b)Monthly occurrence	Month of occurrence
<i>Acremonium</i> sp.	3	0.25	1	Dec
<i>Acrophialophora</i>	4	0.33	1	Jun
<i>Acrophialophora fusispora</i> (S.B. Saksena) Samson	4	0.33	1	Jun
<i>Agaricales</i> sp.	2	0.17	1	Mar
<i>Alternaria</i>	102	8.52	11	Entire year except June
<i>Alternaria alternata</i> (FR.) Keissler	62	5.18	11	Entire year except June
<i>Alternaria brassicicola</i> (Schwein.) Wiltshire	3	0.25	1	Feb
<i>Alternaria chlamydospora</i> Mouchacca	13	1.09	3	Dec,Jan,Feb
<i>Alternaria infectoria</i> E.G. Simmons	12	1.00	1	Mar
<i>Alternaria phragmospora</i> V. Emden	2	0.17	2	Feb,Apr
<i>Alternaria porri</i> (Ellis) Cif.	4	0.33	1	Dec
<i>Alternaria tenuissima</i> (Kunze) Wilt.	6	0.50	2	May,Jul
<i>Aspergillus</i>	124	10.36	12	Entire year
<i>Aspergillus flavus</i> Link	59	4.93	11	Entire year except Jan
<i>Aspergillus fumigatus</i> Fresenius	2	0.17	1	Nov
<i>Aspergillus melleus</i> Yukawa	3	0.25	1	Dec
<i>Aspergillus niger</i> (V. Tiehem) Blochw	9	0.75	3	Jul,Aug,Sep
<i>Aspergillus sulphureus</i> (Fres.) Thom & Church	2	0.17	1	Mar
<i>Aspergillus sydowii</i> (Bainier & Sartory) Thom & Church	32	2.67	4	Dec,Jan,Feb,Mar
<i>Aspergillus terreus</i> Thom	3	0.25	2	Apr,May
<i>Aspergillus versicolor</i> (Vuill.) Tiraboschi	3	0.25	1	Oct
<i>Aspergillus carbonarius</i> (Bainier) Thom	4	0.33	3	Nov,Jan,Feb
<i>Aspergillus japonicas</i> Saito	2	0.17	1	Sep
<i>Aspergillus ustus</i> (Bain.) Thom & Church	5	0.42	2	Jul,Feb

<i>Blastomyces</i> sp.	2	0.17	1	Dec
<i>Cladosporium</i>	721	60.23	12	Entire year
<i>Cladosporium cladosporioides</i> (Fres) de Vries	509	42.52	12	Entire year
<i>Cladosporium herbarum</i> (Pers.) Link	4	0.33	3	Oct,Dec,Feb
<i>Cladosporium macrocarpum</i> Preuss	66	5.51	9	Entire year except Jun, Aug and Sep
<i>Cladosporium oxysporum</i> Berk. & M.A. Curtis	13	1.09	4	Feb,Mar,Apr,May
<i>Cladosporium</i> sp.	80	6.68	1	Apr
<i>Cladosporium sphaerosprumum</i> Penzig	46	3.84	11	Entire year except Nov
<i>Cladosporium tenuissimum</i> Cooke	3	0.25	3	Nov, Feb, Mar Apr, Jun, Jul, Aug, Sep, Oct, Dec
<i>Cochliobolus</i>	18	1.50	8	
<i>Cochliobolus australiensis</i> (Tsuda & Ueyama) Alcorn	6	0.50	4	Jun, Jul, Sep
<i>Cochliobolus hawaiiensis</i> Alcorn (anamorph)	3	0.25	1	Aug
<i>Cochliobolus lunatus</i> Nelson & Haasis (anamorph)	4	0.33	1	Apr
<i>Cochliobolus spicifer</i> Nelson (anamorph)	5	0.42	2	Oct, Dec
<i>Epicoccum</i>	2	0.17	2	May, Jun
<i>Epicoccum nigrum</i>	2	0.17	2	May, Jun
<i>Fusarium</i>	113	9.44	12	Entire year
<i>Fusarium</i> sp.	76	6.35	8	Jan, Feb, Mar, Apr, May, Oct, Nov
<i>Fusarium clamydosporum</i> Wollenweber & Reinking	10	0.84	5	Nov, Dec, Jan, Feb, M ar
<i>Fusarium dimerum</i> Penzig	8	0.67	5	Apr, May, Dec, Feb, Mar
<i>Fusarium moriliforme</i> (A. Braun) Wollenweber	3	0.25	1	Oct
<i>Fusarium oxysporum</i> Schlecht.	16	1.34	6	Oct, Nov, Dec, Feb, Mar, Apr,
<i>Ganoderma</i> spp.	28	2.34	2	Feb, Mar
<i>Geotrichum</i>	2	0.17	1	Jun
<i>Geotrichum candidum</i> Link	2	0.17	1	Jun
<i>Mucor</i> sp.	2	0.17	1	Mar
<i>Myrothecium</i>	2	0.17	1	Dec
<i>Myrothecium verrucaria</i> (Alb. & Schwein) Ditmar	2	0.17	1	Dec
		2.01		

<i>Penicillium</i>	24		4	Jan, Feb, Mar, May, Jun
<i>Penicillium</i> sp.	3	0.25	1	Feb
<i>Penicillium brevicapmactum</i> Dierckx	2	0.17	1	Mar
<i>Penicillium canescens</i> Sopp	3	0.25	1	Feb
<i>Penicillium citrinum</i> Thom	10	0.84	1	May
<i>Penicillium italicum</i> Wehmer	4	0.33	2	May, Jun
<i>Penicillium steckii</i> K.M. Zalesky	2	0.17	1	Jan
<i>Phoma</i>	4	0.33	1	Mar
<i>Phoma glomerata</i> (Corda) Wollenw.	4	0.33	1	Mar
<i>Pleospora</i>	7	0.58	3	Apr, May, DEC
<i>Pleospora tarda</i> Simmons	7	0.58	3	Apr, May, DEC
<i>Rhizopus</i>	5	0.42	2	Jan, Mar
<i>Rhizopus oryzae</i> Went & Prinsen-Geerligs	2	0.17	1	Jan
<i>Rhizopus stolonifer</i> (Ehrenb.) Lind	3	0.25	2	Mar
<i>Stachybotrys</i>	6	0.50	2	Dec, Mar
<i>Stachybotrys chartarum</i> (Ehrenb.) Hughes	3	0.25	1	Dec
<i>Stachybotrys elegans</i> (Pidopl.) W. Gams	3	0.25	1	Mar
<i>Thanatephorus</i>	2	0.17	1	Jan
<i>Thanatephorus cucumeris</i> (Frank) Donk	2	0.17	1	Jan
<i>Ulocladium</i>	15	1.25	4	
<i>Ulocladium botrytis</i> Preuss	9	0.75	2	Mar, Jul
<i>Ulocladium chartarum</i> (Preuss) Simmons	6	0.50	2	Mar, May
UNKNOWN	9	0.75	3	Jan, Mar, May
MOULDS	1197	80.88	12	Entire year
YEASTS	283	19.12	12	Entire year
Total No. of Fungal colonies			1480	

^(a) calculated as a percentage of the total count of mold colonies recovered from the entire study (i.e a total of 1180 colonies from 106 exposures = days).

^(b) number of months of occurrence out of 12 months

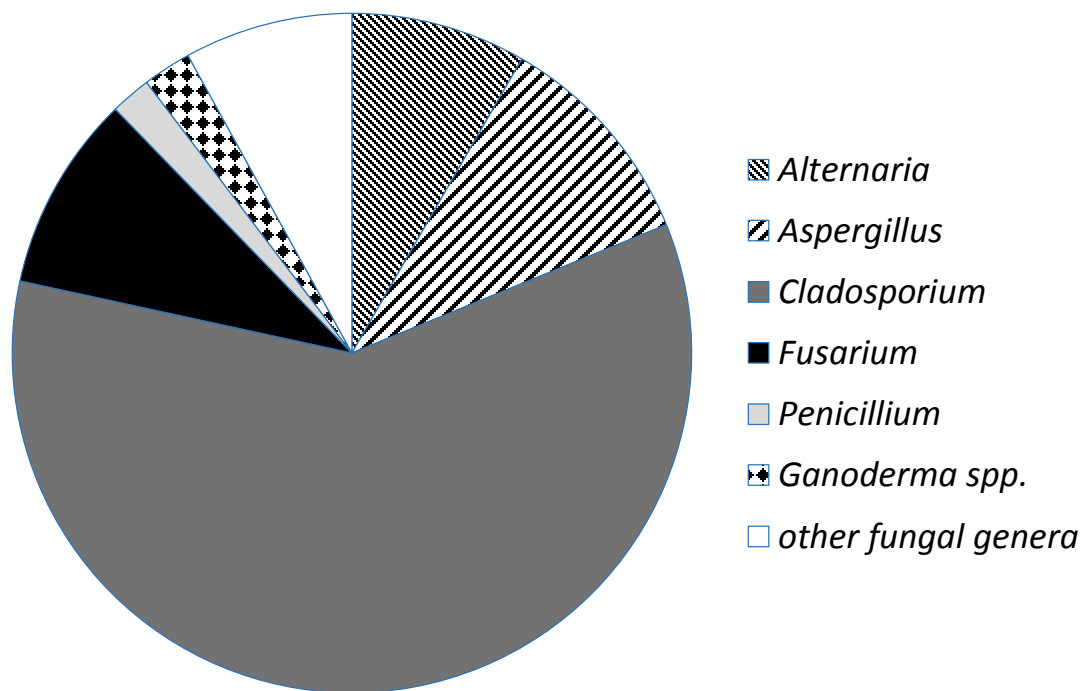


Figure 1. The composition of fungal constituents (%) in the atmosphere of Doha City during (1st of Apr 2015 - 31st of March 2016) using gravimetric method (a total of 106 exposure samplings).

Table 2. Monthly climatic parameters of Doha during the year of study versus results of correlation analysis. The correlation was accomplished based on daily data as well as monthly data and using Pearson correlation coefficient.

	Maximum Temperature (°C)	Minimum Temperature (°C)	Mean Temperature (°C)	Mean Relative Humidity (%)	Total Rainfall (mm)	Mean Wind Speed (m/s)	Mean Wind Direction (degree.)
Apr 2015	35.5	23.6	29.0	27.6	0.0	4.3	255.7=WSW
May	41.0	28.7	34.4	26.0	0.0	4.2	243.3=WSW
Jun	42.2	29.8	35.8	31.7	0.0	4.3	297.1=WNW
Jul	40.7	32.2	36.2	45.0	0.0	3.7	230.0=SW
Aug	41.8	32.1	36.1	58.9	0.0	2.7	214.3=SW
Sep	39.9	30.0	34.3	55.8	0.0	2.5	265.0=W
Oct	36.7	27.2	31.4	59.9	0.0	2.7	308.3=NW
Nov	29.4	21.3	25.0	66.7	0.0	2.4	317.8=NW
Dec	23.7	15.9	19.5	69.7	0.2	3.0	292.5=WNW
Jan 2016	22.9	14.3	18.1	66.3	0.1	3.1	256.3=WSW
Feb	24.0	15.5	19.4	59.5	0.0	2.9	233.1=SW
March	27.6	19.0	23.1	58.3	1.3	3.1	185.7=S
Correlation Coefficients / Daily weather data Vs. Daily data of number of fungal colonies							
<i>Cladosporium</i>	-0.231*	-0.228*	-0.237*	-0.0828	-0.0328	0.427**	0.175
<i>Alternaria</i>	-0.243*	-0.193*	-0.222*	0.0125	-0.0243	0.323**	-0.111
<i>Aspergillus</i>	0.0135	0.0347	0.0209	0.0459	-0.0737	0.0779	0.0909
<i>Fusarium</i>	0.025	0.0906	0.0673	-0.0697	-0.0276	0.329**	0.084
Total daily colony count	-0.23*	-0.215*	-0.231*	-0.102	-0.0519	0.484**	0.155
Total daily number of species	-0.26**	-0.26**	-0.267**	-0.0249	-0.0917	0.257**	0.143
Correlation Coefficients / Monthly weather data Vs. Monthly data of total number of colonies							
<i>Cladosporium</i>	-0.625*	-0.703*	-0.666*	0.14	0.108	0.129	0.267
<i>Alternaria</i>	-0.203**	-0.154*	-0.18**	0.36	0.562**	-0.253	0.569
<i>Aspergillus</i>	-0.724	-0.696	-0.712	0.468	0.77	-0.141	-0.179
<i>Fusarium</i>	0.0846	0.0595	0.0821	-0.217	0.125	0.37	-0.348
Total monthly colony count	-0.684	0.189	0.189	0.189	0.307	0.115	0.181
Total monthly number of species	-0.642	-0.681	-0.66	0.173	0.468	0.022	-0.241

