



## Uncovering the immunological repertoire of the carpet shell clam *Ruditapes decussatus* through a transcriptomic-based approach

Frederico M. Batista<sup>a,b,c,\*</sup>, Allison M. Churcher<sup>a</sup>, Manuel Manchado<sup>d</sup>, Alexandra Leitão<sup>b,e</sup>, Deborah M. Power<sup>a</sup>

<sup>a</sup> Centro de Ciências do Mar (CCMAR), Universidade do Algarve, Campus de Gambelas, 8005-139 Faro, Portugal

<sup>b</sup> Instituto Português do Mar e da Atmosfera, Estação Experimental de Moluscicultura de Tavira, Portugal

<sup>c</sup> Department of Biosciences, College of Science, Swansea University, Swansea, SA2 8PP, Wales, United Kingdom

<sup>d</sup> IFAPA Centro El Toruño, Junta de Andalucía, Camino Tiro Pichón s/n, 11500 El Puerto de Santa María, Cádiz, Spain

<sup>e</sup> Environmental Science Center (ESC), Qatar University, P. O. Box: 2713 Doha, Qatar

### ARTICLE INFO

#### Keywords:

*Ruditapes decussatus*  
Transcriptome  
Immune genes  
Toll-like receptors

### ABSTRACT

The grooved carpet-shell clam *Ruditapes decussatus* is native to the Northern Atlantic and Mediterranean Sea and has a high commercial value. It is one of the main native bivalve species cultured in Europe. The main objective of the present study was to gain further insights into the immunological repertoire of *R. decussatus* through a transcriptomic approach. Pooled mantle samples of eight *R. decussatus* individuals were sequenced using Illumina platform. A total of 67 132 contigs with more than 800 bp were obtained. Manual annotation of these contigs revealed 146 immune-related genes. The gene families in which the highest number of immune-related genes was observed were: C1q domain-containing proteins (63), tumor necrosis factors (15) and toll-like receptors (TLRs, 10). A total of 5 359 putative single nucleotide polymorphisms (SNPs) were identified in the 146 immune-related genes. The density of SNPs ranged between 0.04 and 7.92 SNPs/100 bp. The highest and the lowest SNP density were observed in genes of the C1q domain-containing protein family. Due to the importance of TLRs in innate immunity, we focused our attention on these membrane receptors. Ten TLRs were identified based on protein domain organization. Phylogenetic analysis revealed that *R. decussatus* TLRs were diverse and only 3 showed orthology with TLRs of known immune functions in other bivalve species. Moreover, our analysis suggests that lineage restricted-expansions of TLRs occurred in all mollusc taxa analysed including in venerids.

### 1. Introduction

Aquaculture is one of the fastest-growing food producing sectors in the world and has a high social and economic importance. According to the Food and Agriculture Organization (FAO, 2014), the production of mollusc bivalves represented approximately 22% of world aquaculture production. The carpet shell clam *Ruditapes decussatus* is one of the main native bivalve species produced in Europe, with Portugal leading production in the world with a total production of 2 320 tonnes in 2012 (FAO, 2014).

Recently, there has been a considerable increase in knowledge about mollusc bivalve immunology motivated by the negative impact of infectious diseases on their production. High mortality rates have been observed in *R. decussatus* due to diseases caused by the protozoan parasite *Perkinsus olseni* and bacteria of the genus *Vibrio* (for review see Ruano,

Batista, & Arcangeli, 2015). As in other invertebrates, mollusc bivalves lack a specific immune response and associated immunological memory, and rely on the innate immune system. Several immune-related genes have been identified involved in the cellular and humoral innate response of bivalves, such as recognition of pathogen-associated molecular patterns, activation of signal pathways, haemocyte mediated reactions and production of immune effectors (Schmitt, Gueguen, Desmarais, Bachère, & de Lorgeril, 2010).

The advent of next-generation sequencing (NGS) has greatly accelerated studies in the field of immunology of mollusc bivalves by providing insights into the genetic diversity and transcription of multiple genes simultaneously (Leite et al., 2013; Moreira et al., 2015; Zhang, Guo, Litman, Dishaw, & Zhang, 2015). A better understanding of the immune system in *R. decussatus* will provide the basis for the development of strategies to mitigate the impact of infectious diseases affecting

\* Corresponding author. Department of Biosciences, College of Science, Swansea University, Swansea, SA2 8PP, Wales, United Kingdom.

E-mail address: [fmbatista@yahoo.com](mailto:fmbatista@yahoo.com) (F.M. Batista).

<https://doi.org/10.1016/j.aaf.2018.03.003>

Received 9 December 2017; Received in revised form 4 February 2018; Accepted 21 March 2018

Available online 9 April 2018

2468-550X/© 2018 Shanghai Ocean University. Published by Elsevier B.V. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

this species but can also be of relevance for other bivalves of commercial interest. The main objective of the present study was to map the immunological repertoire of *R. decussatus* using a transcriptomic approach. The importance of toll-like receptors in innate immunity led us to focus our attention on the identification and characterization of these immune-related membrane receptor genes.

