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# **Bio-Waste** *Aloe vera* Leaves as an Efficient Adsorbent for Titan Yellow from Wastewater: Structuring of a Novel Adsorbent Using Plackett-Burman Factorial Design

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Abstract: Titan yellow (TY), a triazene azo dye, was removed from contaminated wastewater samples using a green adsorbent recycled from *Aloe vera* leaves (AV) waste. Two adsorbents were developed—air-dried Aloe vera (ADAV) and thermally treated Aloe vera (TTAV). Adsorption efficacy of both adsorbents was assessed in terms of percent removal (%R) of TY and adsorption capacity  $(q_e)$ . ADAV had a better performance compared to TTAV. Plackett-Burman design (PBD) was exploited to establish the experimental pattern of the study. Four variables were studied: pH, adsorbent dose (AD), dye concentration (DC), and stirring time (ST). Analysis of variance (ANOVA) at 95.0 confidence interval (CI), control, and quality charts helped establish regression model(s). Characterization of both adsorbents was performed using FT-IR/Raman spectroscopy together with TGA/dTGA and SEM/energy dispersive X-ray spectroscopy (EDX) analyses. Textural properties were determined using nitrogen adsorption isotherms at 77 K. Results showed that the surface areas of ADAV and TTAV300 were 3.940 and 7.076 m<sup>2</sup>/g, respectively. Raman analysis showed that the TTAV had clear Dand G-bands. Equilibrium studies revealed that data were well fitted to Freundlich isotherm with a maximum adsorption capacity of 55.25 mg/g using Langmuir equation, and the adsorption was physisorption. Adsorption followed a pseudo-second order that occurred in two steps-diffusion and then adsorption.

**Keywords:** *Aloe vera* (AV) leaves; Plackett–Burman Design (PBD); Titan yellow (TY); %removal and adsorption capacity ( $q_e$ ); characterization; equilibrium; kinetics

# 1. Introduction

Water has ever been the most significant life component for all creatures. With the notable rise in global warming and desertification, water quantity is becoming a worldwide concern [1,2]. While the quantity of water and the share per capita are growing apprehensions, quality of water, which is relapsing progressively—mostly because of the anthropogenic deeds, the population development, the inadvertent expansion, the swift industrialization, and the inappropriate usage of natural water—is becoming a more serious concern [3].

Food, leather, and textile industries release large amounts of contaminated effluents. With the diverse chemical structure of the reagents used in these industries, various problems evolve. Dyes, an example of a major water contaminant, would color large volumes of water even if released as traces as low as 1 ppm. This effect causes not only an undesirable change in water quality in terms of aesthetic properties, but also it cannot be easily reversed or "decolorized". Reports show that, with the huge amount of dyes being produced annually ( $7 \times 10^5$  tons/annum), an amount of 2% of dyes is directly

released into the aquatic system together with an amount of 10% being lost during the tanning process, an issue that signifies the magnitude of the problem [4–6]. Released dyes affect not only aquatic beings but also human health. Their impacts include carcinogenicity, mutagenicity, poisoning, disturbance of the metabolism in aquatic bodies, etc. [7,8].

Titan yellow (TY, Figure 1) is an organic triazene azo dye commonly used in tanning nylon and wool as well as a stain in microscopical examinations. The safety data sheet of TY shows that it is hazardous in case of ingestion with a less hazardous influence on eyes and skin [9,10].

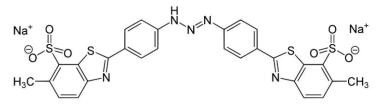


Figure 1. Structure of Titan yellow (TY).

Removal of dyes from wastewater has been done by employing several approaches, e.g., adsorption, bleaching, filtration, biodegradation, and coagulation [11]. These methods possess both pros and cons in terms of cost, availability, efficacy, and surface area and have different influences on the environment. Adsorption, with the capability to totally remove dyes from wastewater, represents an ideal approach. Modeling of an efficient, green, low cost, safe, and reusable adsorbent (in other words, a standard adsorbent) has been the subject of various investigations. While different adsorbents have been used to remove dyes, the most commonly used adsorbents are the activated carbon (char) [12,13].

As developing a standard adsorbent has been the task undertaken in many investigations, a novel category of adsorbents is becoming more prominent–*green* adsorbents. This class of adsorbents offers a dual benefit by removing the contaminants as well as getting rid of the agricultural waste and by-products that would represent a burden on the environment if not properly discarded. Several agricultural waste products (e.g., peels of potatoes, pomegranates, bananas, rice husk, eggshells, crab shells, date pits) have been recycled into adsorbents and were used for the removal of heavy metals, dyes, drugs, etc. [13–18].

Agricultural wastes and products rich in high molecular weight components such as cellulose, hemicelluloses, and lignin would deliver a good quantity of an adsorbent or biomass. Moreover, since these components are rich in functional groups, they are capable of forming different types of bonding (inter- or intramolecular), eventually creating a supramolecular structure capable of adsorbing the contaminants [19]. A literature survey shows that the waste biomasses can be used as they are (untreated or intact) [20,21] following a treatment (activated carbon produced physically or chemically) [22–24] or after conversion into nanoparticles decorated with some functional groups for water remediation [25,26].

*Aloe vera* (AV) is a succulent plant that belongs to the family Asphodelaceae. The origin of AV is the Arabian Peninsula and regions with tropical/subtropical climates. Yet, it is cultivated all over the world because of its medicinal value, where it has been perceived as a standout amongst the most conspicuous therapeutic plants with diverse applications. AV shell, for example, contains over 75 nutrients, 20 minerals, 18 amino acids, and 12 vitamins. Wound healing, anticancer, antioxidant, immunomodulatory cosmetic, and laxative effects are among the medical applications of AV. Food industry, nanotechnology, and the ecological applications are other applications of AV [27–30].

Developing a green adsorbent from AV waste for the removal of TY from contaminated water samples using an effortless, economical, and efficient method as well as optimization of the adsorption procedure and boosting the removal competency of AV leaves were the tasks undertaken in this methodology. As the purpose is to go "green", an experimental protocol that relies on "Design of Experiments" (DoE) was implemented. Plackett–Burman design (PBD) was the DoE of choice.