(**) Highly significant correlation at $P \leq 0.01$

(*) Significant correlation at $P \leq 0.05$

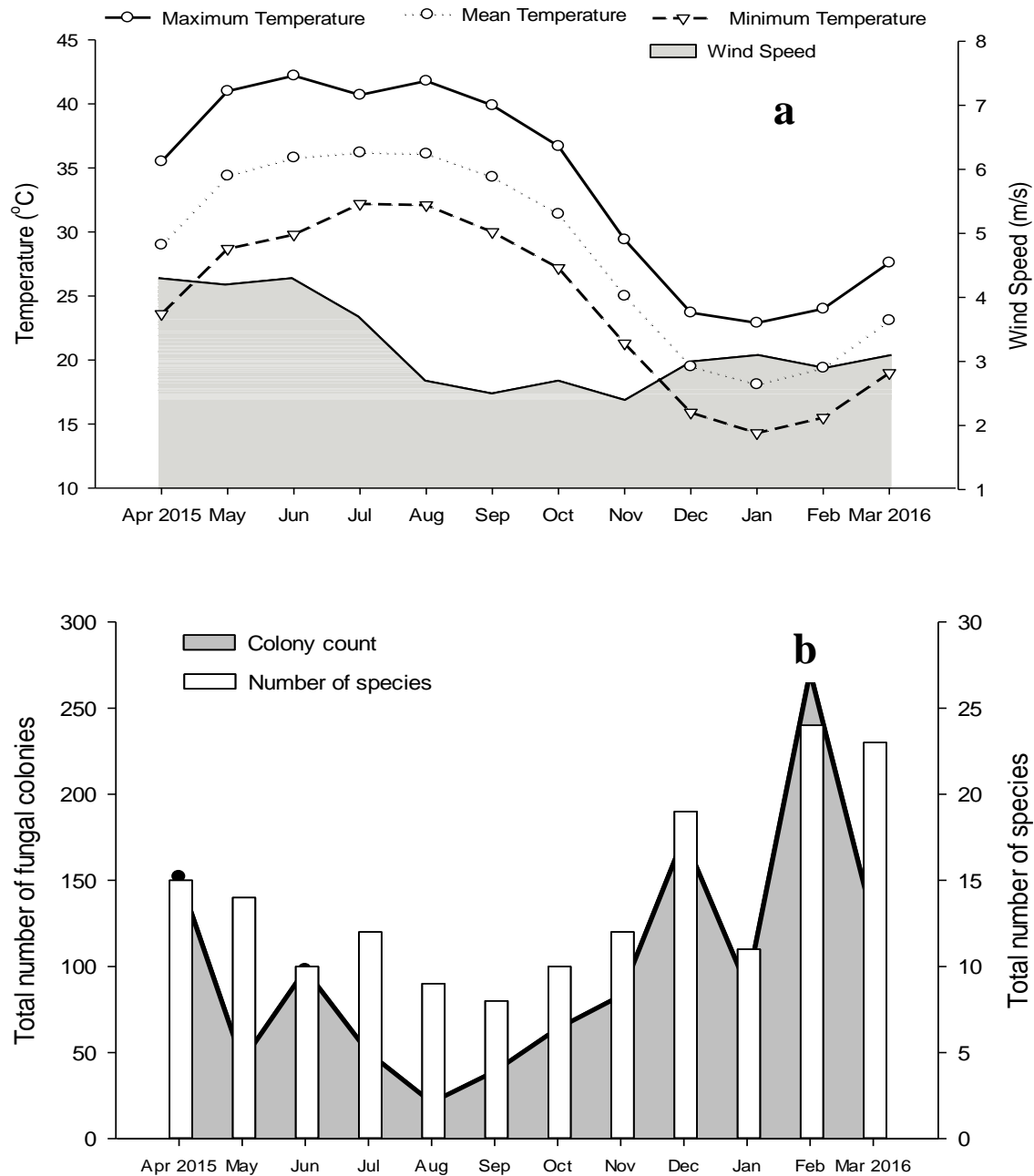


Figure 2. Temperature and wind speed (a) versus total colony count and total number of species (b) recovered each month during the year of study (1st of Apr 2015 - 31st of March 2016) using gravimetric method (a total of 106 exposure samplings). Significant positive correlations were obtained between wind speed and both of colony count and number of species while negative correlations were obtained between any of temperature parameters and both of colony count and species number.

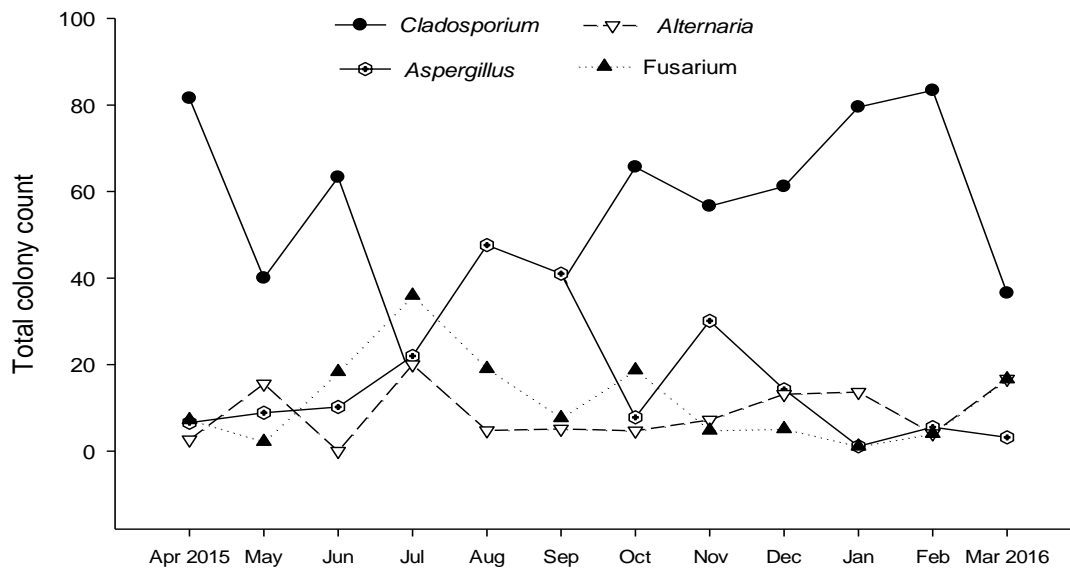
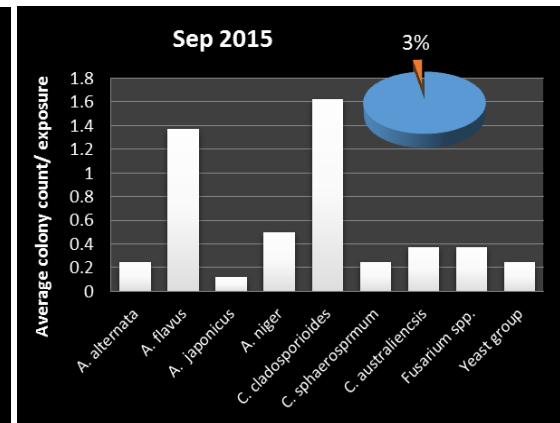
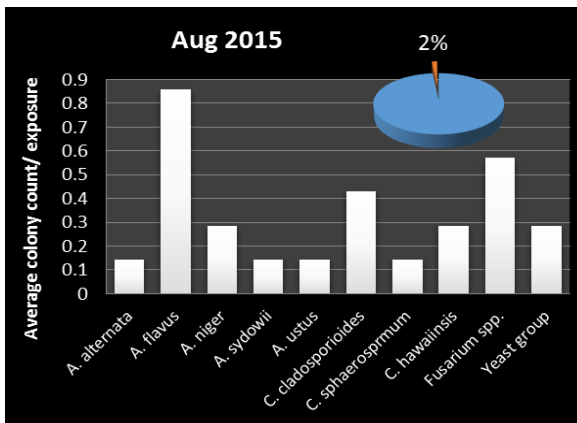
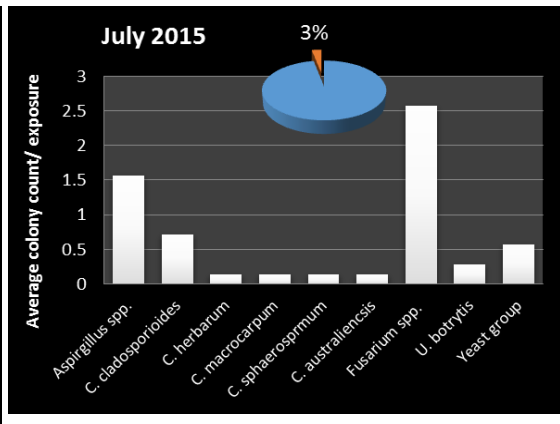
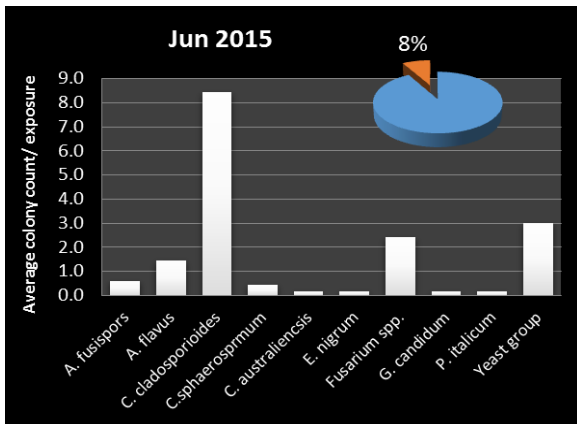
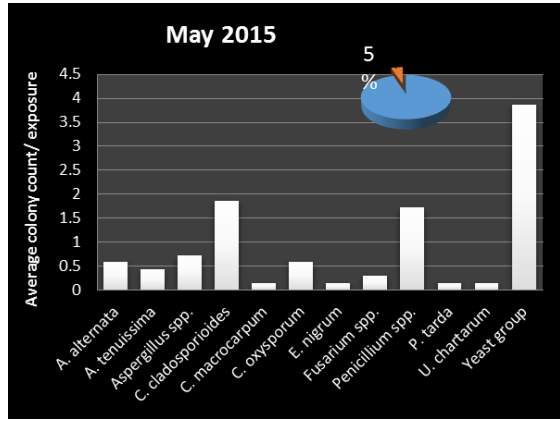
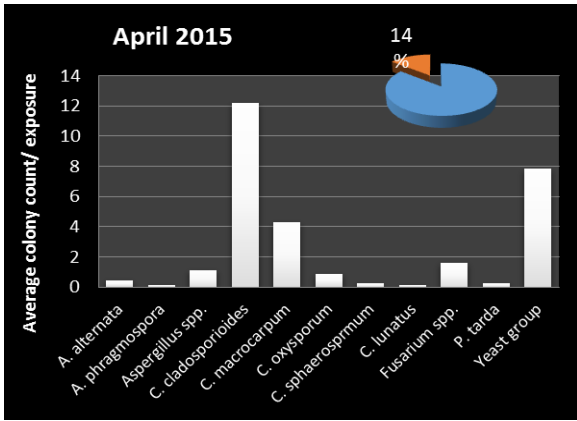


Figure 3. Seasonal variation of the four most common fungal genera in the atmosphere of Doha during the year of study (1st of Apr 2015 - 31st of March 2016) using gravimetric method (a total of 106 exposure samplings).



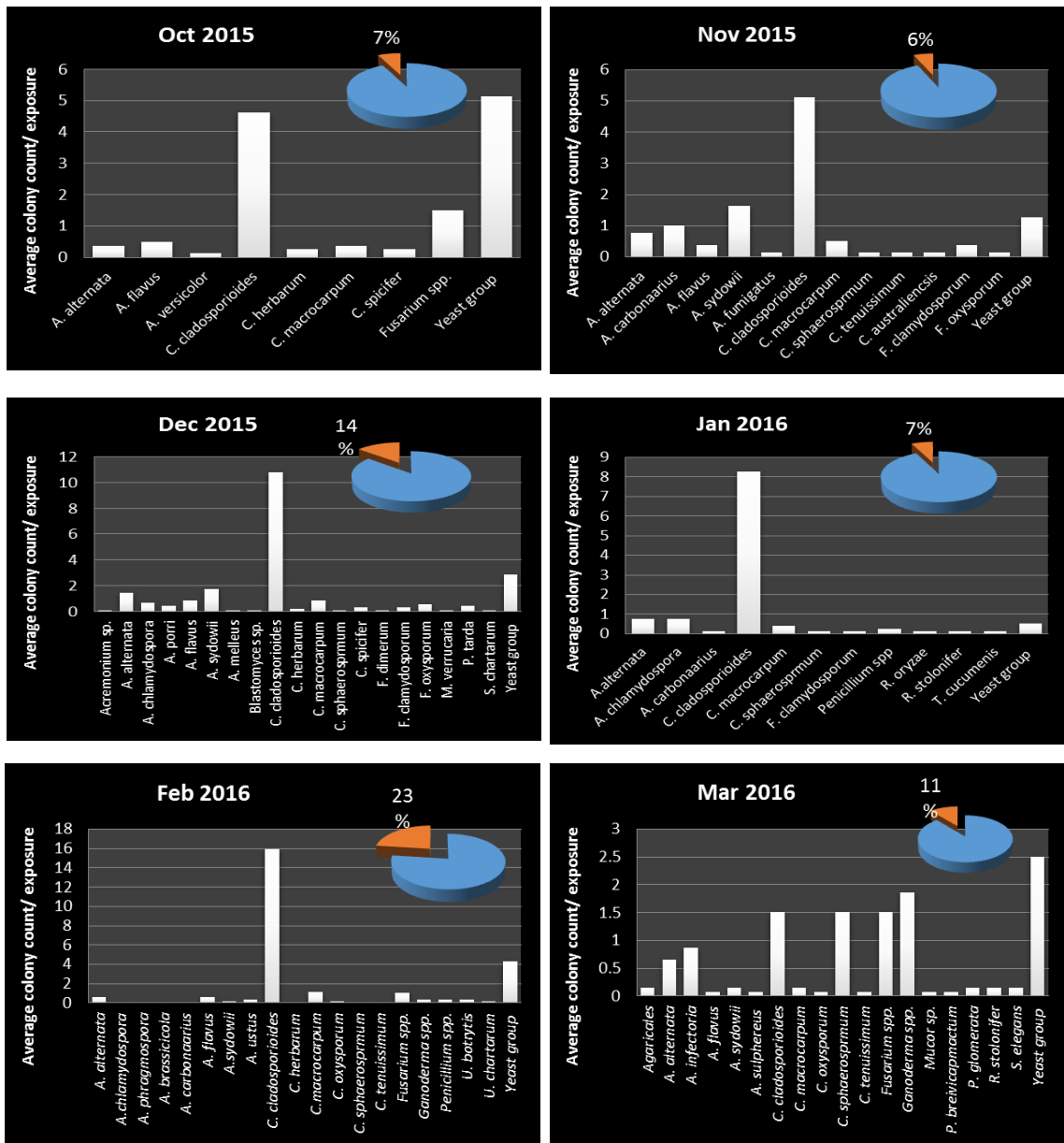


Figure 4. Fungal spore calendar for air-borne fungi in the atmosphere of Doha. The values are based on the average daily colony count of each fungal taxa recovered on PDA culture plates. The Pie graph reflects how much of total fungal colonies (%) reported in a month compared to the summation of all fungal colony counts reported in the whole year. The highest % of fungal colonies occurred during February 2016 (23%), while the lowest occurred in August 2015 (2%).