## 2. Material and methods

### 2.1. RNA isolation, sequencing and de novo assembly

Total RNA was isolated from a piece of mantle edge from eight *Ruditapes decussatus* individuals (wild clams collected in Portugal) using the RNeasy Mini kit (QIAGEN). A Bioanalyzer 2100 (Agilent technology) was used to assess the integrity of the RNA isolated. Total RNA from the 8 individuals was pooled and sequenced using an Illumina HiSeq 2000 platform at the Centre Nacional d'Anàlisi Genòmica (Barcelona, Spain) as reported in Benzekri et al. (2014). After adaptor removal and quality filtration (minimum Phred score of 20) the quality of the reads was confirmed with FastQC (Bioinformatics, 2011) (see Fig. S1). After removing unpaired reads a total of 63 103 312 paired end reads ( $2 \times 76$  bp) were obtained. The GC content was 37% and the mean length and Phred quality were 76 bp and 36.3, respectively. Contigs were then obtained by *de novo* assembly using Trinity (release 2012-06-08) (Grabherr et al., 2011) with default parameters.

### 2.2. Annotation

The Annocript pipeline (Musacchia, Basu, Petrosino, Salvemini, & Sanges, 2015) was used for annotation of *R. decussatus* contigs with more than 800 bp (a total of 67,132 contigs) using BLASTX against TrEMBL/UniRef and SwissProt (cut-off *E*-value  $\leq 10^{-5}$ , threshold 18, wordsize 4), RPSBLAST against CDD profiles (cut-off *E*-value  $\leq 10^{-5}$ ), BLASTN against Rfam and rRNAs (cut-off *E*-value  $\leq 10^{-5}$ ). The Gene Ontology (GO) terms for cellular component, biological process and molecular function were obtained using Annocript. Conserved domains of immune-related genes were manually checked using SMART (Simple Modular Architecture Research Tool) (Letunic, Doerks, & Bork, 2014) and transmembrane regions and signal peptide prediction were performed using Phobius (Käll, Krogh, & Sonnhammer, 2004) and SignalP 4.1 (Petersen, Brunak, von Heijne, & Nielsen, 2011), respectively.

### 2.3. Characterization of toll-like receptors

Contigs were identified as TLRs if they showed at least one leucine rich repeat (LRR) domain, a single transmembrane (TM) region and one C-terminal Toll interleukin-1 receptor (TIR) domain. Multiple alignments were carried out using Clustal Omega software (Li et al., 2015) with amino acid sequences from the TIR domain of *R. decussatus* TLRs identified in the present study, 23 TLRs of *Mytilus galloprovincialis* (Toubiana et al., 2013), 2 TLRs of *Cyclina sinensis* (Ren, Pan, Pan, & Bu, 2016), 12 TLRs of *Lottia gigantea* (Simakov et al., 2012), 9 TLRs of *Crassostrea gigas* (Zhang et al., 2012, 2013, 2011; He et al., 2015) and 1 TLR of *Chlamys farreri* (Qiu, Song, Xu, Ni, & Yu, 2007). The best amino acid substitution model was chosen using MEGA6 software (Tamura, Stecher, Peterson, Filipitski, & Kumar, 2013). Maximum Likelihood (ML) analysis were then conducted using MEGA6 and an LG substitution model and gamma distribution, five discrete rate categories and 1000 bootstrap replicates. Graphical representation of the phylogenetic tree was performed with iTOL (Letunic & Bork, 2016). The RNA-seq reads of *R. decussatus* were aligned with the 146 immune-related genes using Burrows-Wheeler Aligner (BWA)-MEM (Li, 2013). BWA-MEM was used with default settings, under which ambiguous alignments were randomly assigned to one of their possible positions of the immune-related genes. Samtools were then used to perform the SNP calling, which involved converting the SAM files into BAM files and sorting and indexing the BAM files. The

samtools mpileup was used to collect summary information from the BAM files and compute the likelihood of the data and then convert it into BCF format. The bcftools were then used to filter the SNPs and generate a multi-sample variant call format (VCF) file. Default parameters were used for both samtools mpileup and bcftools.

## 3. Results and discussion

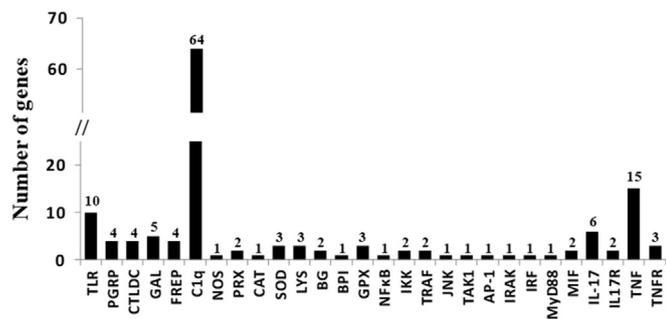
### 3.1. Transcriptome characterization

A total of 173,927 contigs were assembled using Trinity with 67 132 longer than 800 bp. This subset of transcripts were selected for further characterization (mean sequence length of 2 349 bp). Successful Blast searches with at least one significant match ( $E < 10^{-5}$ ) against UniProtKB/Swiss-Prot and UniProtKB/TrEMBL databases were achieved for 29 300 and 39 454 contigs, respectively, and at least one blast result was obtained for 40 839 out of the 67 132 contigs (longer than 800bp). The number of non-coding contigs was estimated to be 2 323. The functional categories distribution of the 40 839 annotated contigs using GO terms is shown in Fig. S2 of the supplementary data. We observed 3 913, 1 510 and 757 GO functional categories belonging to biological process (BP), molecular function (MF) and cellular component (CC), respectively. These results suggest that the transcripts observed in the mantle of *R. decussatus* cover a wide range of biological processes. In the BP category, the “signal transduction” (GO:0007165) and the “transcription, DNA-dependent” (GO:0006351) terms were the most abundant with 6.5 and 5.2%, respectively. In the MF category, the most abundant terms were “ATP binding” (GO:0005524), “zinc ion binding” (GO:0008270) and “metal ion binding” (GO:0046872) with 16.6, 14.0 and 7.4%, respectively. The “integral to membrane” (GO:0016021) term was by far the most common one with 39.9% in the CC category.