The novelty of this approach, therefore, stems from the conservation of method *greenness* implementing DoE, which in turn allows for the hoarding of chemicals and resources as well as high-quality data that can be handled with a high degree of confidence. Moreover, the approach followed herein for getting to the ideal adsorbent status (active biomass) did not involve activation using chemicals, and only the raw/burnt masses were used. Table 1 shows a comparison between the AV used for removal of different adsorbates and the AV synthesized in the current approach. Similarly, Table 1 shows a comparison between different adsorbents employed for the removal of TY and the currently used AV leaves [31–35].

Four variables thought to affect the adsorption process—dye concentration (DC), pH of dye solutions, stirring time (ST), and adsorbent dose (AD)—as well as two responses, percent removal (%R) and adsorption capacity ( $q_e$ ), were evaluated and optimized using PBD as a platform. Adsorbent characterization was performed using scanning electron microscopy (SEM) and Brunauer–Emmett–Teller (BET) surface area analysis. Thermogravimetric analysis (TGA), Fourier-transform infrared (FT-IR), Raman spectroscopy, as well as carbon hydrocarbon nitrogen analysis (CHN) were executed. Understanding the adsorption mechanism was achieved following study of the adsorption equilibrium and kinetics.

**Table 1.** Comparison between the performance of *Aloe Vera* (AV) prepared in the current approach and different AV adsorbents as reported in literature as well as the other adsorbents used to remove TY.

Adsorbent	Modification Method	Analytical Approach Used	Surface Area (m²/g)	Adsorbate	Adsorption Capacity (mg/g)	Percent Removal (%R)	Ref.
*Air dried AV (ADAV) *Thermally treated AV (TTAV300)	Please see the experimental section of the current approach	Multivariate analysis—Plackett-Burma design (PBD)	3.94 n 7.076	TY	55.25	93%	Current work
Aloe Vera leaves wastes-based sulfuric acid modified activated carbon (AV-SAC)	Leaves' waste was washed with deionized water (DIW) and dried at 150 °C for 24 h and then crushed. Particles were carbonized at 550 °C for 20 min. The carbonized sample was treated with sulfuric acid solution for 12 h. The suspension was filtered and washed with DIW. Finally, the modified activated carbon was dried at 105 °C for 12 h. Obtained AV- SAC was crushed and then sieved.	One-factor at a time (OFAT)	*ND	Aniline Methyl Orange (MO)	185.18 196.07	*ND	[31]
Aluminium sulphate- <i>Aloe vera</i> (ALAV) Magnesium sulphate- <i>Aloe vera</i> (MGAV)	Physical blending of AV with either Al or Mg (10%:90%) was performed at room temperature for 24 h.	2 <sup>2</sup> – Full factorial design	*ND	Methylene Blue (MB)	*ND	50–55% 60–70%	[32]
<i>Aloe vera</i> shell ash (ASA) - supported Ni <sub>0.5</sub> Zn <sub>0.5</sub> Fe <sub>2</sub> O <sub>4</sub> magnetic nanoparticles	AV shells were washed, dried in an air oven at 80 °C for 24 h, ground, sieved, and then carbonized at 700 °C for 2 h. Obtained ASA was mixed with the metal nitrates and dissolved in DIW. Egg-white extract was then mixed with the previous mixture at room temperature and stirring was applied for 30 min. Obtained sol-gel was evaporated at 80 °C. The precursor was ground and calcined in muffle furnace at 550 °C for 2 h.	OFAT	814.23	Ag (I)	243.9	98.30%	[33]
Aminated calcium lignosulfonate (ACLS)	Calcium lignosulfonate was modified with triethylene tetramine in the presence of glutaraldehyde as a crosslinking agent.	Orthogonal design	11.93	TY	190.1	91%	[34]
Polyaniline/Fe <sub>3</sub> O <sub>4</sub> (PANI/Fe <sub>3</sub> O <sub>4</sub> ) nanocomposite (NC)	Aniline polymerization was conducted in $H_2SO_4$ medium and in the presence of $Fe_3O_4$ nanoparticles with ammonium persulfate as an oxidant. Obtained NC were further washed and dried.	OFAT	*ND	ΤY	Dec-34	*ND	[35]

ND: Not Determined.

# 2. Experimental

#### 2.1. Materials and Reagents

All reagents and chemicals were of analytical grade and were used without further purification. Sodium hydroxide, sodium tetraborate-10-hydrate (Na<sub>2</sub>B<sub>4</sub>O<sub>7</sub>. 10H<sub>2</sub>O), and hydrochloric acid were purchased from Sigma-Aldrich (Darmstadt, Germany). Ultrapure water (18.2 MΩ) was used to prepare and dilute dye solutions. Titan yellow (TY) was purchased from Riedel-*de*haën (Seelze, NI, Germany). The pH of dye solutions was adjusted using a buffer prepared using a mixture of (Na<sub>2</sub>B<sub>4</sub>O<sub>7</sub>. 10H<sub>2</sub>O), 50 mM) and 0.1M NaOH or 0.1M HCl. Further adjustments of pH were done using small increments of 0.1M NaOH and 0.1M HCl. The color of the dye did not change over the investigated pH range.

#### 2.2. Instrumentation and Software

A UV-Vis spectrophotometer (Agilent diode-array, CA, USA) with 10 mm matched quartz cells was used to measure the absorbance of the samples. A centrifuge (Thermo Scientific, ST8 Benchtop, Waltham, MA, USA) was used for the separation of the supernatant. The pH of the prepared solutions was adjusted using a Jenway pH meter (Jenway, Staffordshire, UK).

The fully dried air-dried *Aloe vera* (ADAV) and the thermally treated *Aloe vera* (TTAV300) samples were sprayed with gold, and the surface morphology of the samples was investigated using scanning electron microscope (SEM, FEI, Quanta 200, Thermo Scientific<sup>™</sup>, Waltham, MA, USA) under an accelerating voltage of 30 kV and by using a standard secondary electron (SE) detector, while the distribution of elements on the surface was determined by energy dispersive X-ray (EDS) detector.

Fourier transform infrared radiation (FT-IR, Bruker Alpha, MA, USA) was used to recognize the functional groups on the surface of the prepared adsorbent. Spectra were collected in the range of 400–4000 cm<sup>-1</sup> with a resolution factor of 4 cm<sup>-1</sup>. The Raman spectrum of the ADAV and the TTAV300 were acquired in the range from 50–3500 cm<sup>-1</sup> using a DXRTM2 Raman microscope (Thermo Scientific<sup>™</sup>, Waltham, MA, USA) with a laser beam at 532 nm as the excitation source and 10 mW power. A thermal gravimetric analyzer (TGA, PerkinElmer-TGA400, Waltham, MA, USA) was employed to investigate the thermal stability of the prepared AV at a temperature range of 50–800 °C.