4.2. Diurnal variation of fungal spore populations in the atmosphere of Doha

Diurnal variations were studied throughout two months (1st of Feb - 31st of March 2016) with a total of 72 exposure samplings. During the two months of the study, the mean daily maximum temperature ranged from 24.6 to 27.3 °C, the mean daily minimum temperature ranged from 15.0 to 18.7 °C. The daily relative humidity ranged from 39 to 84%. Table (3) shows the diurnal variation of fungal species richness and abundance of air-borne fungi in the atmosphere of Doha. Significant differences at $P \leq 0.05$ in colony count and species diversity were found among the four studied time periods. The highest colony count and species diversity were recovered at 18:00 h while the least were reported at 00:00 h (midnight). No significant differences in colony count as well as species diversity occurred between 12:00 and 18:00 h (Table 4). Jaccard similarity index was used to calculate the similarity coefficient among diversity of fungal species recovered from the four time periods. The highest index value reflects the highest similarity in species composition recovered between the two periods. The highest similarity coefficient was obtained between 00:00h and 06:00h, while the lowest similarity coefficient was detected between 18:00 h and 12:00 h (Table 5). The mean daily colony count was negatively correlated ($P \leq 0.05$) with mean daily relative humidity and positively correlated ($P \leq 0.05$) with mean daily temperature (Figure 5). However no significant correlations were found between species diversity and any of weather parameter. Interestingly the mean daily count of the genera *Alternaria* and *Ganoderma* were shown significant positive correlation ($P \leq 0.05$) with the mean daily temperature. The most abundant fungal taxa retrieved throughout the two months at the four time periods were *Cladosporium cladosporioides*, *Alternaria spp.*,

Fusarium spp., *Ganoderma*, *Ulocladium botrytis* and *Aspergillus* (Figure 6). *Cladosporium cladosporioides* represented a mid-day (12:00 h) peak and had a minor peak at 18:00 h, while its concentration significantly declined ($P \leq 0.05$) at midnight (00:00 h) and in the early morning (06:00 h). *Ganoderma* spp. significantly peaked ($P \leq 0.05$) at 18:00 h and shown similar abundance among the other three time periods. *Alternaria* had a significant midday (12:00 h) peak pattern, while the colony count significantly declined in the other three periods. No significant differences in *Fusarium* colony count were reported among the four studied time periods while *Aspergillus* had a major peak pattern at 18:00 h and *Ulocladium botrytis* at midnight (00:00 h) (Figure 6).

Table 3. Diurnal variation of fungal species richness and abundance of air-borne fungi in the atmosphere of Doha, Qatar. A total of 72 exposure samplings during the period (1st of Feb - 31st of March 2016).

Fungal species	Total colony count			
	6:00 AM	12:00 PM	18:00 PM	00.00 AM
<i>Agaricales</i>	0	0	1	0
<i>Alternaria alternata</i>	6	17	18	13
<i>Alternaria chlamydospora</i>	5	2	3	2
<i>Alternaria phragmospora</i>	1	3	0	1
<i>Alternaria infectoria</i>	5	1	2	0
<i>Alternaria tenuissima</i>	0	1	0	0
<i>Aspergillus sp.</i>	1	1	2	1
<i>Aspergillus flavus</i>	1	2	7	4
<i>Aspergillus sydowii</i>	2	0	0	0
<i>Botrytis sp.</i>	0	1	0	0
<i>Cladosporium cladosporioides</i>	23	77	57	11
<i>Cladosporium herbarum</i>	0	0	0	2
<i>Cladosporium macrocarpum</i>	3	5	10	0
<i>Cladosporium oxysporum</i>	5	6	12	4
<i>Cladosporium sphaerosprumum</i>	5	13	4	4
<i>Cladosporium tenuissimum</i>	2	0	4	1
<i>Cochliobolus hawaiiensis</i>	0	0	0	1
<i>Fusarium (genus)</i>	5	7	6	6
<i>Corynascus sepedonium</i>	0	0	2	0
<i>Emericella quadrilineata</i>	0	0	1	0
<i>Fusarium dimerum</i>	11	2	6	4
<i>Fusarium clamydosporum</i>	8	6	2	2
<i>Fusarium oxysporum</i>	9	8	5	4
<i>Fusarium moriliforme</i>	0	2	0	0
<i>Fusarium tenuissimum</i>	0	0	2	0
<i>Ganoderma spp</i>	35	35	54	12
<i>Gilmaniella humicola</i>	0	1	0	0
<i>Gibberella intricans</i>	2	0	1	0
<i>Penicillium canescens</i>	0	0	3	0
<i>Humicola sp.</i>	0	0	1	0
<i>Penicillium chrysogenum</i>	0	0	1	0
<i>Penicillium citricum</i>	3	0	1	0
<i>Penicillium brevicampactum</i>	4	0	1	0
<i>Penicillium italicum</i>	0	0	1	0
<i>Phoma sorghina</i>	1	0	2	0
<i>Rhizopus oryzae</i>	0	1	0	0
<i>Thanatephorus cucumenis</i>	0	1	2	0
<i>Trichoderma viride</i>	5	0	0	0
<i>Ulocladium botrytis</i>	0	3	2	7
<i>Ulocladium chartarum</i>	1	0	1	0
Total colony count	143	195	214	79
Total species	23	23	30	15

Table 4 . Diurnal variation of species richness and abundance of air-borne fungi in the atmosphere of Doha, Qatar, in respect to variation in temperature and relative humidity. Within each column, values with common letters are not significantly different at $P \leq 0.05$ according to Tukey's test. A total of 72 exposure samplings during the period (1st of Feb - 31st of March 2016).

Sampling time	Mean Temp °C	Mean RH %	Total colony count	Total number of fungal species
6:00 h	18.2	70.3	143bc	23ab
12:00 h	24.5	43	195ab	23ab
18:00 h	23.1	52.5	214 a	30 a
00:00 h	19.5	63.5	79c	15b

Table 5 . Similarity table among different diurnal times in respect to fungal species encountered from 72 exposure samplings in the atmosphere of Doha, Qatar during the period (1st of Feb - 31st of March 2016).

Time	0:00	6:00	12:00	18:00
0:00	100%	65%	58%	41%
6:00	65%	100%	48%	56%
12:00	58%	48%	100%	39%
18:00	41%	56%	39%	100%

Jaccard similarity coefficient = $(c/(a+b)-c)*100$. C= number of common species between any two time periods (a & b).

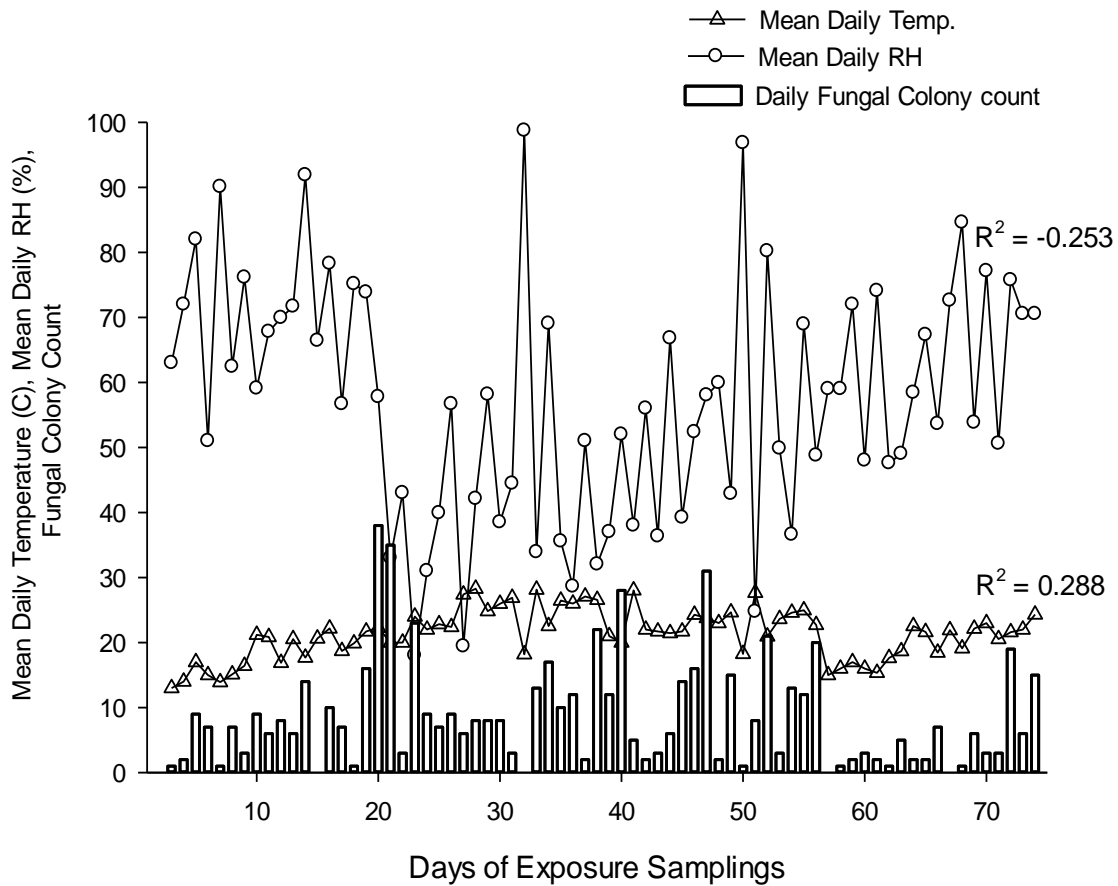


Figure 5. Variations in daily fungal colony counts in respect to changes in mean daily temperature and mean daily relative humidity (RH). Correlation coefficient values (R^2) are significantly different between mean daily counts and each of mean daily temperature and mean daily relative humidity at $P \leq 0.05$ according to Pearson correlation coefficient.

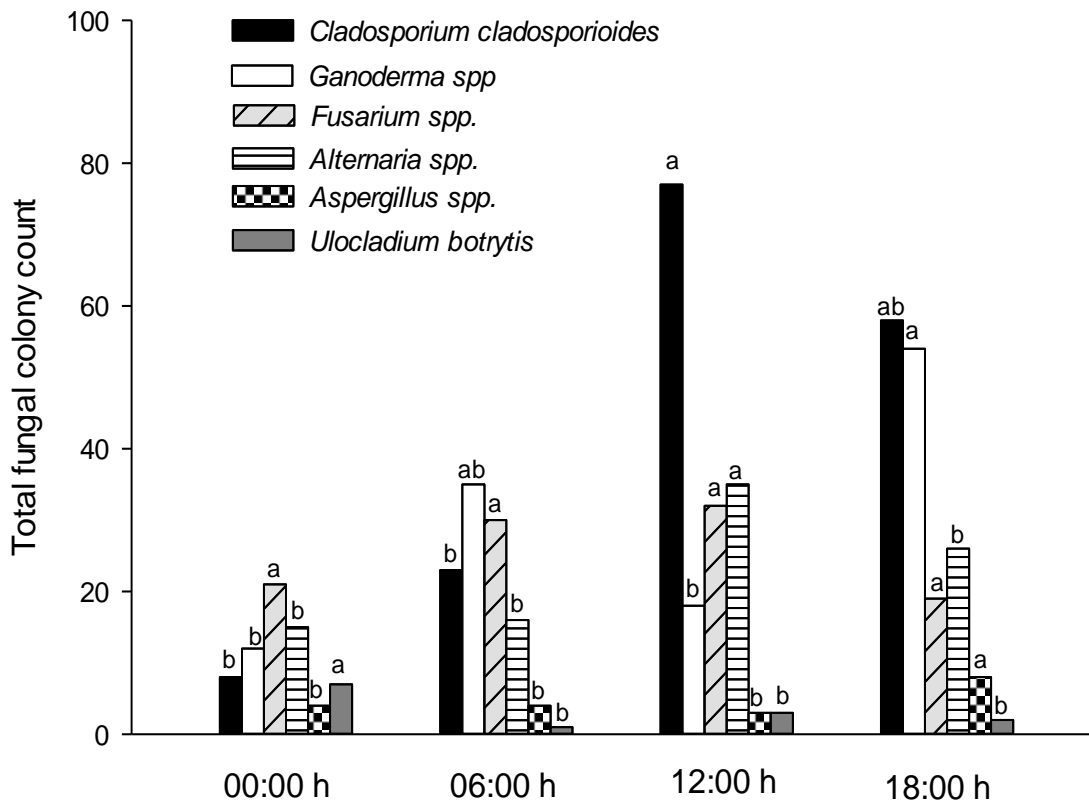


Figure 6. Diurnal variations in total fungal colony counts of the most common species in the atmosphere of Doha during the period (1st of Feb - 31st of March 2016). A total of 72 exposure samplings. Within each fungal taxa, values with common letters are not significantly different at $P \leq 0.05$ according to Tukey's test. Significant correlations were reported between mean daily temperature and each of mean daily counts of *Alternaria* spp. ($R^2 = 0.316$) and mean daily counts of *Ganoderma* spp. ($R^2 = 0.237$) at $P \leq 0.05$ according to Pearson correlation coefficient.

4.3. Investigating the importance of culture media on recovery of fungal diversity and abundance

The total number of sampling plates (exposures) with two different culture media during the study period (1st of Feb - 31st of March 2016) using gravimetric method was 27 for each different culture medium. The exposure samplings were done at the same time (3:00 pm) and in the same location (Qatar University Campus). Table 6 shows the total number of colonies, genera and species of mould fungi (Filamentous fungi) recovered from the two culture media. Yeast fungi were dealt with as one group. From PDA and Rose Bengal culture media, the total number of colonies retrieved was 473 and 336 colonies respectively. According to T-test at $P \leq 0.05$ there was no significant difference in total number of fungal colonies collected at each sampling event using two different culture media (Figure 7). 14 genera and 34 species were collected from PDA, while 13 genera and 25 species were recovered on Rose Bengal media (Table 6). The pair comparison of total number of fungal species collected at each sampling event using the two different media didn't show significant difference with T-test at $P \leq 0.05$ (Figure 8). The most predominant fungi genera which commonly retrieved from both PDA and Rose Bengal media were attributed to *Cladosporium*, *Alternaria*, *Fusarium*, *Penicillium*, *Ganoderma*, *Aspergillus* and *Ulicladium* (Figure 9). Yeast spp. were also isolated from both culture media. For *Cladosporium*, *Aspergillus* and *Penicillium*, higher recoveries were obtained with PDA medium than Rose Bengal medium (Figure 9). The concentration yield of *Fusarium*, *Ganoderma* and *Ulicladium* were higher with Rose Bengal medium than with PDA. The total recoveries of *Alternaria* were similar on both culture media (Figure 9). Yeast spp.

were most often isolated with higher colony- forming unit (CFU) value from PDA medium. *Mucor*, *Agaricales* , *Phoma*, *Stachybotrys* and *Rhizopus* were only isolated from PDA medium, whereas *Humicola*, *Hypomyces* , *Monodyctis* and *Tritirachium* were exclusively obtained with Rose Bengal medium (Table 6).