### 3.2. Identification and characterization of immune-related genes

A total of 146 immune-related genes were identified in the transcriptome of the mantle of *R. decussatus* (for details see Table S1 of supplementary data). Thirty eight percent of the BLAST hits were obtained with members of the family Veneridae followed by 27% with members of the family Ostreidae (see Fig. S3 of supplementary data). The gene families in which the highest number of immune-related genes was observed were: C1q domain-containing proteins (63), tumor necrosis factors (15) and toll-like receptors (10) (Fig. 1). The low number of immune-related genes may be explained by the fact that the individuals used in the present study were not challenged with pathogens and only mantle tissue was analysed.

C1q domain-containing proteins (C1qDCs) may function as pattern recognition receptors (PRRs) and participate in several immune responses. The sequencing and annotation of the genome of the Pacific oyster *Crassostrea gigas* revealed the presence of 321 C1qDCs, which is considerably higher than what has been reported in other protostomes and deuterostomes (Zhang et al., 2015). Zhang et al. (2015) reported that 164 C1qDCs were responsive to biotic stress in the Pacific oyster and hence highlighted their importance in the immune response. A more recent study by Lv et al. (2018) showed that 3 C1qDCs from *C. gigas* could bind lipopolysaccharide (LPS) and exhibited high binding activity towards Gram-negative bacteria. The high number of C1qDCs observed in the present study (64) are in line with the massive expansion of C1qDCs observed in other bivalves (e.g. Zhang et al., 2015). After C1qDCs, the highest number of expressed immune-related genes observed in the present study belonged to the tumor necrosis factor (TNF) family. TNFs are transmembrane proteins that among other functions, are involved in inflammation, apoptosis, cell proliferation and stimulation of the immune system (Goetz, Planas, & MacKenzie, 2004). Initially, TNFs were presumed to be present only in deuterostomes, but more recently they were also identified in arthropods (Mekata et al., 2010) and molluscs (De Zoysa, Jung, & Lee, 2009). Among the 15 TNFs identified in the present



**Fig. 1.** Immune-related genes expressed in the mantle of *R. decussatus*. Bactericidal permeability increasing protein (BPI), big defensins (BG), catalase (CAT), C-lectin domain containing proteins (CTLDC), fibrinogen domain containing proteins (FREP), galectins (GAL), globular head C1q domain containing proteins (C1q), glutathione peroxidases (GPX), inhibitor of nuclear factor kappa-B kinases (IKK), interferon regulatory factor (IRF), interleukins 17 (IL-17), interleukin 17 receptors (IL17R), interleukin-1 receptor-associated kinase (IRAK), lysozymes (LYS), macrophage migration inhibitory factors (MIF), mitogen-activated protein kinase kinase kinase (TAK1), myeloid differentiation primary response gene 88 (MyD88), nuclear factor kappa-B (NF-kB), nitric oxide synthase (NOS), peptidoglycan recognition proteins (PGRP), peroxiredoxines (PRX), stress-activated protein kinase (JNK), superoxide dismutase (SOD), TNF receptor-associated factors (TRAF), toll-like receptors (TLR), transcription factor AP-1 protein (AP-1), tumor necrosis factors (TNF); and tumor necrosis factor receptors (TNFR).