Analysis of the adsorbent's surface area was performed using a Micromeritics ASAP2020 Accelerated Surface Area and Porosimetry System (Micrometrics, Norcross, GA, USA). Degassing of samples was initially employed followed by investigating N<sub>2</sub> adsorption–desorption. Using the N<sub>2</sub> isotherms collected at 77 K and employing the Brunauer–Emmett–Teller (BET) equation, surface area was determined. Pore volume was established using the t-plots and the Barrett–Joyner–Halenda (BJH) equation.

Minitab<sup>®</sup>18 software was purchased from Minitab Inc., State College, Pennsylvania, USA. The software was used to make the list of experiments according to the selected design.

# 2.3. Preparation of Adsorbate

Water samples artificially contaminated with TY were prepared by dissolving the prerequisite amounts of TY in ultrapure water to a final concertation of 100 ppm stock solution. Serial dilutions of the dye solution were prepared in the same solvent, and the pH was adjusted to the desired value using borate buffer. Three calibration curves of TY at the tested pH levels (Table 2, low, central, and high) were constructed by measuring different concentrations of TY at 406 nm.

Independent Variables and Their Codes			Range	s and Levels				
					-1,	Low	0, Central	+1, High
	pН	(pH, <b>A</b> , pH ı	unit)		4	.00	8.00	12.00
	-	dose (AD, B,			100		300	500
Dye concentration (DC, $\vec{C}$ , ppm)				-	10		100	
Stirring time (ST, <b>D</b> , min.)					5	32.5	60	
Run		Coded levels	s of variable	s	Respons	e (Y1, %R)	Response (Y2, capacit	
	A <sup>a</sup>	B <sup>b</sup>	C c	D <sup>d</sup>	* Obs.	** Pred.	* Obs.	** Pred.
01	-1	+1	+1	-1	85.00	83.94	7.68	10.5
02	0	0	0	0	79.34	79.61	7.20	7.25
03	+1	+1	+1	-1	52.50	42.09	5.24	3.54
04	+1	+1	-1	+1	13.24	13.97	0.13	0.13
05	+1	+1	-1	+1	11.00	13.97	0.07	0.13
06	0	0	0	0	79.88	79.61	7.30	7.25
07	-1	-1	-1	-1	30.05	34.93	1.47	2.10
08	-1	-1	+1	+1	41.74	48.85	20.9	27.9
09	-1	+1	+1	+1	93.00	95.63	9.27	7.27
10	+1	-1	+1	-1	26.90	25.73	10.9	13.6
11	-1	-1	-1	+1	30.13	23.80	2.25	1.45
12	-1	+1	-1	-1	72.84	79.88	0.73	0.55
13	+1	-1	+1	+1	18.66	17.53	11.4	9.35
14	+1	-1	-1	-1	12.10	12.53	0.88	0.71

**Table 2.** PBD design matrix for the coded independent variables. Response is shown as observed and predicted %R of TY as well as the adsorption capacity ( $q_e$ ) using ADAV biomass.

\* Obs.: experimental values, \*\* Pred.: predicted values following the process of response transformation.

### 2.4. Adsorbent Preparation

#### 2.4.1. Air Dried Aloe Vera (ADAV)

Fresh, green AV leaves were collected from a yard/home garden in Doha-Qatar. The collected leaves were washed copiously with deionized water (DIW) several times to remove dust and impurities. The gel of AV leaves was carefully removed using a razorblade. Leaves were washed again with DIW several times. Clean, empty leaves were then cut into small pieces of approximately  $1 \times 1$  cm and left to dry in shade and in the presence of the air conditioner for 5 successive days. AV leaves were thereafter dried in an oven at 50–52 °C for 2 h. It is noteworthy to mention that the oven temperature was kept at less than 60 °C so as to eliminate water and to conserve the organic components simultaneously. The dried AV shells were then ground into very fine powder using a non-metallic electric grinder, sieved using a 1 mm mesh size, and divided into two portions. One portion was kept in the desiccator and was used for the adsorption experiments without any further treatment and was labeled as ADAV. The second portion underwent thermal treatment as in the subsequent section.

#### 2.4.2. Thermally Treated Aloe Vera (TTAV)

ADAV prepared under the aforementioned mild conditions was thermally treated at different temperatures ranging from 300–500 °C for 1 h and labeled as TTAV300, TTAV400, and TTAV500.

#### 2.5. Evaluation of Adsorption Capability of the Prepared Adsorbent(s)

Performance of the prepared adsorbents was assessed by comparing the %R and the  $q_e$  for the four adsorbents (ADAV, TTAV300, TTAV400, and TTAV500). The two quantities were measured using Equations (1) and (2). Assessments showed that ADAV and TTAV300 had the best performance with ADAV being superior, as shown in Table 3. The impact of the four variables on the performance of ADAV as an adsorbent was tested following the experimental setup shown in Table 2. Two sets of

15 mL centrifuge tubes were prepared. An amount of 30–150 mg of ADAV was added to each tube. The pH of TY solutions (10–100 ppm) was adjusted using borate buffer to the desired pH value (Table 2). Blanks were similarly prepared but without TY in the second set. The two sets were then centrifuged at 4200 rpm for the stirring time (ST) specified in Table 2. The absorbance of the supernatant was measured using UV-Vis spectrophotometer.

$$(\%R, Y1) = \frac{C_0 - C_e}{C_0} \times 100\%$$
(1)

$$(qe, Y2) = \frac{C_0 - C_e}{W} V \tag{2}$$

where  $C_e$  (mg L<sup>-1</sup>) is the concentration of TY aqueous solution at equilibrium,  $C_0$  (mg L<sup>-1</sup>) is the initial concentration of TY in aqueous solution, *V* is the volume of the solution in liters (L), and *W* is the weight of the ADAV in (g).

**Table 3.** A comparison between the adsorption capabilities of the four adsorbents in terms of %R and  $q_e$  (AD = 200 mg/50 mL, pH = 9, ST = 30 min, and DC = 100 ppm). The responses shown were calculated using Equations (1) and (2).