Table 6. Total number of colonies and fungal species retrieved from the two culture media during the study period (1st of Feb - 31st of March 2016) using gravimetric method (a total of 27 exposure samplings).

#	Fungal Taxa	Total colony count	
		PDA medium	Rose Bengal medium
1	<i>Agaricus spp.</i>	1	0
2	<i>Alternaria sp</i>	2	0
3	<i>Alternaria alternate</i>	12	4
4	<i>Alternaria brassicicola</i>	1	0
5	<i>Alternaria chlamydospora</i>	6	26
6	<i>Alternaria infectoria</i>	10	0
7	<i>Alternaria phragmospora</i>	1	0
8	<i>Alternaria tenuissima</i>	0	2
9	<i>Aspergillus sp.</i>	1	0
10	<i>Aspergillus carbonarius</i>	1	0
11	<i>Aspergillus flavus</i>	8	11
12	<i>Aspergillus sydowii</i>	4	0
13	<i>Aspergillus sulphureus</i>	1	0
14	<i>Aspergillus versicolor</i>	0	1
15	<i>Aspergillus ustus</i>	4	1
16	<i>Cladosporium cladosporioides</i>	226	146
17	<i>Cladosporium herbarum</i>	1	0
	<i>Cladosporium</i>	16	
18	<i>Macrocarpum</i>		6
19	<i>Cladosporium oxysporum</i>	3	6
20	<i>Cladosporium sphaerosprum</i>	22	0
21	<i>Cladosporium tenuissimum</i>	2	0
22	<i>Fusarium (genus)</i>	18	22
23	<i>Fusarium dimerum</i>	5	6
24	<i>Fusarium clamydosporum</i>	2	3

25	<i>Fusarium oxysporum</i>	6	13
26	<i>Ganoderma spp</i>	25	34
27	<i>Gilmaniella humicola</i>	1	1
28	<i>Humicola grisea</i>	0	1
29	<i>Hypomyces chrysosperum</i>	0	1
30	<i>Humicola sp</i>	0	1
31	<i>Humicola insolens</i>	0	4
32	<i>Monodyctys sp.</i>	0	2
33	<i>Mucor sp</i>	1	0
34	<i>Penicillium (genus)</i>	3	0
35	<i>Penicillium canescens</i>	1	1
36	<i>Penicillium citricum</i>	0	0
37	<i>Phoma sp</i>	1	0
38	<i>Pithomyces</i>	1	5
39	<i>Rhizopus stolonifer</i>	1	0
40	<i>Stachybotrys elegans</i>	1	0
41	<i>Tritirachium sp</i>	0	1
42	<i>Ulocladium atrum</i>	0	1
43	<i>Ulocladium botrytis</i>	1	14
44	<i>Ulocladium chartarum</i>	1	0
45	Yeast group	83	23
Total number of colonies from the whole study period		473	336
Total number of genera recorded		14	13
Total number of species recorded		34	25

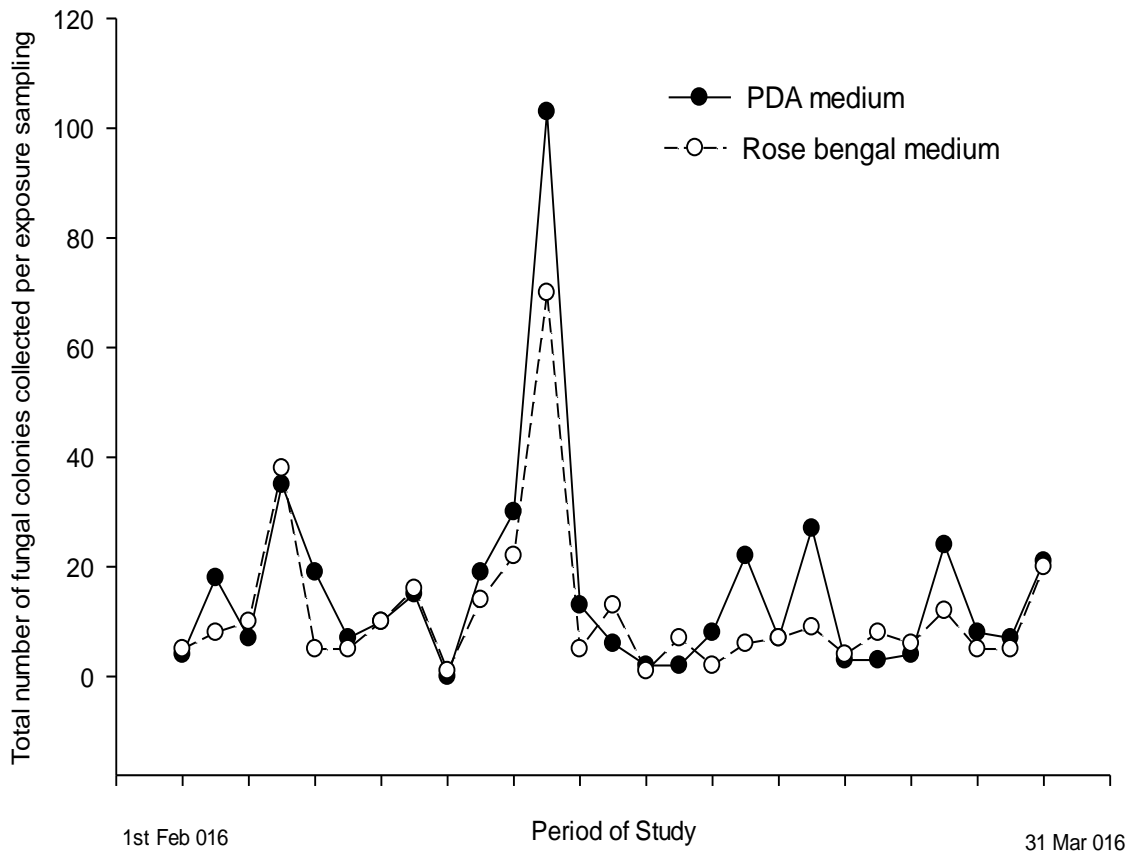


Figure 7. Total number of fungal colonies collected at each sampling event during the study period (1st of Feb - 31st of March 2016) in each of the two culture media used. According to T-test at $P \leq 0.05$ there was no significant difference due to type of culture media used (a total of 27 exposure samplings).

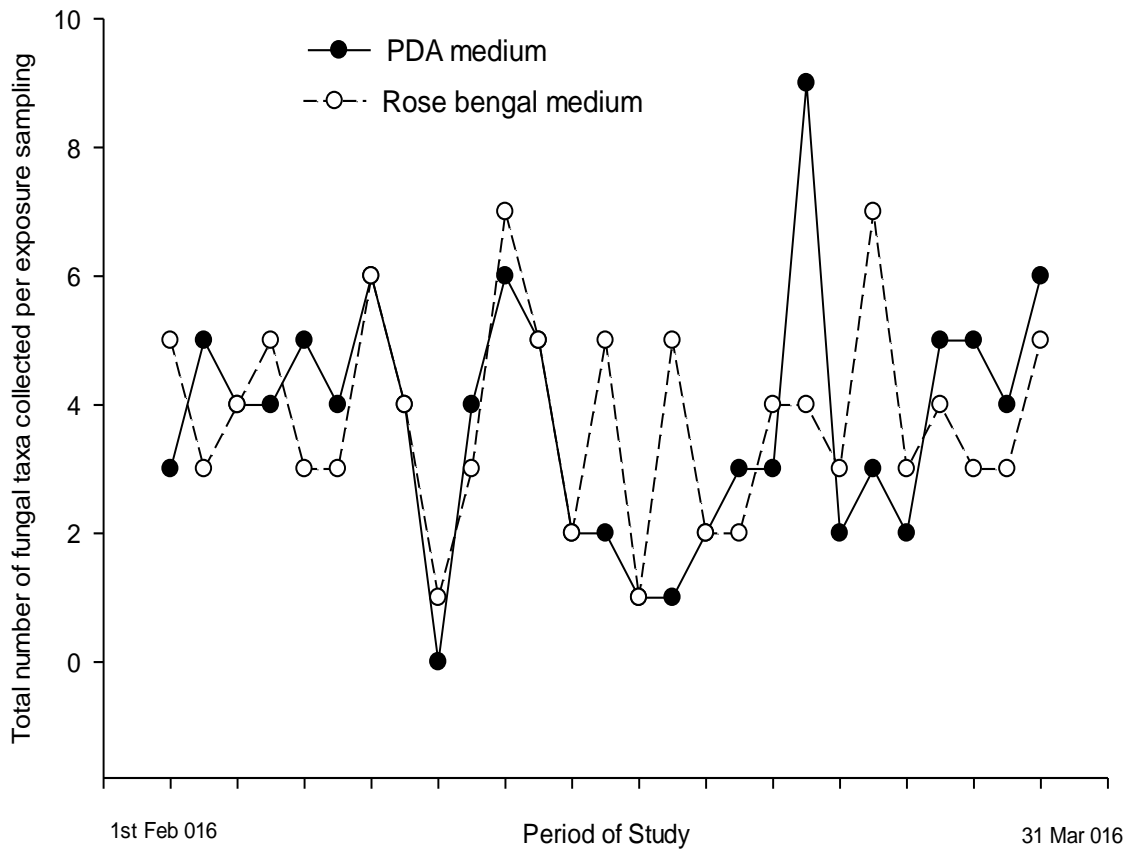


Figure 8. Total number of fungal species collected at each sampling event during the study period (1st of Feb - 31st of March 2016) in each of the two culture media used. According to T-test at $P \leq 0.05$ there was no significant difference in number of species due to type of culture media used (a total of 27 exposure samplings).

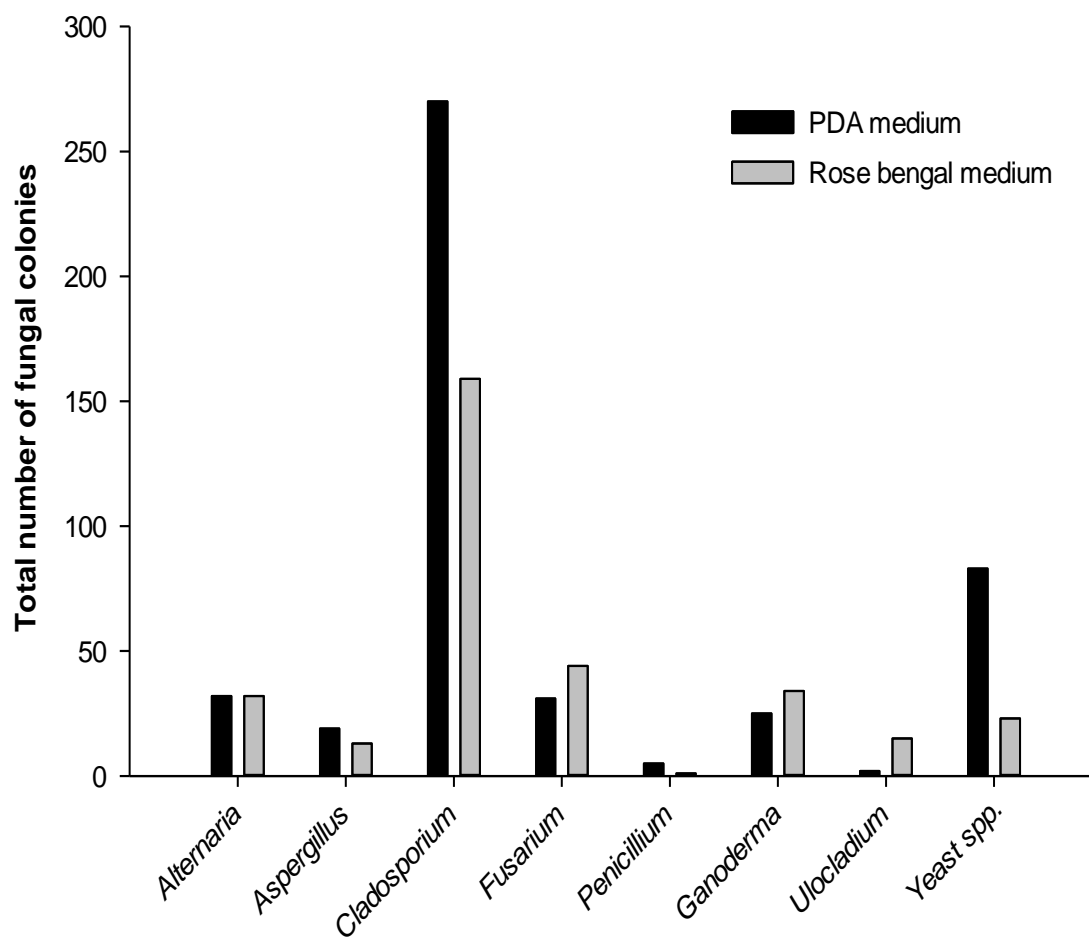


Figure 9. Total number of fungal colonies grown from the most common encountered genera during the study period (1st of Feb - 31st of March 2016) in each of the two culture media using gravimetric method (a total of 27 exposure samplings).