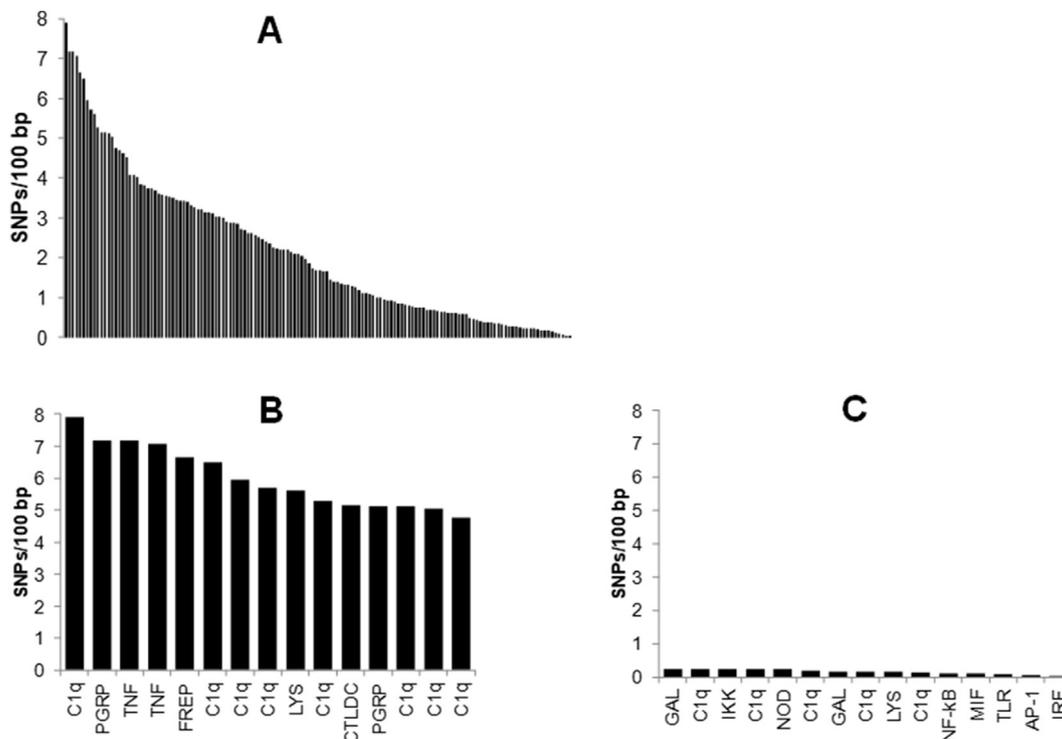
study, 9 were predicted to have a typical TNF domain, namely a transmembrane region and a tumor necrosis factor domain (see Table S1). In *C. gigas* a total of 18 TNFs were identified in the whole genome (Zhang et al., 2015). One of them (CgTNF-1) was studied in more detail and is presumed to have a crucial role in the modulation of the immune response in apoptosis, phagocytosis and regulation of anti-bacterial

activity (Sun et al., 2014). However, none of the TNFs identified in the present study were homologs of CgTNF-1, which suggest that *R. decussatus* may have more than 15 TNFs. Among the immune-related genes identified in the present study, it is also noteworthy the identification of lectins (4 C-type lectins and 5 galectins), big defensins (2) and lysozymes (3), which are presumed to be involved in the recognition and elimination of *P. olsenii* (Soudant, Chu, & Volety, 2013).

A total of 5 359 SNPs were identified in the 146 immune-related genes of *R. decussatus*. The mean density of SNPs in the 146 immune-related genes was 2.17 SNPs/100 bp. The density of SNPs varied between 0.04 and 7.92 SNPs/100 bp (Fig. 2). The level of DNA polymorphism observed (with one SNP every 46 bp) was similar to that previously reported by Sauvage, Bierne, Lapègue, and Boudry (2007) in *C. gigas* and one of the highest described in metazoa. Among the 15 immune-related genes showing the highest SNP density, 8 belonged to the C1q domain-containing protein family. Within each gene family, considerable variations in SNP density was observed. For example, the SNP density in TLRs varied between 0.07 and 3.58 SNPs/100 bp and the mean density was 3.28 SNPs/100 bp, which was higher than the mean density of all immune-related genes identified in the present study.

### 3.3. Toll-like receptors

Toll-like receptors (TLRs) are evolutionarily conserved pattern recognition receptors (PRRs) found in metazoans that have a key role in the innate immune system (Leulier & Lemaitre, 2008). TLRs recognize pathogen-associated molecular patterns (PAMPs) present in pathogens such as fungi, bacteria and viruses. Gene expansion and functional divergence of TLRs was reported recently in bivalves (Zhang et al., 2015). The sequencing of the genome of *C. gigas* revealed the existence of 83 TLRs (Zhang et al., 2012, 2015), although some of them were not predicted to have all the characteristic domains of TLRs (i.e. at least one LRR domain, one TM region and one TIR domain). The analysis of the transcriptome of other bivalve species also revealed a high number of TLRs



**Fig. 2.** SNPs per 100 bp (in both coding and non-coding regions) detected in immune-related genes expressed in the mantle of *R. decussatus*. A, distribution of SNPs/100 bp in the 146 immune-related genes. B, the 15 genes with the highest number of SNP/100 bp. C, the 15 genes with the lowest number of SNP/100 bp.

namely, 23 TLRs in the Mediterranean mussel *M. galloprovincialis* (Toubiana et al., 2013) and 32 TLRs in the Venus Clam *Cyclina sinensis* (Pan, Ren, Gao, & Gao, 2015). In the present study, we identified 10 TLRs in the transcriptome of the mantle of *R. decussatus* (Fig. 3). The number of LRRs in the 10 TLRs of *R. decussatus*, predicted using SMART, varied between 3 and 18 (Fig. 3). Four TLRs were predicted to have a signal peptide (RdTLR-2, -7 -9 and -10). The absence of a predicted signal peptide in the other 6 TLRs may indicate that the nucleotide sequences are incomplete. Nevertheless, TLRs without signal peptides have been reported in other bivalve species with complete coding sequences namely in *C. gigas* (Zhang et al., 2015). Nine TLRs were single cysteine cluster TLRs (sscTLRs) and only one (RdTLR-2) was a multiple cysteine cluster TLR (mccTLR).