<b>Tested Adsorbent</b>	% Removal (%R)	Adsorption Capacity (q <sub>e</sub> , mg/g)
ADAV	79.44	11.92
TTAV300	70.85	10.63
TTAV400	37.07	5.561
TTAV500	31.08	4.662

# 2.6. Experimental Design

PBD was the design of choice in the current approach. Four independent variables were investigated at two levels (Table 2). Two responses were measured as a function of the four variables. In this itinerary, 14 base runs were generated in two blocks and with two added central points. The design matrix in terms of uncoded variables is shown in Table 2 [14,36,37].

# 2.7. Equilibrium and Kinetics Studies

For equilibrium studies, a stock solution of 1000 ppm TY was prepared. Further dilutions of the stock solution (5–400 ppm) were prepared in the same solvent, and the pH was adjusted to pH  $4.00 \pm 0.20$  using borate buffer solution. Equal quantities of ADAV (0.1 g  $\pm$  0.005) were added to 13 mL of the previously prepared solutions, and then the solution was shaken using the automatic shaker at 150 rpm for 2 h and then filtered. Absorbance of the filtrate was measured at 406 nm. The kinetics studies were conducted using 200 mL TY dye solution (200 ppm, pH 4.00  $\pm$  0.20) and ~1.5 g of ADAV with shaking. One sample was taken at time range around 1 min over a time span of 15 min.

# 3. Results and Discussion

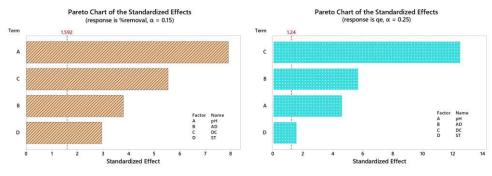
#### 3.1. Plackett–Burman Design (PBD)

As previously mentioned, the purpose of the current investigation was not only to recycle the agro-waste into a convenient adsorbent but also to augment the adsorption efficiency of the selected bio-sorbent without using extensive chemical or physical treatment. This objective could be attained via the application of PBD as a platform. The experimental setup was established following the scenario exhibited in Table 2. As a screening approach, PBD is commonly used to investigate the main effects, where they are usually deeply confounded with the factorial interactions [14,36,37]. Two responses were considered for the current investigation: %R and  $q_e$ . The target was to maximize each response.

# 3.1.1. Investigation of Statistically Significant Variables

A Pareto chart of the standardized effects was used to decide upon the statistically significant variables. As shown in Figure 2, the left panel shows that pH (A) was the most statistically significant variable affecting the %R compared to dye concentration (DC) (C) in the case of  $q_e$  (right panel). Analysis of variance (ANOVA) data are shown in Table 4. The data shown further confirm the findings of the Pareto chart with normal and half-normal probability plots (not shown). Statistical significance of the tested variables at 95.0 confidence intervals (95.0 CI) was estimated by F-test and *p*-value < 0.05. It is noteworthy to mention that a significant improvement in data quality was observed following response transformation by employing Box-Cox transformation [Equation (3)] together with stepwise analysis in the case of %R and forward selection in the case of  $q_e$  [38–40].

$$Y' = (Y_{\lambda} - 1)/\lambda \tag{3}$$



where Y' is the transformed response and  $\lambda$  is the transformation factor.

Figure 2. Pareto chart of standardized effects obtained following response optimization.

Response Source	DF *	% <b>R</b>			$q_e$				
	DI	Adj SS *	Adj MS *	<i>f-</i> Value	<i>p</i> -Value	Adj SS *	Adj MS *	F-Value	<i>p</i> -Value
Model	5	7.33116	1.46623	28.99	0.000	37.9965	7.5993	45.25	0.000
Linear	4	5.87250	1.46813	29.03	0.000	35.6767	8.9192	53.11	0.000
pН	1	3.15187	3.15187	62.32	0.000	3.5806	3.5806	21.32	0.002
ÂD	1	0.72725	0.72725	14.38	0.005	5.4182	5.4182	32.26	0.000
DC	1	1.55138	1.55138	30.67	0.001	26.2590	26.2590	156.35	0.000
ST	1	0.44201	0.44201	8.74	0.018	0.4190	0.4190	2.50	0.153
Curvature	1	1.45866	1.45866	28.84	0.001	2.3198	2.3198	13.81	0.006
Error	8	0.40463	0.05058			1.3436	0.1679		
Lack-of-Fit	6	0.38743	0.06457	7.51	0.122	1.1124	0.1854	1.60	0.432
Pure Error	2	0.01720	0.00860			0.2312	0.1156		
Total	13	7.73579				39.3401			
		* DD 1	C C 1	00 ·	C	N/G :	(		

Table 4. Results of ANOVA for the transformed responses.

\* DF is degrees of freedom; SS is sum of squares; MS is mean of squares.

Equations (4) and (5) are the regression models that describe the relation between the measured responses and the investigated variables and accordingly can be used to predict the R and the  $q_e$ . These models were obtained following response transformation.

$$\ln(Y1, \ \%R) = 3.898 - 0.1281 \text{ pH} + 0.001231 \text{ AD} + 0.00799 \text{ DC} - 0.00698 \text{ ST} + 0.922 \text{ Ct. Pt.}$$
(4)

$$\ln(Y2, q_e) = 1.331 - 0.1366 \text{ pH} - 0.003360 \text{ AD} + 0.03287 \text{ DC} - 0.00680 \text{ ST} + 1.163 \text{ Ct. Pt.}$$
(5)

To test the applicability of the proposed models, the experimental values of the two responses—%R and  $q_e$ —were compared to the model predicted values (Table 2). Results show a good agreement between both. The values of the coefficient of determination (R<sup>2</sup>) were high enough (0.9477 and 0.9658 for %R and  $q_e$ , respectively) to indicate the model linearity. The potential of the suggested models to foresee new responses was spotted from the high values of R<sup>2</sup> (predicted) observed following response transformation (0.8463 and 0.8996 for %R and  $q_e$ , respectively). As shown from the two regression equations, while the adsorbent dose (AD) was positively impacting the %R, it negatively impacted the  $q_e$ . The direction of the rest of the effects was the same for both responses.