4.4. The impact of atmospheric status (CO₂ concentration) on the dynamics of airborne fungi

Samples were collected in parallel and at the same time at Qatar University Campus and close to the Industrial area sites (around 25 km in distance) during the study period (1st of Feb - 31st of March 2016) using gravimetric methods. Table (7) represents the total number of colonies and fungal species obtained from the two study sites. 358 colony-forming unit (CFU), 16 fungi genera and 35 species were counted at the Industrial area site. From Qatar University Campus site, a total of 312 colonies, 12 fungal genera and 31 fungal species were collected. No significant differences were observed in the total number of colonies and fungal species between the two study sites, although daily concentration of CO₂ was higher, according to average, median and range values of CO₂ at the Industrial area site than at Qatar University Campus (Figure 10). The common and most abundant fungal taxa recorded at the two study sites were attributed to *Cladosporium*, *Ganoderma*, *Fusarium* and *Alternaria*. Corresponding to CO₂ concentrations, the daily colony count of *Cladosporium* was significantly higher at Qatar University Campus than at Industrial area ($P = 0.05$) (Figure 11). Contrary to *Cladosporium*, the concentration of *Fusarium* and *Alternaria* were significantly higher ($P \leq 0.05$) at Industrial area site (Figure 11). The main constituents of fungal genera and their abundance rate in the atmosphere of the two study sites were widely variable between the two sites (Figure 12). *Cladosporium* was prominently higher in relative abundance (67%) at the university campus constituting about two third of all fungal composition. Other fungi like *Ganoderma*, *Fusarium*, and *Alternaria* are also main constituents with relative abundances of 11%, 7%, 6%, respectively. Other

taxa at the University Campus had 10% abundance (Figure 12). While *Cladosporium* is still the main constituent of fungal taxa at the Industrial area site but its relative abundance (29%) is very low compared to the university site (Figure 12). Interestingly, *Ganoderma* showed similar relative abundances (11%) in the two sites (Figure12). The other genera of fungi *Alternaria* and *Fusarium* are considered main constituents with relative abundances of 19% and 17% respectively; however other fungal taxa are still represented by 23% (Figure 12).

Table 7. Total number of colonies and fungal species retrieved from the two study sites during the study period (1st of Feb - 31st of March 2016) using gravimetric method (a total of 22 exposure samplings). No significant correlations at $P = 0.05$ were obtained neither for total colony count or number of species reported.

#	Fungal Taxa	Total colony count	
		Qatar University Campus	Industrial area
1	<i>Agaricales</i> sp.	1	0
2	<i>Alternaria</i>	20	70
3	<i>Alternaria alternata</i> (FR.) Keissler	12	43
4	<i>Alternaria brassicicola</i> (Schwein.) Wiltshire	1	0
5	<i>Alternaria chlamydospora</i> Mouchacca	1	21
6	<i>Alternaria infectoria</i> E.G. Simmons	2	1
7	<i>Alternaria phragmospora</i> V. Emden	4	3
8	<i>Alternaria</i> sp.	0	0
9	<i>Alternaria tenuissima</i> (Kunze) Wilt.	0	2
10	<i>Aspergillus</i>	17	8
11	<i>Aspergillus flavus</i> Link	7	5
12	<i>Aspergillus</i> sp.	1	0
13	<i>Aspergillus sulphureus</i> (Fres.) Thom & Church	1	0
14	<i>Aspergillus sydowii</i> (Bainier & Sartory) Thom & Church	4	1
15	<i>Aspergillus carbonarius</i> (Bainier) Thom	0	2
16	<i>Aspergillus ustus</i> (Bain.) Thom & Church	4	0
17	<i>Cladosporium</i>	210	103
18	<i>Cladosporium cladosporioides</i> (Fres) de Vries	173	83
19	<i>Cladosporium macrocarpum</i> Preuss	12	11
20	<i>Cladosporium oxysporum</i> Berk. & M.A. Curtis	2	3
21	<i>Cladosporium sphaerospermum</i> Penzig	22	2
22	<i>Cladosporium tenuissimum</i> Cooke	1	4
23	<i>Cochliobolus</i>	0	1
24	<i>Cochliobolus australiensis</i> (Tsuda & Ueyama) Alcorn	0	1
25	<i>Fusarium</i>	22	62
26	<i>Fusarium</i> sp.	10	52
27	<i>Fusarium clamydosporum</i> Wollenweber & Reinking	3	1
28	<i>Fusarium dimerum</i> Penzig	3	3
29	<i>Fusarium oxysporum</i> Schlecht.	6	6
30	<i>Ganoderma</i> spp.	30	42

Table 7. continue

31	<i>Humicola</i>	0	2
32	<i>Humicola grisea</i> Traaen	0	2
33	<i>Mucor sp</i>	1	0
34	<i>Neurospora sp.</i>	0	32
35	<i>Penicillium</i>	5	6
36	<i>Penicillium sp.</i>	3	0
37	<i>Penicillium canescens</i> Sopp	1	3
38	<i>Penicillium citrinum</i> Thom	1	1
39	<i>Penicillium chrysogenum</i> Thom	0	1
40	<i>Penicillium corylophilum</i> Dierckx	0	1
41	<i>Phoma</i>	1	5
42	<i>Phoma glomerata</i> (Corda) Wollenw.	0	4
43	<i>Phoma sorghina</i> (Saccardo) Boerema, Dorenbosch & Kesteren	0	3
44	<i>Phoma sp</i>	1	0
45	<i>Rhizopus</i>	1	1
46	<i>Rhizopus oryzae</i> Went & Prinsen-Geerligs	0	1
47	<i>Rhizopus stolonifer</i> (Ehrenb.) Lind	1	0
48	<i>Scopulariopsis</i>	0	1
49	<i>Scopulariopsis candida</i> Vuillemin	0	1
50	<i>Stachybotrys</i>	1	3
51	<i>Stachybotrys chartarum</i> (Ehrenb.) Hughes	1	0
52	<i>Stachybotrys elegans</i> (Pidopl.) W. Gams	0	5
53	<i>Thanatephorus</i>	0	1
54	<i>Thanatephorus cucumeris</i> (Frank) Donk	0	1
55	<i>Trichoderma</i>	0	1
56	<i>Trichoderma viride</i> Persoon	0	1
57	<i>Ulocladium</i>	2	15
58	<i>Ulocladium botrytis</i> Preuss	1	14
59	<i>Ulocladium chartarum</i> (Preuss) Simmons	2	1
	UNKNOWN	2	4
Total number of colonies from the whole study period		312	358
Total number of genera recorded		12	16
Total number of species recorded		31	35

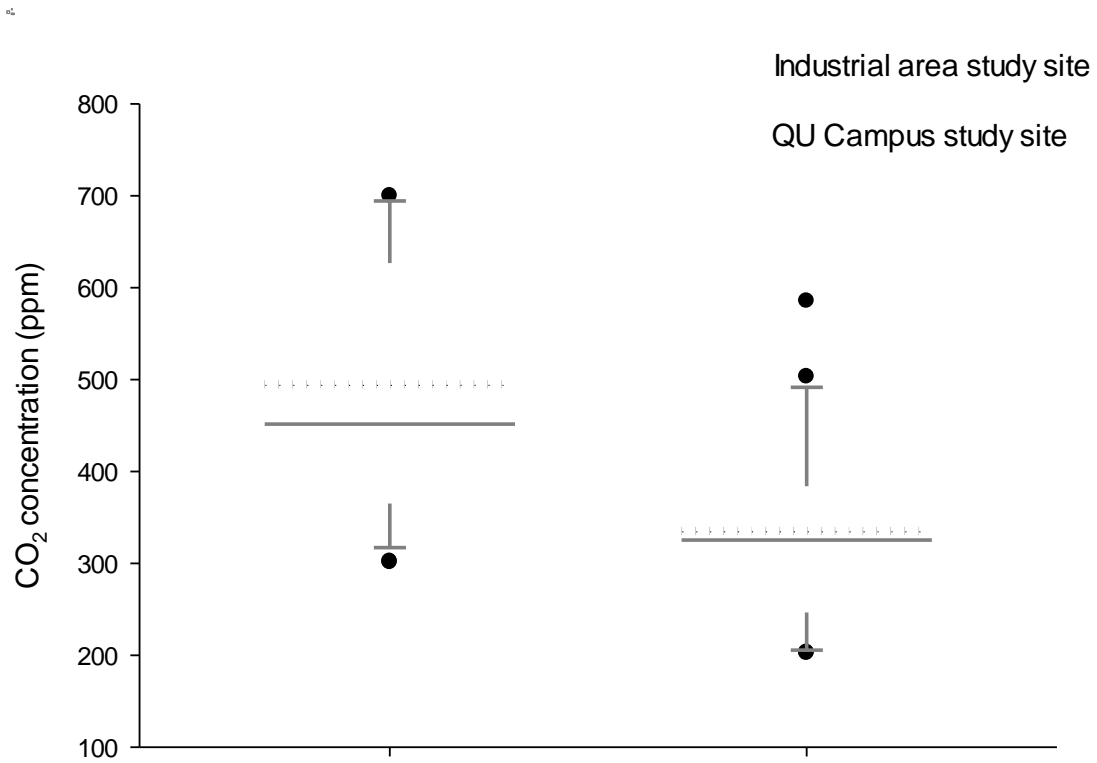


Figure 10. Box plot represents the daily concentration of CO₂ recorded in each of the two study sites during the period of (1st Feb – 31 Mar 2016). The dotted lines refer to mean daily count. The blacked circles represent the outlier values.

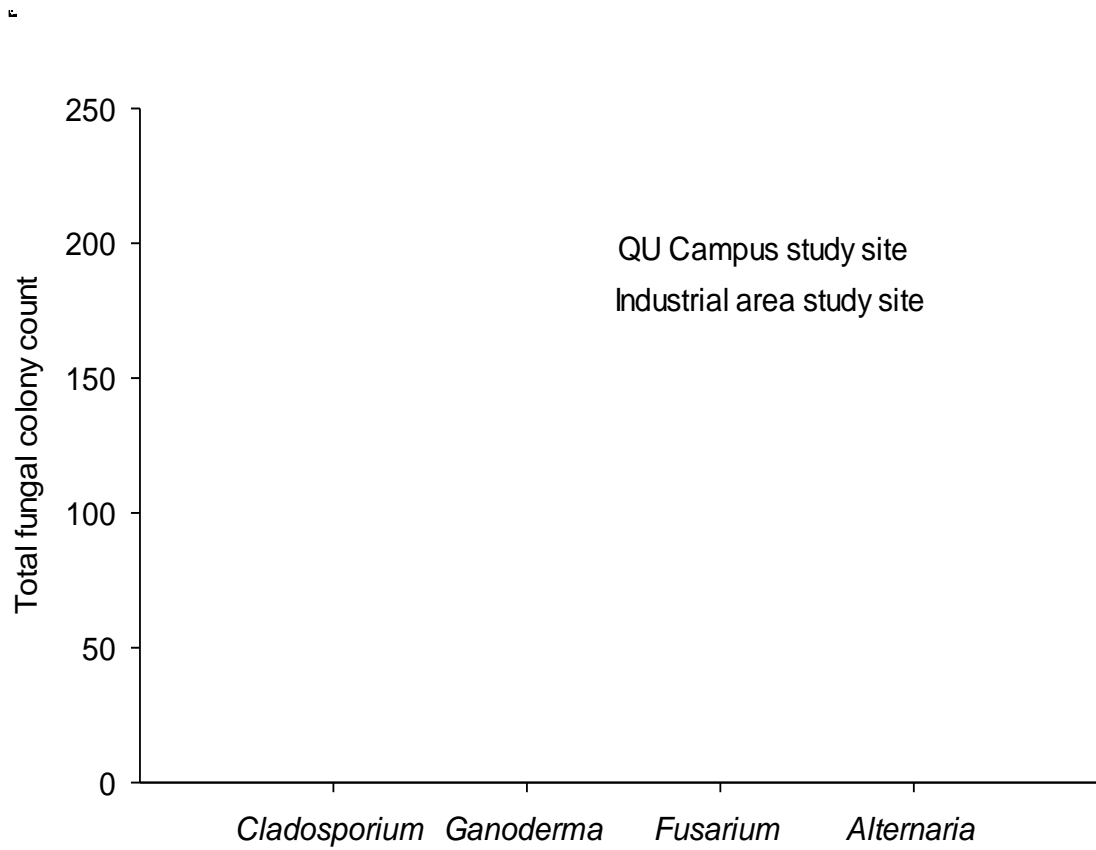


Figure 11. Fungal taxa that shown significant abundance differences between the two study sites (according to t- test of daily colony count at $P = 0.05$) during the period of (1st Feb – 31 Mar 2016) with a total of 22 exposure samplings.

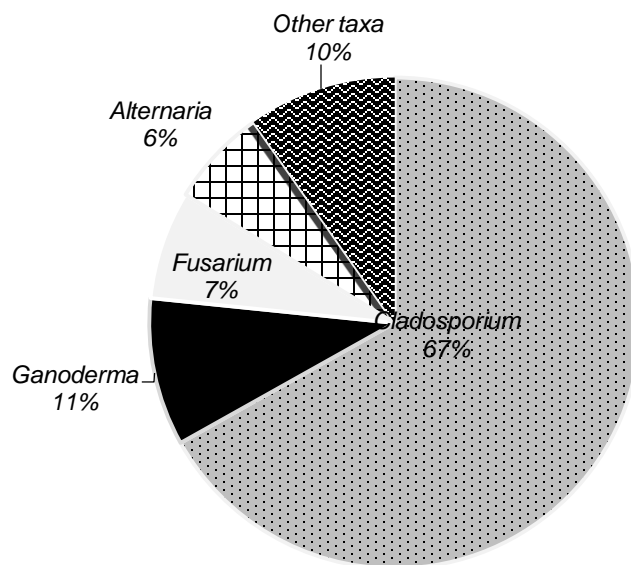
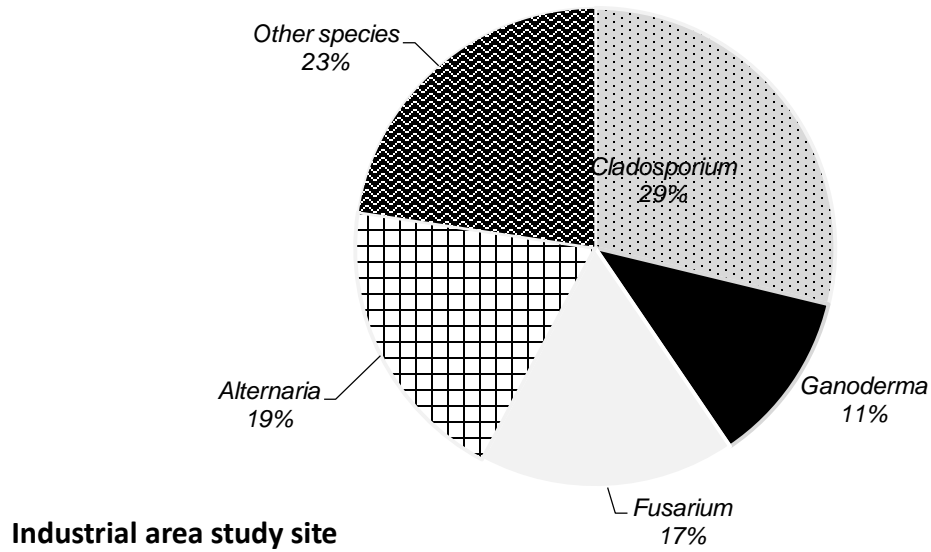


Figure 12. Pie graph to show the main constituents of fungal genera and their relative abundances in the atmosphere of the two study sites during the period of (1st Feb – 31 Mar 2016) with a total of 22 exposure samplings.