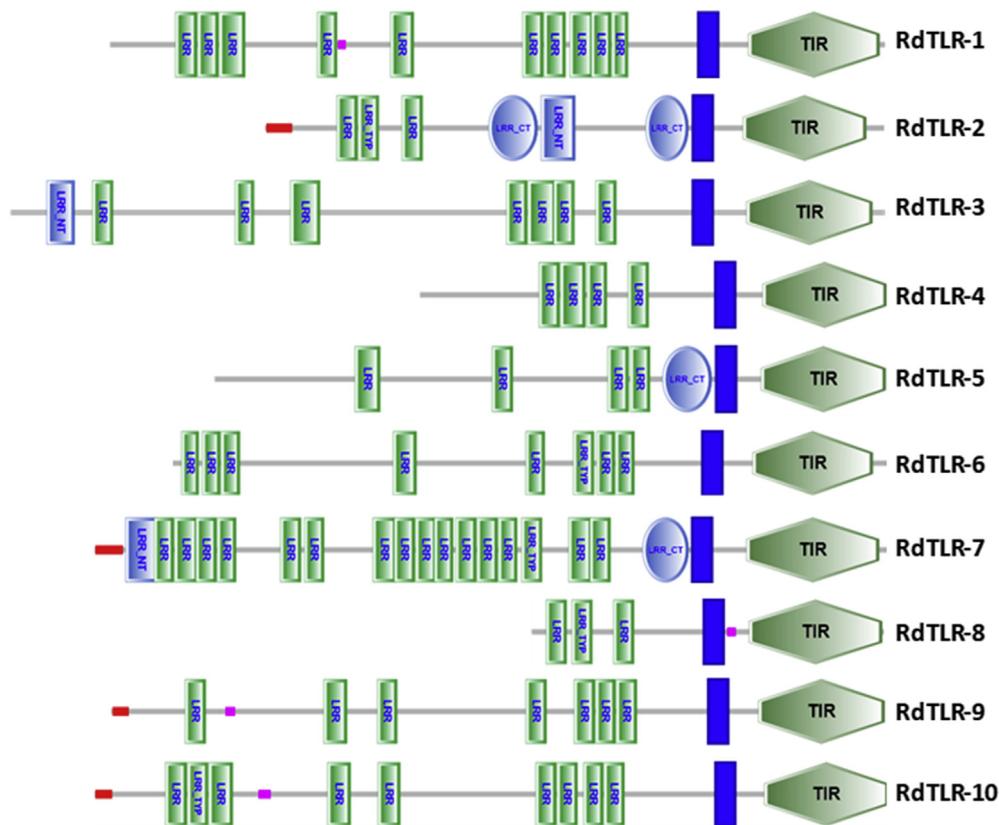
Our phylogenetic analysis suggests that multiple lineage restricted-expansions may have occurred in bivalves such as in mytilids (clusters M1, M2 and M3) and venerids (cluster V1), and in the marine gastropod *Lottia gigantea* (cluster L1) (Fig. 4). These results suggest that some TLR duplication events occurred early in mollusc evolution, but also more recently. Our phylogenetic analysis also shows that the TLRs of *R. decussatus* were diverse (Fig. 4). The highest amino acid sequence similarity was observed between RdTLR-1 and RdTLR-6 (51.4%) and the lowest between RdTLR-2 and RdTLR-8 (7.3%). This contrasts with *M. galloprovincialis* in which higher amino acid sequence similarities were observed among some TLRs (Toubiana et al., 2013). For example, a high amino acid sequence similarity of 78, 74 and 71% were observed for MgTLR-e/MgTLR-h, MgTLR-r/MgTLR-j and MgTLR-m/MgTLR-f, respectively.

Several TLRs of bivalves were shown to be differentially expressed following challenge with pathogens (bacteria, virus and fungi) or pathogen-associated molecular patterns (PAMPs) (He et al., 2015; Qiu et al., 2007; Ren et al., 2016; Toubiana et al., 2013; Zhang et al. 2011,