# 3.1.2. Contour and Surface Plots

The 2D and the 3D plots for the response surface versus the significant factorial interactions are shown in Figure 3. The left panel is a sample 2D plot that shows the factorial interactions, where pH and DC are represented on the X- and the Y-axes, while the %R is denoted by the contour lines. As shown on the graph and as per the associated legend, the darkest green zone signifies a zone with the maximum response (greater than 90% removal of TY). The right panel of Figure 3 displays a similar liaison; however, it is shown in the 3D arrangement where the measured %R is represented on the Z-axis. As shown, the maximum %R (exceeding 75%) would be attained by employing a DC between 50–70 ppm and a pH value of 4–7.

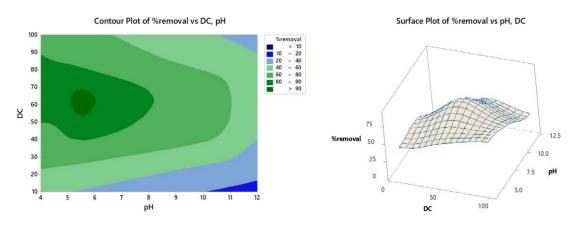


Figure 3. Sample contour (2D) and surface (3D) plots for the relationship between DC, pH, and %R.

Optimum conditions for maximizing each response (individually) were obtained using the Minitab response optimizer. Data obtained show that maximum %R was achieved with a desirability of d = 1.0000 and using a factorial combination of pH = 4, AD = 500 mg/50 mL, DC = 100 ppm, and ST = 5 min. This high value of desirability function (d) indicates that the proposed factorial combination was in favor of maximum %R. The increase in %R with the increase of AD and DC implies that adsorption sites were not getting saturated, even with higher dye concentrations. Maximum  $q_e$ , however, was obtained using a combination of pH = 4, AD = 100 mg/50 mL, DC = 100 ppm, and ST = 5 min with a desirability of d = 1.0000. This can be explained since  $q_e$  could be maximized using the lowest AD and the highest DC, as shown in Equation (2).

# 3.2. Adsorbent Characterization

#### 3.2.1. TGA Analysis of ADAV

Thermogravimetric analysis of ADAV was performed under  $N_2$  and with a heating rate of 10 °C/min. As shown in Figure 4, the weight loss in the samples occurred over three steps:

1. Loss of adsorbed water and solvent molecules at 25–150 °C, while the crystalline water was lost at 200 °C, which represented 6.5% of the sample;

- 2. In this step, more than 51% of the sample was decomposed between (200–600 °C), and there were four peaks—two major peaks at 273.2 °C and 316.3 °C, and two other small peaks at 370.6 °C and 436.8 °C. Therefore, at least four decomposition mechanisms could be involved [41];
- 3. The third step (600–800 °C) represented nearly 9.9% of the sample and could be attributed to the carbonization of polymeric material [42].

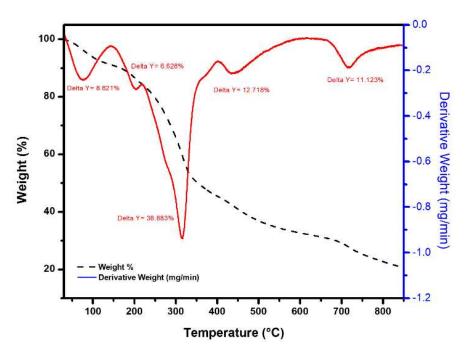


Figure 4. TGA/dTGA analysis of ADAV sample.

# 3.2.2. FT-IR Analysis of ADAV and TTAV300

The FT-IR spectra of ADAV and TTAV300 are shown in Figure 5. The revealed spectrum of ADAV exhibited a broad band positioned at 3300 cm<sup>-1</sup>, which was attributed to the stretching vibration of the –OH groups that characterized the carbohydrate monomers together with mannose and uronic acid. Moreover, the spectrum showed two absorption bands at 2916 and 2850 cm<sup>-1</sup>, which could be assigned to the symmetrical and the asymmetrical C–H stretching. The absorption band at 1731 cm<sup>-1</sup> was characteristic for C=O stretching and confirmed the presence of a carbonyl moiety. The absorption peak at 1586 cm<sup>-1</sup> may have been associated with the asymmetrical and the symmetrical –COO<sup>-</sup> stretching of carboxylate compounds. Furthermore, the two absorption peaks at 1250 and 1153 cm<sup>-1</sup> corresponded to the presence of C–C stretching and C–O–C stretching of aliphatic ether, respectively. The absorption peak at 1019 cm<sup>-1</sup> may have been due to C–O stretching associated with rhamnogalacturonan, a side-chain component of pectins [43,44].

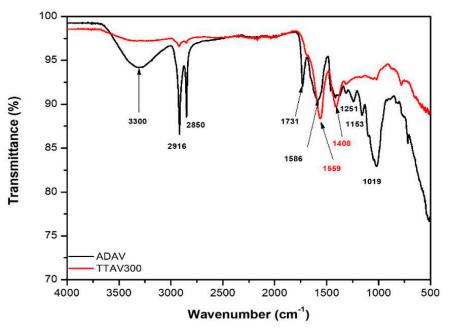


Figure 5. FT-IR spectra of ADAV and TTAV300.

On the other hand, the spectrum of TTAV300 showed the presence of some peaks similar to those of ADAV, however, with a lower absorption intensity due to the thermal treatment. Moreover, two strong absorption peaks appeared at 1559 and 1409 cm<sup>-1</sup>, which could be attributed to the presence of –COO<sup>-</sup> stretching of carboxylate compounds in the thermally treated sample.

# 3.2.3. CHN Analysis

Data presented in Table 5 confirmed the findings of the FT-IR analysis. As shown, the percentage of hydrogen decreased from 5.34% to 4.22% following the thermal treatment. This could have been caused by the loss of water during the heating of samples. On the other hand, the percentage of nitrogen was almost the same in both adsorbents. The percentage of carbon, as expected, increased from 35% to 53% in the thermally treated sample, and this also confirmed the presence of carbon–oxygen and carbon–nitrogen functional groups in both adsorbents.

Adsorbent	N (%)	C (%)	Н (%)
ADAV	0.96	35.79	5.34
<b>TTAV300</b>	1.10	52.73	4.22

Table 5. Carbon hydrocarbon nitrogen (CHN) analysis of the prepared adsorbents.