CHAPTER FIVE

DISCUSSION

5.1. Seasonal variations of fungal spore populations in the atmosphere of Doha

5.1.1. Fluctuations of abundant species

In the present study, the dynamics of fungal airspora in the atmosphere of Doha, Qatar were investigated over one year (1st of Apr 2015 - 31st of March 2016) using plate exposure method in which a total of 1197 colony counts and 21 genera assigned to 62 species were retrieved (Table 1). A similar study has been conducted in Doha by Al-Subai (2002) during the period of March 1997–March 1998. The author used similar method (plate exposure method) to study the effect of meteorological factors on the airborne fungi population in the air of Doha; however there were major differences between the present study design and the Al-Subai's design. In the present study, one Petri dish was exposed on the roof of the Qatar University building (about 12 m above ground level) , where at this height the composition of airborne fungal spores is more stable, for 15 minutes at 3:00 pm on each of 3 days a week (alternate days). In Al-Subai (2002), plate exposure events were assigned to 24 days monthly for 10 minutes at a level of about 1.5 m at 9 a.m. In Al-Subai (2002) study, 1% glucose-Czapek's agar + 0.05% yeast extract culture medium, while in this study Potato Dextrose Agar (PDA) was used as a culture medium. Both, the height level of collection and culture media used are considered variables that affected the species composition and abundance of air mycoflora (Burge *et al.*, 1977; Herrero *et al.*,1996; Khattab & Levetin, 2008). The above differences in the methodology of the two studies as well as the long

time between the two studies (about 17 years) may explain the differences in the results obtained and make them not comparable. Importantly to mention that huge anthropogenic changes including urbanization, industrialization, and establishment of housing compounds, shopping centers and high-way roads were occurred in Doha within the last 15 years. Those anthropogenic changes are also expected to affect the dynamics and species composition of air-borne fungal spores. According to Al-Subai (2002), the major obtained airborne fungal genera were: *Cladosporium*, *Alternaria* and *Ulocladium*. From our study, *Cladosporium* (60.2%), *Aspergillus* (10.4 %), *Fusarium* (9.4 %), *Alternaria* (8.5 %), *Ganoderma* spp. (2.3%) and *Penicillium* (2.0 %) (Figure 1) were the main component of the air mycoflora. Several studies demonstrated the presence of the above mentioned fungal taxa as part of the main aeromycota composition in different Middle Eastern countries (Abdel-Hafez, 1984; Al-Subai, 2002; Sen & Asan, 2009; Abu-Dieyeh *et al.*, 2010; Quintero *et al.*, 2010; Shams-Ghahfarokhi *et al.*, 2014). Our study showed that *Cladosporium* was the most common fungal genus among others and presented in all months with double peaks one in April and the other in February. Many authors presented *Cladosporium* as the main air mycoflora in different countries world-wide (In Jordan: Abu-Dieyeh and Barham (2014); In Japan: Takahashi 1997; in Turkey: Sen and Asan 2001; in Qatar: Al-Subai 2002; In Egypt: El-Morsy (2006). Many factors might support the abundance of *Cladosporium* throughout the year. Ali *et al* (1976) and Al-Subai (2002) demonstrated that the structural features of *Cladosporium* conidia such as chlamydospore-like structure make them more resistant to solar radiation and physicochemical agents; however other authors explained this phenomenon by referring to the small size, thin exine

and smooth wall of the *Cladosporium* spores which help the dissemination of conidia (Shaheen, 1992; Asan *et al.* 2004).

Aspergillus sp. was the second predominant fungal genera and assigned for ten species (Table 1). *Aspergillus* represented a seasonal pattern; the highest concentration was in August and the lowest in January (Figure 3). Khan *et al.*, (1999) demonstrated that, *Aspergillus* spp. were the predominant in the outdoor environment of Kuwait. Similarly, Abdel-Hafez (1984) reported that *A. sydowi*, *A. flavus*, *A.niger* and *A. versicolor* among the most common species in the air of Taif, Saudi Arabia., and *Aspergillus* sp. were also found to be the most abundant fungal genus in the atmosphere of Saudi Arabia (Abdel-Hafez, 1985). The abundance of *Aspergillus* sp. in the atmosphere of Doha and other Gulf regions is consistent with the findings provided by Christensen & Tuthill (1986) who found that the tropical/subtropical habitats accommodated higher number of *Aspergillus* species in comparison to temperate regions. Accordingly, they concluded that *Aspergillus* genus is highly abundant in the tropics/subtropics especially in the saline and cultivated soils.

In the present study, *Fusarium* was ranked as the third among other most common airborne fungi (Figure 1). *Fusarium* is a cosmopolitan fungus which is well known as plant/animal pathogen (Smith, 2007). *Fusarium* germination can't be developed well at low water activity (Nelson *et al.*, 1994). The dispersion of *Fusarium* macroconidia was found to follow a rain or irrigation event (Fernando *et al.*, 2000). Coincides with that Abu-Dieyeh *et al.* (2010) reported that *Fusarium* was abundant in the atmosphere of the Zarqa area, Jordan and correlated this abundance with the continuous drip irrigation of olive plantations in the study site and also the existence of many farms around the study sites. The previous

findings may explain the abundance of *Fusarium* in the atmosphere of Doha since the study site, Qatar University campus, receives extensive and frequent irrigations to accommodate the need of the vegetation cover (turfgrass areas and others) and the high evaporation rate. However rainfall might interact with density of *Fusarium* but it's difficult to achieve any significant correlation trends since the rainfall in Qatar is very low and limited to few days of the year.

Alternaria represented the fourth most prevalent fungal taxa in the air with seven identified species (Figure 1). *Alternaria* represented a seasonal trend, with high colony count in May and July while the colony count declined a little and continue all over the year with almost minor fluctuation (Figure 3). Similar to *Cladosporium* spores, the conidia of *Alternaria* are of dry spore's type, dark color which tolerate the solar radiation and survive the whole year (Al-Subai, 2002). Other studies showed that the highest spore concentration of *Alternaria* spp. was reported in July and August, followed a wet weather (Gregory & Hirst, 1957; Grinn-Gofroń & Rapiejko, 2009). In India, when the condition is suitable for plant decomposition, after winter, the dispersal of *Alternarai* become the maximum (Mishra, 1971). *Ganoderma* spp. was one of the major constituents of air mycoflora of Doha (2.4 %), but it was exclusively retrieved in February and March of the whole year (Table 1; Figure 1). In the present study, *Ganoderma* spp. has been identified for the first time in the atmosphere of Doha. Similar to our findings ,in Saudi Arabia, Jizan a considerable concentration of *Ganoderma* basidiospores was detected within the airspora (Hasnain *et al.*, 2004). According to the later authors, airborne *Ganoderma* spp. was not frequently detected in many countries with similar ecological features in the Gulf region. *Ganoderma*

basidiospores have ovoid or egg shape, the internal wall of spores is colored with a colour varying from golden to dark brown, the external wall is smooth and transparent where the two walls are connected by pillars (Pegler & Young, 1973). A flattened basal apiculus is another feature of the spores (Southworth, 1974).

Ganoderma was first suggested to be associated with respiratory allergy by Gregory & Hirst (1952). In Saudi Arabia, Jizan the concentration of *Ganoderma* basidiospores was significantly correlated with the high incidence of asthma in Jizan (Hasnain *et al.*, 2004). From our study, *Ganoderma* basidiospores exclusively appeared in late February and March of the whole year (Table 1; Figure 1). The seasonal pattern of *Ganoderma* is widely varied worldwide. In Jizan region in Saudi Arabia, the seasonal trend of *Ganoderma* was shown from October to March with a peak in December and January when the temperature is moderate and the humidity is relatively high (Hasnain *et al.*, 2004). In Poland, Stępańska & Wołek (2005) found August to be the month of the highest spore incidence when the weather is warm and humid. In USA, the spores of *Ganoderma* were reported on more than 95% of the days, lasting from June through October and reaching maximum concentration in late August until mid-October with a positive correlation with temperature and precipitation (Craig & Levetin, 2000). In India, Delhi, *Ganoderma* had seasonal pattern from July to September (Singh *et al.*, 1995). The presence of *Ganoderma* basidiospores is determined by the abundance of vegetation coverage (Cabarroí *et al.* 2008), and also the maximum spore concentration was reported to occur during the rainy months (Almaguer *et al.*, 2014). From the above studies, the incidence of *Ganoderma* basidiospores seems to be more correlated with moderate temperature and high moisture or rainfall which may

explain our findings about why they only reported in February (10%) and March (90%) as March was the month of the highest rainfall during the study period (Table 2).

5.1.2. Fungal species fluctuation versus climatic factor

According to our results, February was the month of greatest colony count and fungal species richness in the atmosphere of Doha and this can easily be explained due to the relatively moderate temperature and the highest reported rainfall (Figure 3). While August and September were the months of lowest fungal abundance and diversity (Figure 3). These months are characterized by relatively the highest temperature and relatively very high humidity and these conditions might interacted to not facilitate the dissemination of fungal spores from their sources (soil and plants) and/or deteriorated them especially if we also add the factor of sun radiation intensity which is also expected to be elevated during these months.

In the present study, a strong significant ($P \leq 0.01$) positive correlation coefficient ($R^2 = 0.517$) were obtained between the total daily fungal counts and the total daily fungal taxa recorded which may indicate that similar interactions of weather conditions may determine the dynamics and fluctuation of both parameters.

Both daily and monthly counts of either of *Cladosporium* or *Alternaria* spores showed significant negative correlations ($P \leq 0.05$) with daily maximum, minimum and mean temperatures. While the correlations of *Cladosporium* colony count with both daily or monthly relative humidity and rainfall were non-significant, the monthly rainfall data

showed highly significant correlation ($P \leq 0.01$) with *Alternata* colony counts. Highly significant ($P \leq 0.01$) positive correlation was appeared between daily counts of *Cladosporium* and *Alternaria* and daily wind speed data (Table 2). In contrary to our results, in Poland, Grinn-Gofroń & Strzelczak, (2013) found that the concentration of air spores of *Cladosporium* and *Alternaria* were significantly and positively correlated with temperature and negatively with relative humidity, however the authors couldn't clearly rank which weather parameter is more important for the viability of those fungal spores in the air. Similarly, in Spain, Sabariego *et al* (2012) concluded that the mean daily count of *Alternaria* was significantly and positively correlated with temperature but negatively and significantly with relative humidity and rainfall and wind speed. In Turkey, both *Alternaria* and *Cladosporium* were significantly and positively correlated with temperature and humidity (Erkara *et al.*, 2009). Our results showed similar trends with studies that have been conducted in countries with similar environment like Qatar, Hasnain *et al.*, (2012) demonstrated that the occurrence of *Alternaria* spores is negatively and significantly correlated to temperature and Hameed *et al.*, (2007) mentioned a negative insignificant correlation with relative humidity. Çolakoglu (1996) and Nourian *et al.* (2007) found that the concentration of *Cladosporium* spores increased significantly as the temperature decreased which is in support to our findings. The cumulative rainfall was positively correlated with the level of *Alternaria* incidence (Stennett & Beggs, 2004), in the same regards, Gregory and Hirst (1957) noted that the dispersal of *Alternaria* spores was higher in August following a period of wet weather. In general raindrops accelerate the release of dry spores (Inglod, 1971) like *Alternaria* spores. All the above mentioned findings agreed

with our result about the presence of a significant positive correlation between monthly data of *Alternaria* and rainfall.

Interestingly, the meteorological parameters didn't show an influence on the spore incidence of *Aspergillus* in the atmosphere of Doha (Table 1). The nature of *Aspergillus* spores as they are dry, relatively small make them easily and passively released by even minor wind speed or vibration (Lin & Li 2000). Nevertheless, it seems that the inconsistency of the meteorological impacts on the dynamic of airborne fungal spores is due to the differences in the climates between different countries. Our findings agreed with Oliveira *et al.* (2009) who found that *Aspergillus* didn't display correlation with any of the meteorological parameters. In contrary to our results, *Aspergillus* level was positively and significantly correlated with temperature, while significantly and negatively correlation was found with relative humidity (Sen & Asan, 2009). *Aspergillus* was found to be positively and significantly correlated with temperature, though negatively and significantly with relative humidity (Hameed *et al.*, 2012).