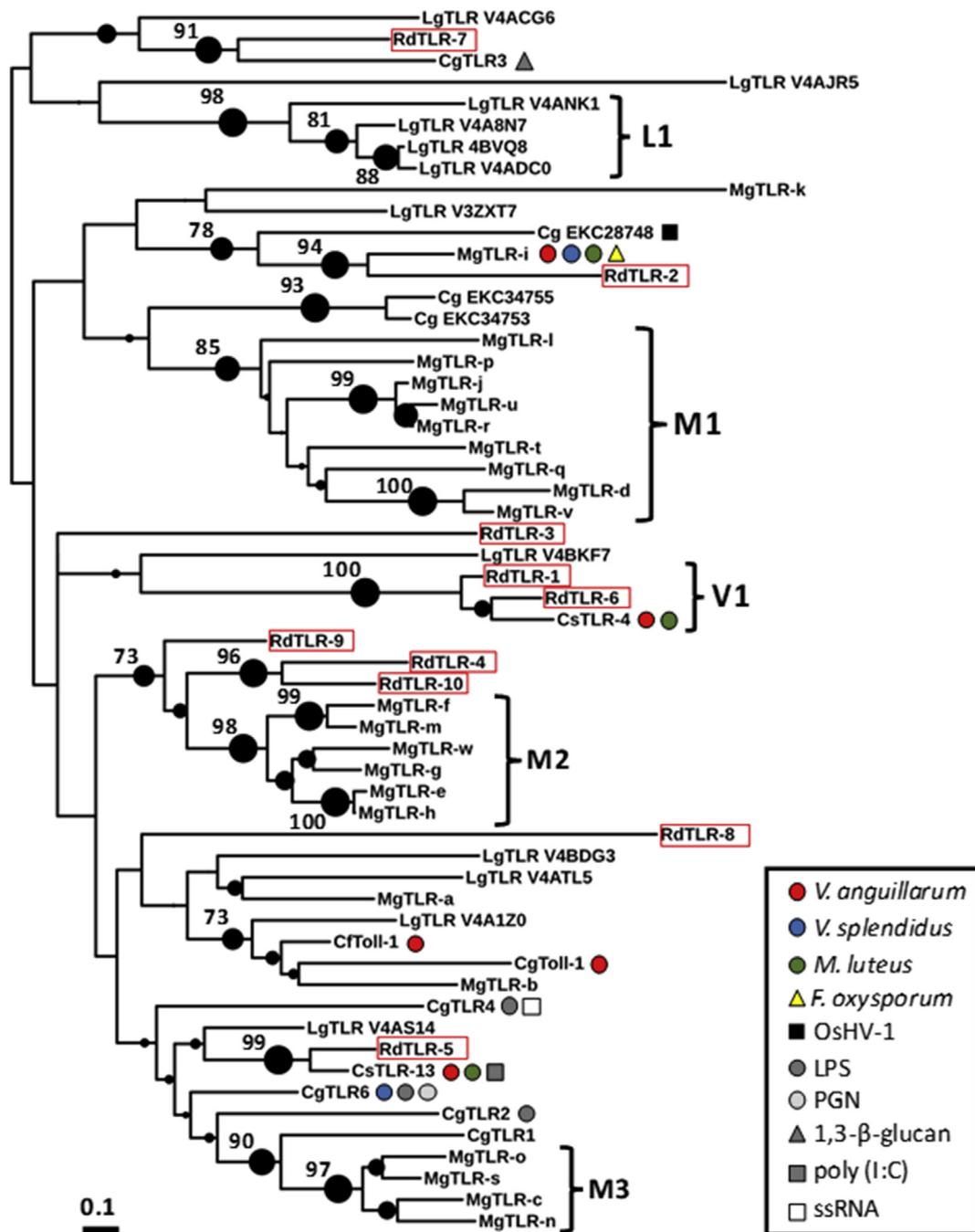
2013) (Fig. 4). Zhang et al. (2015) observed that 19 out of 78 *C. gigas* TLRs were differentially expressed following challenge with *Vibrio* spp., ostreid herpesvirus 1 or LPS. Our phylogenetic analysis revealed that 3 TLRs of *R. decussatus* were orthologues with other bivalve counterpart with known immune function. RdTLR-2 clustered robustly with MgTLR-i with a bootstrap value of 94% (Fig. 4). Toubiana et al. (2013) found that MgTLR-i was significantly up regulated in haemocytes of *M. galloprovincialis* challenged with bacteria (*V. anguillarum*, *V. splendidus* or *Micrococcus luteus*) or fungus (*Fusarium oxysporum*). It is noteworthy that both RdTLR-2 and MgTLR-i are predicted to be mccTLRs. RdTLR-6 is an orthologue of CsTLR-4, which was upregulated in haemocytes of *C. sinensis* challenged with *V. anguillarum* or *V. splendidus* (Ren et al., 2016). Finally, our results also indicate that RdTLR-5 and CsTLR-13 are orthologues. Ren et al. (2016) observed that CsTLR-13 was up-regulated in haemocytes of *C. sinensis* following challenge with *V. anguillarum*, *M. luteus* or poly (I:C). Hence, it is likely that RdTLR-2, -5 and -6 have similar immune functions to their orthologues identified in *M. galloprovincialis* and *C. sinensis*.

#### 4. Conclusions

A total 146 immune-related genes were identified in the transcriptome of the mantle of *R. decussatus*. The gene families in which the highest number of immune-related genes was observed were the: C1q domain-containing protein (64), tumor necrosis factor (15) and toll-like receptor (10) families. The density of SNPs in immune-related genes of *R. decussatus* ranged between 0.04 and 7.92 SNPs/100 bp. The highest and the lowest SNP densities were observed in genes of the C1q domain-containing protein family. Based on amino acid sequence similarity and the predicted protein domain organization, we identified 10 TLRs in *R. decussatus*. The phylogenetic analysis of the TIR domain of



**Fig. 3.** Toll-like receptors of *Ruditapes decussatus*. Red rectangle: signal peptide; Green rectangle: leucine-rich repeat (LRR) domain; Light blue rectangle: LRR N-terminal domain; Light blue ellipse: LRR C-terminal domain; Dark blue rectangle: transmembrane region; Purple rectangle: low complexity regions; Green hexagon: toll-interleukin 1 receptor (TIR) domain. (For interpretation of the references to colour in this figure legend, the reader is referred to the Web version of this article.)