#### 3.2.4. Raman Analysis of ADAV and TTAV300

Raman spectra of ADAV and TTAV300 are shown in Figure 6. The spectrum of ADAV did not show any peaks, probably because carbon exists as an organic matter. On the other hand, the spectrum of TTAV300 had a clear D- band, which appeared at 1351 cm<sup>-1</sup>, and a clear G- band at 1585 cm<sup>-1</sup>. These two peaks were distinctive in the spectra of all carbonaceous materials, and their arrangement was the same as graphene oxide [45].

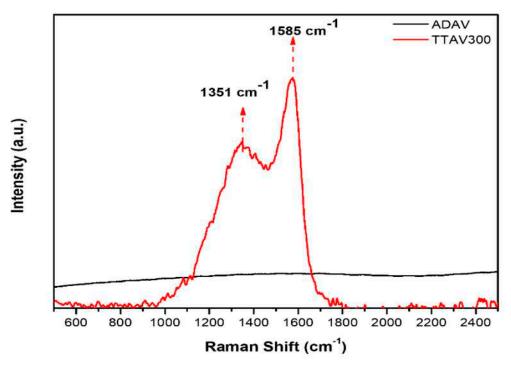
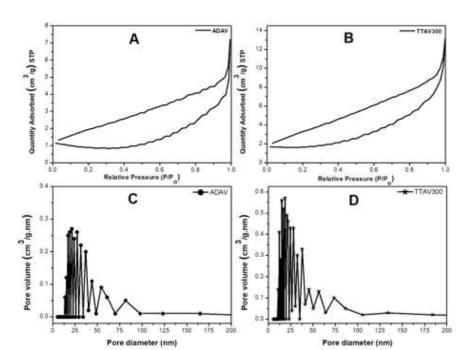


Figure 6. Raman spectra of ADAV and TTAV300.

The properties of the carbon lattice, including defects and sizes, were displayed by the D- band; however, it did not display the chemical conformation of the carbon materials. The wavelength of the laser beam mainly affected the position of the D- band. On the other hand, the G- band identified the stretching of C–C for the  $sp^2$  system [46]. Obtained data confirmed the formation of activated carbon in the case of TTAV300, and this was confirmed by the presence of the two bands (G- and D-).

# 3.2.5. BET Analysis

The surface areas (SAs) of ADAV and TTAV300 were measured, as shown in Figure 7. The obtained data showed that Langmuir SAs were 3.940 and 7.076 m<sup>2</sup>/g for ADAV and TTAV300, respectively. Besides, the single point total pore volumes were 0.011197 and 0.020461 cm<sup>3</sup>/g for ADAV and TTAV300 samples, respectively. Moreover, the single point average pore radii were found to be 98.6°A for ADAV and 68.4°A in the case of TTAV300. The obtained data indicated that the thermal treatment process of the AV samples did not have a significant impact on the surface area and the pore size of the prepared samples and, as a result, the adsorption efficiency of TTAV samples was not affected by thermal treatment [45,46]. On the other hand, using the International Union of Pure and Applied Chemistry (IUPAC) definition for mesoporous and macroporous materials, the pore diameter patterns of both ADAV and TTAV300 showed two types of pores: mesoporous (less than 50 nm in diameter) and macrospores (greater than 50 nm in diameter). These types of pore systems serve to increase the surface area of the adsorbent and hence its adsorption capacity [16].



**Figure 7.** Nitrogen adsorption–desorption isotherms at 77 K for ADAV (**A**) and TTAV300 (**B**) and the relation between pore volume and diameter for both ADAV (**C**) and TTAV300 (**D**).

# 3.2.6. SEM and EDX Analyses

Surface morphologies of ADAV and TTAV300 were investigated using SEM. Micrographs shown in Figure 8A,B indicate that ADAV and TTAV300 contained a number of pores with different radii. These data confirmed the loss of some volatile and organic matter at 300 °C and the formation of carbonaceous mass with an advanced pore structure. These findings are in alignment with the BET surface area and the TGA analyses of other thermally treated adsorbents [14–16]. On the other hand, energy dispersive X-ray spectroscopy (EDX) analysis was used to study the elemental compositions of ADAV and TTAV300. Obtained results show that the TTAV300 was mainly composed of carbon (92.75%) in comparison to 72.55% for ADAV (Figure 8C,D). Other elements such as Na, Mg, P, Cl, Ca, and Ca were also present, and their total concentration was less than 2%. These results confirm the formation of carbonaceous material following the thermal treatment.

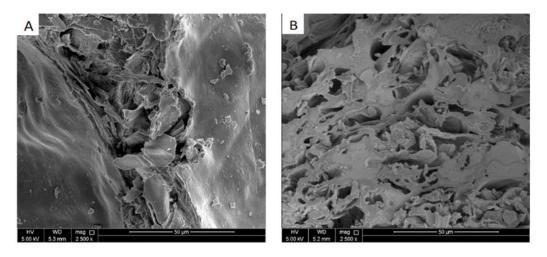
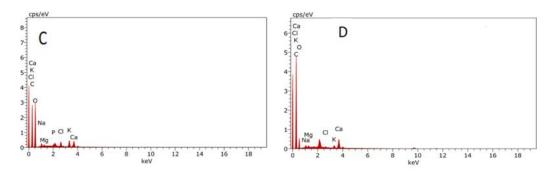


Figure 8. Cont.



**Figure 8.** SEM micrographs of ADAV (**A**), TTAV300 (**B**), as well as energy dispersive X-ray spectroscopy (EDX) analysis of ADAV (**C**) and TTAV300 (**D**).

#### 3.3. Equilibrium and Kinetics Studies

Preliminary studies have shown that the ADAV is more powerful compared to the TTAV samples in biosorption removal of TY. Therefore, the equilibrium and the kinetics studies were conducted on the ADAV. Equilibrium and kinetic studies play an imperative role in assessing the efficiency of adsorbents. The adsorption isotherms are very useful in determining the maximum quantity ( $q_m$ ) that can be adsorbed, the pattern of adsorption on the surface of the biomass, the interaction between adsorbate and adsorbent's surface, and whether it is chemisorption or physisorption. On the other hand, the kinetic studies can be used to determine the rate of adsorption, the thickness of the formed layer around the sorbent, and whether the reaction is controlled by diffusion or adsorption mechanisms. The following sections reveal a discussion on the adsorption isotherms as well as the kinetics of the adsorption of TY on ADAV.