Fusarium didn't exhibit any significant correlation with any of the weather data except for wind speed. Fernando *et al.* (2000) noted that similar environmental conditions are favorable for some *Fusarium* species to release their spores, also and even though the rain had an influence on the sporulation rate of *Fusarium*, it was difficult to address the impact of other factors such as soil surface temperature and wind velocity. *Fusarium* germination is well known to develop well in the presence of high moisture (Nelson *et al.*, 1994) and as mentioned earlier, the abundance of *Fusarium* in the study site is due mainly to extensive and frequent irrigations to accommodate the need of the vegetation cover (turf grass areas

and others) and the high evaporation rate. Total daily colony count of all encountered fungi had negative significant correlations ($P \leq 0.05$) with any of the daily temperature parameters. A highly significant positive correlation ($P \leq 0.01$) was also obtained between the total daily or monthly colony count and daily or monthly wind speed (Table-2 & Figure 3) and a negative but insignificant correlation with daily mean relative humidity and rainfall. However no significant correlations were reported between total colony count and either of relative humidity or rainfall. (Figure.2). Similar to our results, Hasnain *et al.* (2012) found that the concentration of several fungal spore counts to be significantly and negatively correlated with temperature, though a positive and insignificant correlation was detected with relative humidity. Similarly, Nourian *et al.* (2007) detected a significant and negative correlation between temperature and colony count while had a positive with relative humidity. Other studies indicated contrary results as the concentration of airborne fungal spores was positively and significantly correlated with temperature (Segvić Klarić & Pepeljnjak, 2006). Based on our statistical correlation analysis between weather data and fungal aerosols, the temperature might have the greatest influence on the dynamics of airborne fungal spores, a fact that have been highlighted by many authors, among those: (Stepalska & Wołek, 2005; Sousa *et al.*, 2008; Grinn-Gofron, 2010; Grinn-Gofroń, & Bosiacka, 2015). Interestingly, the majority of retrieved fungal airospora are mesophilic (optimal temperature for growth 20–40 °C) (Gravesen 1979). Our results showed that relative humidity had no significant impact on the airborne fungi of Doha which might be attributed to the adaptation of germination, growth and propagation of airborne fungi to relative humidity (Nourian *et al.*, 2007). Another reason might be that the high temperature

plus the sun radiation (not studied) are more limiting factors than the humidity in certain months of the year, particularly from July to September. Wind speed was found to exaggerate the number of fungal spores in the atmosphere significantly (Al-Subai, 2002; Erkara, *et al.*, 2008; Erkara *et al.*, 2009; Sabariego *et al.*, 2012) which corresponding with our finding for all fungal taxa, total number of colonies and fungal species, however the impact of wind speed was related to the daily data which means that the impact of wind velocity on the fungal airospra is temporally and on daily basis rather than the accumulative effects (Figure 2). On the other hand, wind can act as a dispersal and diluting factor for spores when the wind is high and so decrease the concentration of spores significantly (Quintero *et al.*, 2010). According to Lin &Li, (2000), when the wind speed is less than 5 m/s, the fungal spore's concentration decreased, but when the wind velocity is more than 5m/s the concentration of fungal spores increased. The change in wind direction didn't exert any effect of fungal abundance or fungal diversity. McDonald & O'Driscoll (1980) found a remarkable impact of wind direction on the count of airborne fungi. On the other hand and coinciding with our findings Al-Subai, (2002) did not find any regular correlation between wind direction and the concentration of fungal spores, which might be related to the fact that Doha is a coastal city and surrounded by the Persian Gulf.

Based on our findings, a fungal spore calendar for air-borne fungi in the atmosphere of Doha and their seasonal patterns was constructed (Figure 4). The highest density of fungal colony counts was reported in February, while the lowest was recorded in August. The highest number of fungal species was reported in February, while the lowest was obtained in August and September. This fungal spore's calendar is clinical important as it will

provide baseline knowledge about concentrations, diversity and dynamics of airborne fungal spore in the atmosphere of Doha. These scientific knowledge are necessary to facilitate and enhance the work for allergists and thus helping them to suggest the risk of allergic sensitization for patients that are sensitive to certain allergic fungal species. Mycologists, applied ecologists and plant pathologists can get benefits from this spore calendar as well. In the present study, *Cladosporium*, *Aspergillus*, *Fusarium*, *Alternaria*, *Ganoderma* spp. and *Penicillium* represented the major fungal airospora in the atmosphere of Doha, Qatar. All these fungi are frequently associated with allergy (Wu *et al.*, 2007; Sen & Asan, 2009; Hasnain *et al.*, 2012; Knutsen *et al.*, 2012). In the Middle East and particularly in the Gulf region only few respiratory fungal studies have been attempted to assess the prevalence of allergic rhinitis, conjunctivitis, bronchial asthma, and allergic bronchopulmonary mycoses developed from the frequent exposure to allergenic spores. In Qatar, Taj-Aldeen *et al.* (2003) investigated the allergic fungal rhinosinusitis (AFS) cases caused by *Aspergillus flavus*, where they found huge quantity of allergic fungal mucin and dark crusts fully colonizing the sinuses which required a course of systemic and topical corticosteroids. The same author demonstrated that in Qatar, an observational study was carried out to estimate the size of fungal infections at public level. The data analysis revealed that, 1486 people were affected by severe asthma with fungal sensitization, 1126 patients were diagnosed with allergic bronchopulmonary and 176 individuals complained from chronic pulmonary aspergillosis (Taj-Aldeen *et al.*, 2015). In Saudi Arabia, even though an aeromycological study was not purposely conducted as respiratory fungal study, a considered relation was found between the high prevalence of asthma in children and the

highest concentration of *Ganoderma* spp. recorded among three study sites (Hasnain *et al.*, 2004). *Cladosporium*, *Alternaria* and *Aspergillus* were believed to significantly causing allergic rhinitis and allergic asthma (Achatz *et al.*, 1995).

In the present study, the spore's concentration of *A. alternata* contributed to more than the half of total *Alternaria* (Table 1). Bush & Sanchez (1997) Achatz *et al.* (1995) considered *A. alternata* as one of the most significant aeroallergens. From our study, *A. flavus* was the predominant of the total *Aspergillus* (Table 1). *A. fumigatus*, *A. niger*, *A. flavus* and *A. oryzae* were frequently correlated with the respiratory allergic cases, *A. falvus* and due to its large spores it deposited in the upper respiratory tract and commonly cause fungal sinusitis (Hedayati *et al.*, 2007). *Cladosporium herbarum* is an important and main cause of inhalant fungal allergens among other *Cladosporium* (Kurup *et al.*, 2000), in this study, *C. cladosporioides* was the main species among *Cladosporium*, *C. herbarum* showed a minor participation to the genus *Cladosporium*. However, *C. herbarum* was more frequently associated with respiratory dysfunctions (Aukrust & Borch, 1979; Reijula *et al.*, 2003; Heinzerling *et al.*, 2005). Skin prick testing (SPT) is an essential technique for diagnosing allergic sensitization such as fungal allergens which will improve the diagnostic accuracy, In Qatar there is an ongoing research about the reactivity of Skin Prick test to common aeroallergen among children with allergic respiratory diseases (Personal communication with Dr. Mohammad Ehlayel, MD, Hamad Mediacorporation, Qatar).

5.2. Diurnal variation of fungal spore populations in the atmosphere of Doha

In the present study, the atmospheric concentrations of fungal spores under the influence of intra-diurnal fluctuations were studied during the month's period 1st of Feb - 31st of March 2016. Several aeromycological studies investigated the diurnal periodicity of airborne fungal spores (Sreeramulu, 1959; Helander & Pessi, 1991; Li & Kendrick, 1995; Al-Subai, 2002; Hameed *et al.*, 2009; Stępańska & Wołek, 2009; Abu-Dieyeh *et al.*, 2010). Our findings revealed an intra-diurnal periodicity pattern of fungal spore concentrations where we found significant differences in the total colony counts and fungal diversity among different time periods (Table 3). Both parameters, total colony count and species fungal diversity were peaked significantly in the atmosphere of Doha at 18:00h and declined significantly at 00:00h (Table 3). A negative correlation between colony count and relative humidity while a positive correlation with temperature was noted (Figure 5). During February and March a homogeneous temperature readings were obtained that might lead to this finding which was completely against to what we found in the relation between seasonal temperature data and total colony count of the whole year. Interestingly, a concentration gradient of fungal spores was observed which occurred in upward flux (started from 00:00 h) coordinating positively with recorded temperature and negatively with relative humidity. Our findings partially agreed with Al-Subai (2002) who studied the aeromycology of Doha during the year (1997-1998) and concluded that the highest concentration of fungal spores occurred at 12:00 when the highest temperature and lowest

relative humidity were indicated, while the lowest concentration was recorded at 00:00h and 06:00h at reverse weather conditions. Other findings indicated that the diurnal pattern of airborne fungi is affected by solar temperature and decreased relative humidity (Joy Royes, 1987). Stępańska & Wołek (2009) demonstrated that the highest spore's concentration was recorded at midday and afternoon due to reduction in the relative humidity. On the other hand, Fengxiang *et al* (1991) found that the viability of airborne fungi was higher during the nighttime than in daytime due to the darkness and high relative humidity that increase the concentration of fungal spores. However the period of study is so important to determine the diurnal periodicity of fungal spores (Abu-Dieyeh & Barham 2014). Our study was conducted during February and March which matches the spring time in Qatar and the diurnal range of minimum humidity is still not detrimental to negatively reduce fungal spore dissemination and this may allow temperature to be the effective variable in the diurnal cycle of fungal spore dissemination. The highest similarity coefficient was obtained between 00:00h (midnight) and 06:00h (early morning), while the lowest similarity coefficient was detected between 18:00h (afternoon) and 12:00h (noon) (Table 5). The most abundant fungal taxa were *Cladosporium cladosporioides*, *Alternaria spp.*, *Fusarium spp.*, *Ganoderma*, *Ulocladium botrytis* and *Aspergillus* (Figure6). *Cladosporium* and *Alternaria* were found in higher concentrations at midday time (Figure6). These two fungal taxa are of clinical important due to their allergenic properties (Kurup *et al.* 2002). The maximum concentrations of *Cladosporium* and *Alternaria* were found to occur at the midday time (Gregory, 1973; Troutt & Levetin, 2001; Ho *et al.*, 2005).

Similar to our findings, *Cladosporium* and *Alternaria* were found to have a positive correlation with temperature but a negative correlation with relative humidity (Recio *et al.*, 2012; O'Connor *et al.*, 2014). The spores of *Cladosporium* are considered dry air spores, thus the midday weather conditions when the temperature is relatively high and relative humidity decreases make it optimum for the dispersion of *Cladosporium* spores (Elvira, 2001). Recio *et al.* (2012) concluded from their study that the highest correlation coefficients in term of maximum, mean and minimum temperature to be for *Alternaria* accordingly they recommended to predict the concentration of *Alternaria* spores as a function of temperature. It was demonstrated that the maximum release of *Ganoderma* spores occurred when the humidity increased and temperature diminished (O'Connor *et al.*, 2014). Similarly, Calderon *et al.* (1995) concluded that the release of basidiospores are negatively correlated with temperature and declines at higher temperature more than 27 °C. Coincides to what's mentioned above, in this study, we reported *Ganoderma* only during February and March in the atmosphere of Doha. We also found that *Ganoderma* significantly peaked at 18:00 h where the average of main daily temperature is 23.1 °C and the average of mean daily humidity is 52.5%. Even though most of the authors highlighted temperature as more detrimental in diurnal fungal cycle, we believe that a minimum degree of humidity is still needed.

Mean rainfall during the study period was the highest throughout the year, the spores of basidiomycetes are wet spores and their dispersal was mainly and directly affected by the precipitation (Lyon *et al.*, 1984). Sufficient moisture, either during rainfall or when humidity increases is required for the spore production of *Ganoderma* (Ho *et al.*, 2005;

Stepalska & Wołek, 2009). By the splash and “tap-and-puff” mechanism, whereby the raindrops hit the leaves and cause the attached spore to release from their colonies into the air (Ho *et al.*, 2005). From this study, the sporulation of *Ganoderma* is more conjunction with rainfall but not with increasing humidity. *Aspergillus* had a major peak pattern at 18:00 h (Figur.6). There is no significant diurnal periodicity of *Aspergillus /Penicillium* due to the nature of their spores as they are dry, relatively small and and thus they can be released passively by even minor wind speed (Lin & Li 2000). Hameed *et al* (2009) observed double peaks pattern of *Aspergillus* 10:00 h and at 20:00 h. The highest concentration of *Aspergillus/Penicillium* occurred at 00:00h (Ho *et al.*, 2005). In the current study, *Ulocladium botrytis* peaked at midnight (00:00 h). Coincides with Al-Suwaine *et al.* (1999) who fund that, the concentration of *Ulocladium* is negatively correlated with temperature

5.3. Investigating the importance of culture media on recovery of fungal diversity and abundance

The nutrients composition and water content of the culture media have been introduced as major factors that influence the growth and recovery rates of fungal spores (Ahamed & Vermette, 2009; Domingues *et al.*, 2000). Two different culture media were used in this study, Potato Dextrose Agar (PDA) and Czapek’s/Rose-Bengal agar, to differentiate between the potentialities of fungal growth on different media. The dextrose concentration

in PDA is double what is found in Czapek's/Rose-Bengal agars, which make it very rich with carbohydrates source. Rose Bengal (dye) and chloramphenicol (antibiotic) are main ingredients of Czapek's/Rose-Bengal agar while they are missing in PDA medium. Our results didn't reveal a significant difference in the total colony number and fungal diversity due to type of culture media (Figure 7 and 8). Hameed *et al.*, (2007) used four culture media, Czapek Dox agar, malt extract agar, Potato dextrose agar and Sabouraud dextrose agar Petri dishes to investigate the examine the yield rate of fungal spores on different sampling media compare the response, however there was no significant difference between the total colony counts using the different culture media, which consistent to our findings. Burge *et al.* (1977) concluded from using eight different culture media to compare the growth pattern of fungal spores that, the total growth were similar on Sabouraud dextrose agar (SDA) , Malt Extract (MALT),V8 juice (V8), and Potato dextrose agar (PDA) , whereas MALT and SDA exhibited the highest occurrences of recovery rate. Selective agents such as dye are usually used to suppress the bacterial growth, among these dyes is Rose Bengal which particularly used to control the growth of spreading molds, reducing the efficiency of colony formation by yeast, reducing the diameter of the fungal colony and might inhibit the bacteria development on the culture media (Ottow, 1972; Henson, 1981; Bragulat *et al.*, 1991). In this study, yeast spp. were most often isolated with higher colony forming unit (CFU) value from PDA medium than with Czapek's/Rose-Bengal agar medium (Figure 9). Also, the recovery of spreading fungi such as *Rhizopus* and *Mucor* on the Rose Bengal medium was absent (Table 6). Furthermore, the observed fungal colony diameters developed on Czapek's/Rose-Bengal agar medium were less than

the diameter of mold colony recovered with PDA medium (data not shown). Interestingly, *Ulicladium* spp. was highly recovered with Czapek's/Rose-Bengal agar medium compared to its growth rate on PDA medium. *Ulocladium* was a highly selective fungal genera where its growth rate on cellulose-Czapek's agar was much higher than with glucose- Czapek's agar (Abdel-Hafez, 1984). In the same regard, *Ulicladium* spp. was the third predominant fungal genera in the atmosphere of Doha using Czapek's agar medium (Al-Subai, 2002). Even though PDA medium is the richest in carbon content, it is not always support the highest recovery pattern of all molds (Whipps & Magan, 1987), this fact support our findings which revealed that higher recovery pattern of *Cladosporium*, *Aspergillus* and *Penicillium* were obtained with PDA medium, while the growth rate of *Fusarium*, *Ganoderma* and *Ulicladium* were higher with Czapek's/Rose-Bengal agar medium (Figure 9).