**Fig. 4.** Phylogenetic tree of the TIR domain of toll-like receptors (TLRs) of *Lottia gigantea* (LgTLR), *Ruditapes decussatus* (RdTLR), *Cyclina sinensis* (CsTLR), *Chlamys farreri* (Cf), *Mytilus galloprovincialis* (MgTLR) and *Crassostrea gigas* (CgTLR) inferred through a maximum-likelihood analysis. TLRs of *R. decussatus* are indicated with a red rectangle. The size of the dark circles near nodes is proportional to the strength of bootstrap values. Only bootstrap values higher than 70% are shown near nodes. Symbols next to TLRs indicate they were differentially expressed following challenge with the gram-negative bacteria, *Vibrio anguillarum* (red circle) and *V. splendidus* (blue circle), the gram-positive bacterium, *Micrococcus luteus* (green circle), the fungus, *Fusarium oxysporum* (yellow triangle), the virus, OsHV-1 (black square), lipopolysaccharide (LPS, dark gray circle), peptidoglycan (PGN, light gray circle), 1,3-β-glucan (gray triangle), poly (I:C) (gray square) and ssRNA (white square). Differential expression data is based on the results of studies by Qiu et al. (2007), Zhang et al. (2011), Zhang et al. (2013), Toubiana et al. (2013), He et al. (2015) and Ren et al. (2016). (For interpretation of the references to colour in this figure legend, the reader is referred to the Web version of this article.)

*R. decussatus* TLRs revealed they were diverse and 3 TLRs (RdTLR-2, -5 and -6) were orthologues of TLRs from other bivalves with known immune functions. Moreover, our analysis suggests that lineage restricted-expansions of TLRs occurred in most of the mollusc taxa analysed including the venerids. The present study increases our knowledge about the immunological repertoire of *R. decussatus* and sheds light on the evolution of mollusc TLRs and by analogy with other bivalve TLRs, their

putative function.

**Acknowledgements**

This study was financially supported by the projects: AQUAGENET (SOE2/P1/E287) funded by the European Regional Development Fund within the program INTERREG IVB SUDOE; COMPETE/FEDER (FCOMP-

01-0124-FEDER-010607); PTDC/MAR/103550/2008 and CCMAR/Multi/04326/2013 funded by the Foundation for Science and Technology (FCT). F.M.B. and A.M.C were supported by postdoctoral fellowship grants from FCT (SFRH/BPD/108591/2015 and SFRH/BPD/85408/2012). We would like to thank two anonymous referees for their constructive comments.

## Appendix A. Supplementary data

Supplementary data related to this article can be found at <https://doi.org/10.1016/j.aaf.2018.03.003>.