#### 3.3.1. Equilibrium Isotherms

The adsorption isotherm expresses the specific relationship between the degrees of accumulation on the adsorbent surface and the concentration of sorbate at constant temperature [18]. Four isotherm models were used to investigate the adsorption process of TY on ADAV. These models included Langmuir, Freundlich, Temkin, and Dubinin–Radushkevich (DR) isotherm.

Langmuir isotherm explains the single-layer homogeneous adsorption on a surface, as shown in Figure 9A and Table 6. The basic characteristic of Langmuir equation is shown in Equation (6).

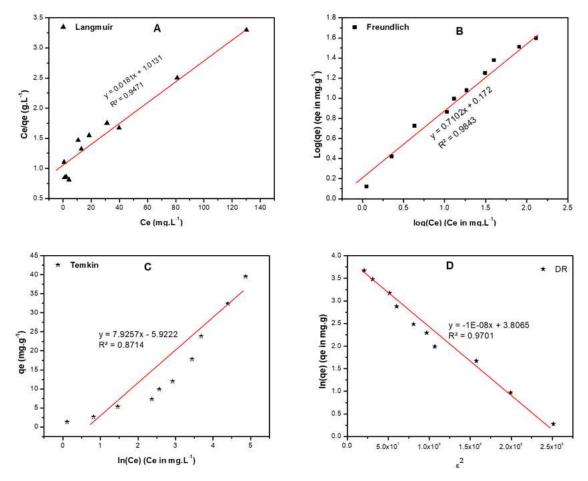
$$q_e = \frac{q_m K_L C_e}{1 - K_L C_e} \tag{6}$$

where  $q_m$  is the monolayer adsorption capacity and  $K_L$  is the Langmuir equilibrium coefficient. In addition, the Langmuir equation can be stated using the dimensionless equation:

$$R_L = \frac{1}{1 - K_L C_0}$$
(7)

where  $R_L$  is separation factor, and  $C_0$  (mg/L) is the initial concentration. Based on previous reports [47,48], the adsorption favorability is dependent on the value of  $R_L$ . In general, if  $R_L$  is >1, the adsorption process is unfavorable, if  $R_L = 1$ , adsorption is linear, and with a value between 0–1, the adsorption is favorable (occurs spontaneously); if the value = 0, adsorption is irreversible. The calculated  $R_L$  values in the current investigation were <1 and >0, implying that the adsorption process was spontaneous with maximum adsorption ( $q_{max}$ ) = 55.25 mg/g.





**Figure 9.** Adsorption isotherms of TY on ADAV including (**A**) Langmuir, (**B**) Freundlich, (**C**) Temkin, and (**D**) Dubinin–Radushkevich (DR).

Table 6. General and linearized equations of Langmuir, Freudlich, Temkin, and Dubinin–Radushkevich
isothems beside their parameters for the adsorption of TY on ADAV.

Isotherm	Equations (Generalized/Linearized Forms)	Parameters	Value
	$q_e = rac{q_m K_L C_e}{1 - K_L C_e}$	$q_m (\mathrm{mg/g})$	55.25
Langmuir		$K_L$ (L·mole <sup>-1</sup> )	0.017
	$rac{C_e}{q_e} = rac{1}{q_m  K_L} + rac{C_e}{q_m}$	$\mathbb{R}^2$	0.947
	$q_e = K_F C_e^{\frac{1}{n}}$	$rac{1}{n} K_F$	0.710
Freundlich	$\log(q_e) = \log(K_F) + \left(\frac{1}{n}\right)\log(C_e)$	K <sub>F</sub> (mole/g)(L/mole) <sup>1/n</sup>	1.486
	$\log(q_e) = \log(\kappa_F) + (\frac{1}{n})\log(C_e)$	$R^2$	0.984
	$q_e = \frac{RT}{h_T} \ln(A_T C_e)$	$b_T$ (J/mole)	311.014
Temkin	$q_e = \frac{RT}{h_r} \ln(A_T) + \frac{RT}{h_r} \ln(C_e)$	$A_T$ (L/mole)	0.473
	$q_e = \frac{1}{b_T} \ln(A_T) + \frac{1}{b_T} \ln(C_e)$	$\mathbb{R}^2$	0.8714
	$\ln(q_e) = \ln(q_m) - \beta \epsilon^2$	β	$1 \times 10^{-8}$
DB	$\mathbf{PT}(1 + 1)$	E (kJ/mole)	7.07
DR	$\epsilon = RT \left( 1 + rac{1}{C_e}  ight)$	$q_s (\mathrm{mg/g})$	44.98
	$E = \frac{1}{\sqrt{2\beta}}$	R <sup>2</sup>	0.9701

The Freundlich isotherm used to describe heterogeneous surface energies is given by the equation:

$$q_e = K_F C_e^{\frac{1}{n}} \tag{8}$$

Here, the  $C_e$  is the equilibrium concentration of TY (mg·L<sup>-1</sup>),  $q_e$  is the amount of TY adsorbed/unit mass (mg·g<sup>-1</sup>), and  $K_F$  (mole·g<sup>-1</sup>)(L·mole<sup>-1</sup>)<sup>1/n</sup> and1/n are Freundlich coefficients (Figure 9B and Table 6). The Freundlich plot (Figure 9B) showed a good fit with a coefficient of determination,  $R^2 = 0.9843$  with 1/n = 0.71 and n = 1.487. Therefore, the adsorption potential (A = nRT) = 4.12 kJ, thus any TY with a potential energy less than 4.12 kJ would be adsorbed on the surface of ADAV, and reactions would tend to be irreversible and favorable. These data confirm that the Freundlich isotherm was more applicable to study the adsorption process of TY on ADAV.

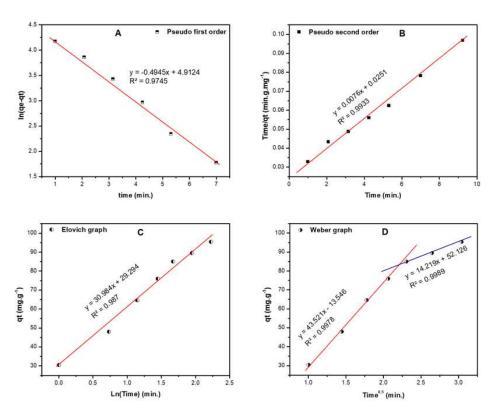
The Temkin isotherm (Figure 9C) gives an idea about the adsorbate–adsorbent interaction by the factor that describes the fact that the heat of adsorption of all the molecules in the layer decreases linearly with the adsorbent–adsorbate interactions. The data shown in Table 6 shows that the  $R^2$  value was 0.8714, signifying that this isotherm was not applicable in this study.