The composition of culture media makes them very selective for the development of fungal genera and thus determines the recovery and growth rates, accordingly the integration of more than one medium in aeromycological studies might yield different fungal taxa and even different colony counts, colony diameters and morphology of the same fungal genera or species. Thus, there are no perfect media for all molds.

5.4. The impact of atmospheric status (CO₂ concentration) on the dynamics of airborne fungi

The influence of atmospheric CO₂ concentration on the composition and abundance of airborne fungi at two different areas in Qatar were examined in order to investigate the effect of urbanization and air pollution on the availability of aeromycota. The concentrations of CO₂ and other air pollutants have higher levels in response to increased anthropogenic activities (Hameed *et al.*, 2012; Chung *et al.*, 2006). There were no significant differences in the composition and diversity of the airborne fungal population between the two study sites, though daily concentration of CO₂ was higher at the Industrial area site than at Qatar University Campus (Figure 10). Coinciding with our results, from Taiwan, Chung *et al.* (2006) and from USA, Klamer *et al.* (2002) demonstrated that the fungal abundance and their species richness were not significantly influenced by elevated CO₂.

Even though the main constituents of the airospora at the two study sites were attributed to similar fungal taxa *Cladosporium*, *Ganoderma*, *Fusarium* and *Alternaria*, their main concentrations and distribution rate in the air of the two study area were significantly varied (Figure 12). *Cladosporium* showed higher and significant concentration at the Qatar University site in comparison to *Fusarium* and *Alternaria* which presented at higher abundance rate at Industrial area study site, while *Ganoderma* had relative similar distribution rate (Figure 11). This may be explained by the fact that the elevated concentration of atmospheric CO₂ rises the sporulation rate in some fungal genera (Cotty, 1987), also higher CO₂ concentration might decline or altering fungal metabolism which

subsequently either modify, stimulate or even inhibit the fungal growth pattern, spore production and other reproduction processes (Klironomos *et al.*, 2005). Rather than the atmospheric CO₂, other air pollutants such as ozone (O₃), nitrogen dioxide (NO₂), sulphur dioxide (SO₂) and particulate matter PM10 might influence the availability and biological activities of airborne fungal taxa (Hameed *et al.*, 2012). Grinn-Gofroń *et al.* (2011) revealed that the concentration of *Cladosporium* spp. was significantly and negatively correlated with (NO₂), sulphur dioxide (SO₂) and particulate matter PM10, but positively correlated to O₃. Unfortunately the above pollutants were not monitored in the current study, even though we expected to have higher levels of those pollutants at the Industrial area compared to university campus due at least to the big traffic status and this may explain why *Cladosporium* had higher abundance at Qatar University.

In this study, the concentrations of *Alternaria* spp. and *Fusarium* spp. were significantly higher at Industrial area site in corresponding to CO₂ than at Qatar University site. Our findings were harmonized with the results of Klironomos *et al.* (1997) who found that the spore production levels of *Fusarium* spp. and *Alternaria* spp. were stimulated under the condition of elevated CO₂ concentration. Wolf *et al.* (2010) demonstrated that the sporulation of *Alternaria alternata* was significantly amplified by the impact of increasing CO₂ concentration level. It was demonstrated that, *Alternaria alternaria* exhibited higher spore production corresponding to elevated CO₂ concentration. Because increased CO₂ concentration provide the plant with a higher carbon/ nitrogen ratio, pathogenic plant fungi accelerate their growth and sporulation rate (Lake & Wade, 2009; Knutsen *et al.*,

2012). Interestingly, airborne fungal spores were increased by 4-fold responding elevated CO₂ concentration (Klironomos *et al.*, 2005; Klironomos *et al.*, 1997).

CONCLUSIONS & RECOMENDATIONS

In conclusion, our study demonstrated the seasonal and intra-diurnal pattern of the airborne fungi in the atmosphere of Doha, Qatar in addition to that certain interacting factors were also studied. The main fungal airospora were attributed to *Cladosporium*, *Aspergillus*, *Fusarium*, and *Alternaria*. Among the meteorological parameters, temperature might be the main determinant of the fungal spore's incidence and diversity in the atmosphere of Doha. *Ganoderma* spp. is an allergic fungus which was determined as a major fungal constituent in the atmosphere of Doha during February and March. *Ganoderma* spp. was not encountered in Qatar prior to this study. A spore calendar was constructed to reflect the dynamics and fluctuations of different fungal taxa among different months of the year. This spore calendar represents an important piece of knowledge to fill the gap of data shortage about Doha atmosphere and also to provide baseline information for allergists, plant pathologists, meteorologists and other scientists. Different culture media can support the recovery of a different variety of fungal species and this may be responsible for mistaken interpretations and comparison of results among different aeromycological studies. The concentration of atmospheric CO₂ at two locations showed an influence on the occurrence of certain fungal taxa, however other pollutants may also contribute to this influence.

After completing this study and based on the results obtained, we can add the following recommendations for any further or ongoing research about aeromycology of Doha atmosphere:

- Although February and March are the best months of any fungal study in Qatar, since they matched the highest fungal density and diversity, it is recommended to study the intra-diurnal variations on the dynamics of air borne fungi in at least three different seasons to get a better idea about which intra-diurnal period is with the highest fungal spore incidence and diversity.
- It would be of interest to examine the impact of atmospheric concentration of CO₂ on the incidence of fungal airospra for a period of one year. Other pollutants like Sulfer and nitrogen oxides should also be monitored and correlated with prevalent fungal species abundance and diversity.
- This study should be used to produce another study if statistical data about allergic incidence of fungi among Qatari population is available on daily and/or monthly basis
- The impact of *Ganoderma* as an inhalant allergen on the asthmatic patients, particularly among children, is required to be deeply investigated in such environment.
- A further collaboration research should be created between medical doctor allergists in Qatar and scientists from biological backgrounds to investigate the status of dominant allergic fungi and their dynamics not only in outdoor but also in indoor environment, especially in schools, university campuses, hospitals, kinder

gardens and any other dwellings that have a mass of population for a certain period of time.

- A future study considering the socioeconomic and hygienic factors as parameters affecting the fluctuation of indoor fungal airspora is recommended, also the relationship between indoor and outdoor fungal flora should be clarified.
- Although fungal spore calendars are not subject to major changes within few years from the year when it's established, however it can be greatly affected if the environment around the source of fungi (soil and vegetation) due to urbanization, ecosystem changes, agriculture, soil and air pollution were changed. The global climate change is also responsible for changing in dynamics and diversity of airospora Therefore spore calendar data should be reviewed and updated each few years.

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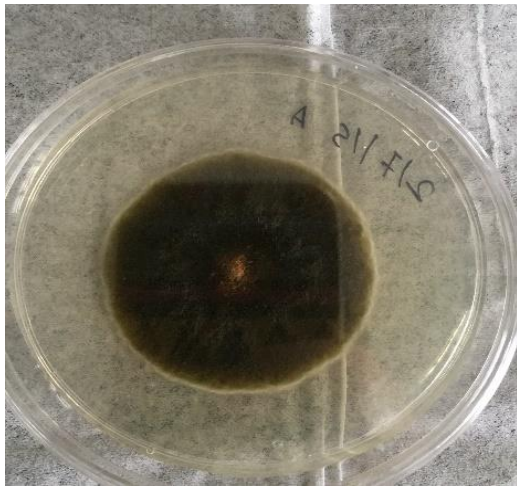
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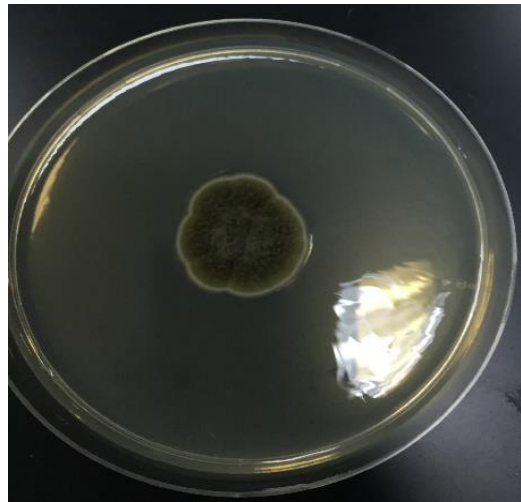
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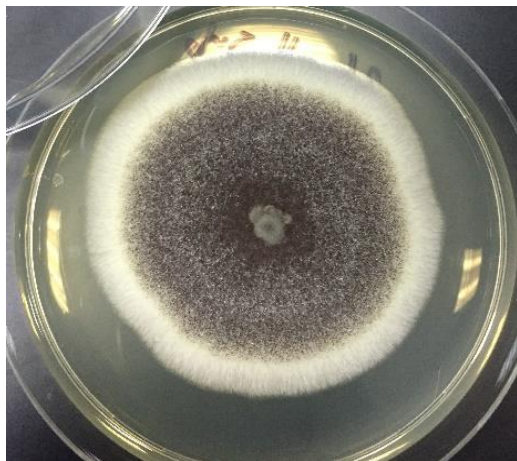
Appendix A: Samples of pictures for selected fungal colonies on Potato Dextrose Agar (PDA) medium.



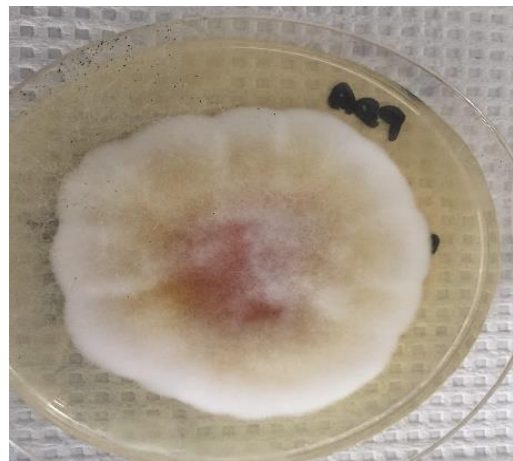
A



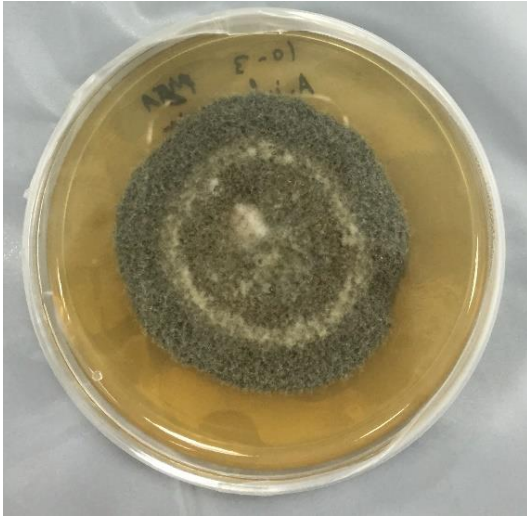
B 3



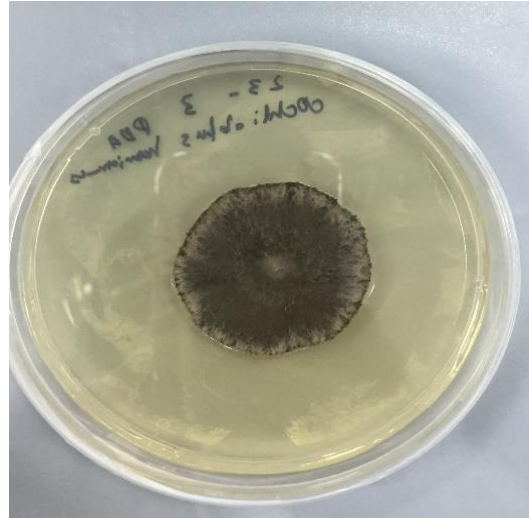
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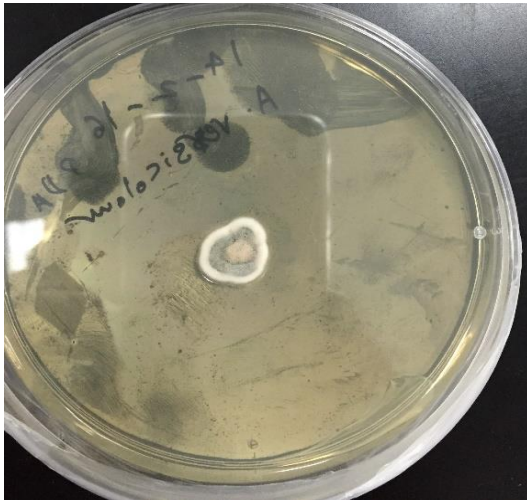
F



E



F



G

- A. *Alternaria alternata*
- B. *Cladosporium cladosporioides*
- C. *Aspergillus carbonarius*
- D. *Fusarium oxysporum*
- E. *Alternaria infectoria*
- F. *Cochliobolus hawaiiensis*
- G. *Aspergillus versicolor*

Appendix B: Samples of pictures for selected fungal spores on slides which photographed under light microscope using the 40X objective lens.



A



B



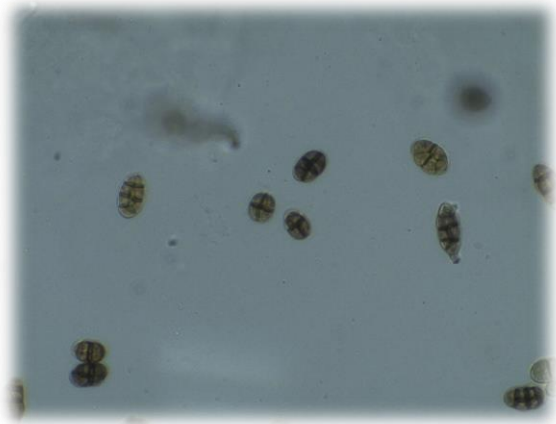
C



D



E



F



G



H

- A. *Aspergillus versicolor*
- B. *Alternaria alternate*
- C. *Cladosporium cladosporioides*
- D. *Cochliobolus hawaiiensis*
- E. *Alternaria infectoria*
- F. *Ulocladium botrytis*
- G. *Ganoderma* spp.
- H. *Aspergillus flavus*