## References

- Benzekri, H., Armesto, P., Cousin, X., Rovira, M., Crespo, D., Merlo, M. A., et al. (2014). De novo assembly, characterization and functional annotation of Senegalese sole (*Solea senegalensis*) and common sole (*Solea solea*) transcriptomes: Integration in a database and design of a microarray. *BMC Genomics*, *15*, 952.
- Bioinformatics. (2011). *FastQC: A quality control tool for high through-put sequence data*. Cambridge, United Kingdom: Babraham Institute.
- De Zoysa, M., Jung, S., & Lee, J. (2009). First molluscan TNF- $\alpha$  homologue of the TNF superfamily in disk abalone: Molecular characterization and expression analysis. *Fish & Shellfish Immunology*, *26*, 625–631.
- Food and Agriculture Organization of the United Nations (FAO). (2014). *FishStat Plus - 392 Universal software for fishery statistical time series*, 393 <http://www.fao.org/fi/statist/FISOFT/FISHPLUS.asp>. (Accessed 14 April 2015).
- Goetz, F. W., Planas, J. V., & MacKenzie, S. (2004). Tumor necrosis factors. *Developmental & Comparative Immunology*, *28*, 487–497.
- Grabherr, M. G., Haas, B. J., Yassour, M., Levin, J. Z., Thompson, D. A., Amit, I., et al. (2011). Full-length transcriptome assembly from RNA-seq data without a reference genome. *Nature Biotechnology*, *29*(7), 644–652.
- He, Y., Jouaux, A., Ford, S. E., Lelong, C., Sourdain, P., Mathieu, M., et al. (2015). Transcriptome analysis reveals strong and complex antiviral response in a mollusc. *Fish & Shellfish Immunology*, *46*(1), 131–144.
- Käll, L., Krogh, A., & Sonnhammer, E. L. (2004). A combined transmembrane topology and signal peptide prediction method. *Journal of Molecular Biology*, *338*, 1027–1036.
- Leite, R. B., Milan, M., Coppe, A., Bortoluzzi, S., Anjos, A., Reinhardt, R., et al. (2013). mRNASeq and microarray development for the Grooved carpet shell clam, *Ruditapes decussatus*: a functional approach to unravel host-parasite interaction. *BMC Genomics*, *14*, 741.
- Letunic, I., & Bork, P. (2016). Interactive tree of life (iTOL) v3: An online tool for the display and annotation of phylogenetic and other trees. *Nucleic Acids Research*, *44*(W1), W242–W245.
- Letunic, I., Doerks, T., & Bork, P. (2014). SMART: Recent updates, new developments, and status in 2015. *Nucleic Acids Research*, *43*, 257–260.
- Leulier, F., & Lemaitre, B. (2008). Toll-like receptors—taking an evolutionary approach. *Nature Reviews Genetics*, *9*, 165–178.
- Li, H. (2013). *Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM*. arXiv:1303.3997.
- Li, W., Cowley, A., Uludag, M., Gur, T., McWilliam, H., Squizzato, S., et al. (2015). The EMBL-EBI bioinformatics web and programmatic tools framework. *Nucleic Acids Research*, *43*(W1), W580–W584.
- Lv, Z., Qiu, L., Wang, M., Jia, Z., Wang, W., Xin, L., et al. (2018). Comparative study of three C1q domain containing proteins from pacific oyster *Crassostrea gigas*. *Developmental & Comparative Immunology*, *78*, 42–51.
- Mekata, T., Sudhakaran, R., Okugawa, S., Inada, M., Kono, T., Sakai, M., et al. (2010). A novel gene of tumor necrosis factor ligand superfamily from kuruma shrimp (*Marsupenaeus japonicus*). *Fish & Shellfish Immunology*, *28*, 571–578.
- Moreira, R., Pereira, P., Canchaya, C., Posada, D., Figueras, A., & Novoa, B. (2015). RNA-seq in *Mytilus galloprovincialis*: Comparative transcriptomics and expression profiles among different tissues. *BMC Genomics*, *16*, 728.
- Musacchia, F., Basu, S., Petrosino, G., Salvemini, M., & Sanges, R. (2015). Annotcript: A flexible pipeline for the annotation of transcriptomes also able to identify putative long noncoding RNAs. *Bioinformatics*, *31*(13), 2199–2201.
- Pan, B., Ren, Y., Gao, J., & Gao, H. (2015). De novo RNA-Seq analysis of the Venus clam, *Cyclina sinensis*, and the identification of immune-related genes. *PLoS One*, *10*(4), e0123296.
- Petersen, T. N., Brunak, S., von Heijne, G., & Nielsen, H. (2011). SignalP 4.0: Discriminating signal peptides from transmembrane regions. *Nature Methods*, *8*, 785–786.
- Qiu, L., Song, L., Xu, W., Ni, D., & Yu, Y. (2007). Molecular cloning and expression of a Toll receptor gene homologue from Zhikong Scallop, *Chlamys farreri*. *Fish & Shellfish Immunology*, *22*(5), 451–466.
- Ren, Y., Pan, H., Pan, B., & Bu, W. (2016). Identification and functional characterization of three TLR signaling pathway genes in *Cyclina sinensis*. *Fish & Shellfish Immunology*, *50*, 150–159.
- Ruano, F., Batista, F. M., & Arcangeli, G. (2015). Perkinsiosis in the clams *Ruditapes decussatus* and *R. philippinarum* in Northeast Atlantic and Mediterranean Sea: A review. *Journal of Invertebrate Pathology*, *131*, 58–67.
- Sauvage, C., Bierné, N., Lapègue, S., & Boudry, P. (2007). Single Nucleotide polymorphisms and their relationship to codon usage bias in the Pacific oyster *Crassostrea gigas*. *Gene*, *406*, 13–22.
- Schmitt, P., Gueguen, Y., Desmarais, E., Bachère, E., & de Lorgeril, J. (2010). Molecular diversity of antimicrobial effectors in the oyster *Crassostrea gigas*. *BMC Evolutionary Biology*, *10*, 23.
- Simakov, O., Marletaz, F., Cho, S. J., Edsinger-Gonzales, E., Havlak, P., Hellsten, U., et al. (2012). Insights into bilaterian evolution from three spiralian genomes. *Nature*, *493*, 526–531.
- Soudant, P., Chu, F.-L. E., & Volety, A. (2013). Host-parasite interactions: Marine bivalve molluscs and protozoan parasites, *Perkinsus* species. *Journal of Invertebrate Pathology*, *114*, 196–216.
- Sun, Y., Zhou, Z., Wang, L., Yang, C., Jianga, S., & Song, L. (2014). The immunomodulation of a novel tumor necrosis factor (CgTNF-1) in oyster *Crassostrea gigas*. *Developmental & Comparative Immunology*, *45*, 291–299.
- Tamura, K., Stecher, G., Peterson, D., Filipiński, A., & Kumar, S. (2013). MEGA6: Molecular evolutionary genetics analysis version 6.0. *Molecular Biology and Evolution*, *30*, 2725–2729.
- Toubiana, M., Gerdol, M., Rosani, U., Pallavicini, A., Venier, P., & Roch, P. (2013). Toll-like receptors and MyD88 adaptors in *Mytilus*: Complete cds and gene expression levels. *Developmental & Comparative Immunology*, *40*, 158e166.
- Zhang, G., Fang, X., Guo, X., Li, L., Luo, R., Xu, F., et al. (2012). The oyster genome reveals stress adaptation and complexity of shell formation. *Nature*, *490*, 49e54.
- Zhang, L., Guo, X., Litman, G. W., Dishaw, L. J., & Zhang, G. (2015). Massive expansion and functional divergence of innate genes in a protostome. *Nature Scientific Reports*, *5*, 8693.
- Zhang, Y., He, X., Yu, F., Xiang, Z., Li, J., Thorpe, K. L., et al. (2013). Characteristic and functional analysis of toll-like receptors (TLRs) in the lophotrochozoan, *Crassostrea gigas*, reveals ancient origin of TLR-mediated innate immunity. *PLoS One*, *8*(10), e76464.
- Zhang, L., Li, L., & Zhang, G. (2011). A *Crassostrea gigas* Toll-like receptor and comparative analysis of TLR pathway in invertebrates. *Fish & Shellfish Immunology*, *30*, 653–660.