Finally, the DR isotherm (Figure 9D) is usually applied to differentiate between the adsorption mechanisms on a heterogeneous surface. The physical adsorption occurred when the free energy was <8.0 kJ/mol, while the chemical adsorption occurred when the free energy was >8.0 kJ/mol. The free energy for adsorption of TY on ADAV was physical, as shown in Table 6, where the E equals 7.07 KJ.

#### 3.3.2. Kinetic Studies

Four kinetics models, namely, pseudo-first order (PFO), pseudo-second order (PSO), Elovich, and Weber–Morris, were applied to simulate the kinetics of the adsorption of TY onto ADAV. Figure 10A,B show the plots of  $\ln(q_e - q_t)$  and the time/ $q_t$  vs time for the PFO and the PSO kinetic models, respectively. The calculated parameters of the two models are listed in Table 7. By comparing the coefficient of determination values (R<sup>2</sup>) of the two models, it could be concluded that the experimental data were in good agreement with the PSO model (R<sup>2</sup> = 0.993) [49,50]. Therefore, the reaction is represented as follows:

$$\Gamma Y + ADAV \stackrel{\kappa}{\to} \{TY - ADAV\} \tag{9}$$



**Figure 10.** (**A**) First order, (**B**) second order, (**C**) Elovich, and (**D**) intra-particle diffusion (Weber) curves of adsorption of TY on ADAV.

Models	Parameter	Val	lue	
Pseudo-first order (PFO)	$K_1 ({\rm min}^{-1})$	0.4	94	
$\ln(q_e - q_t) = \ln(q_e) - k_1 t$	$q_e (mg/g)$	135	5.9	
$\operatorname{III}(q_e - q_t) = \operatorname{III}(q_e) - \kappa_1 \iota$	R <sup>2</sup>	0.974		
Pseudo-second order (PSO) $\frac{t}{t} = \frac{1}{t} + \frac{1}{t}$	$K_2$ $(g \cdot mg^{-1} \cdot min^{-1})$	2.3 ×	10 <sup>-3</sup>	
$rac{t}{qe}=rac{1}{k_2q_e^2}+rac{1}{q_e}\mathbf{t}$	$q_e$ (mg/g)	131	.57	
Where $K_2$ is rate constant $(g \cdot mg^{-1} \cdot min^{-1})$	$R^2$	0.993		
Elovich equation is $q_t = \beta \ln(\alpha\beta) + \beta \ln(t)$ and is used to	α	0.083		
predict the sorption mechanism, where $q_t$ is adsorbed quantity at time t, while $\alpha$ and $\beta$ are initial sorption concentration rate	В	30.984		
(mg·g <sup><math>-1</math></sup> .min <sup><math>-1</math></sup> ) and desorption constant (g/mg), respectively.	R <sup>2</sup>	0.987		
Weber–Morris intraparticle diffusion model is used to study	K <sub>I</sub>	43.52	14.22	
the formed layers around the adsorbent and the rate-controlling step, which is expressed as $q_t = K_I t^{0.5} + C$ ,	С	-13.46	52.13	
where $K_I$ is intraparticle diffusion rate constant (mg·g <sup>-1</sup> ·min <sup>-0.5</sup> ), and C is boundary thickness effect.	$\mathbb{R}^2$	0.998	0.999	

Table 7. The kinetics study results corresponding to Figure 10.

Consequently, the rate of the reaction equaled k[TY][ADAV], indicating that the adsorption rate depended on both the adsorbate and the adsorbent.

According to the Elovich model shown in Figure 10C, the high  $R^2$  values (0.987) show that the desorption rate was higher than the initial sorption. From the Weber–Morris intraparticle diffusion model (Figure 10D), it is clear that there were two regions; the first region had almost no boundary layer, the diffusion rate was very fast, and the negative sign indicated the penetration of TY to the surface of ADAV. After a while, a TY layer formed around the particles, preventing further penetration of TY with a boundary layer equal to 52.13 mg/g, which was close to the  $q_{max}$  calculated by the Langmuir isotherm. Therefore, the reaction in the beginning was controlled by diffusion but later on by adsorption.

# 4. Conclusions

As per the introduced data, *Aloe vera* (AV) leaves could be recycled into a low-cost, non-conventional green adsorbent. The removal efficiency as well as the adsorption capacity of ADAV proved to be better compared to the thermally treated samples (TTAV). Optimization of the adsorption process was done using a contemporary approach (PBD). Two responses (%R and  $q_e$ ) were measured as functions of four variables (pH, AD, DC, and ST). The target was set to maximize both responses. Two mathematical models were therefore developed to predict the removal efficiency and the adsorption capacity. As per the regression models, increasing the concentration of TY enhanced both responses, contrary to pH and ST. Moreover, while the sorbent dosage had a positive impact on the %R, it had a negative influence on  $q_e$ .

CHN analysis showed that the carbon content increased significantly with the thermal treatment from 35.8% to 52.7%. Increasing of the carbon content was proven by using EDX analysis. Raman spectra showed the conversion of ADAV into a carbonaceous material similar to graphene oxide, which was also confirmed by FT-IR, which showed the disappearance of O–H and C–H groups and the presence of the C=O in TTAV300. SEM micrographs showed a formation of porous material by thermal treatment.

Equilibrium studies revealed that the adsorption of TY on ADAV was well described by the Freundlich isotherm, where n > 1, and the adsorption was favorable. The maximum adsorbed quantity = 55.25 mg/g, as calculated by Langmuir isotherm. Moreover, the calculated adsorption energy (7.07 KJ) by the Dubinin–Radushkevich isotherm signified that the adsorption occurred through physisorption. The kinetic study revealed that the reaction was second order reaction and was controlled at the beginning by a diffusion reaction followed by adsorption when the boundary layer = 52 mg/g.

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administration, M.E.-A.; Resources, A.A.I.; Software, M.E.-A.; Supervision, M.E.-A.; Validation, M.E.-A., A.S.E.-S.; Visualization, M.E.-A.; Writing—original draft, M.E.-A.; Writing—review & editing, M.E.-A., A.S.E.-S. and A.A.I.

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