

QATAR UNIVERSITY

COLLEGE OF ENGINEERING

DIFFERENTIAL ARCHITECTURE SEARCH IN DEEP LEARNING FOR DNA
SPLICE SITE CLASSIFICATION

BY

SHABIR MOOSA

A Thesis Submitted to
the Faculty of the College of Engineering
in Partial Fulfillment of the Requirements for the Degree of
Masters of Science in Computing

June 2019

© 2019 Shabir Moosa. All Rights Reserved.

COMMITTEE PAGE

The members of the committee approve the Thesis of
Shabir Moosa defended on 17/04/2019.

Prof. Abbes Amira
Thesis Supervisor

Dr. Sabri Boughorbel
Thesis Co-Supervisor

Dr. Puthen Veettil Jithesh
Committee Member

Dr. Abdelkarim Erradi
Committee Member

Approved:

Abdel Magid Hamouda, Dean, Engineering

ABSTRACT

Moosa, Shabir, Masters:

June: 2019, Masters of Science in Computing

Title: Differential Architecture Search in Deep Learning for DNA Splice Site Classification

Supervisor of Thesis: Prof. Abbas Amira

The data explosion caused by unprecedented advancements in the field of genomics is constantly challenging the conventional methods used in the interpretation of the human genome. The demand for robust algorithms over the recent years has brought huge success in the field of Deep Learning (DL) in solving many difficult tasks in image, speech and natural language processing by automating the manual process of architecture design. This has been fueled through the development of new DL architectures. Yet genomics possesses unique challenges as we expect DL to provide a super human intelligence that easily interprets a human genome. In this thesis, the state-of-the art DL approach based on Differential Architecture Search (DARTS) mechanism was adapted for interpretation of biological sequences. This method has been applied to the splice site recognition task on raw Deoxyribonucleic Acid (DNA) sequences to discover high-performance convolutional architectures by automated engineering. The discovered architecture outperformed fixed DL architectures. As part of the study, the DARTS model was benchmarked on CPU and multiple GPU architectures in terms of computational time and classification performance. The results have shown a potential of using this automated architecture search mechanism for solving other problems in genomics.

DEDICATION

“For my family and friends, for their constant support, love and faith in me”

ACKNOWLEDGEMENTS

First and foremost, I thank Allah, the almighty, for giving me the strength, knowledge, ability and opportunity to undertake this research study and be able to complete it satisfactorily. Without his blessings, this achievement would not have been possible.

I would also like to express my heartfelt gratitude to my parents for their unconditional love, encouragement and prayers. I thank with love to my wife and daughter who are the biggest source of strength and happiness for me. My acknowledgement would be incomplete without thanking my friends for keeping me going on my path to success and for ensuring that good times keep flowing.

I would like to submit my sincere and deepest gratitude to my supervisors, Dr. Abbas Amira and Dr. Sabri Boughorbel for their meticulous supervision, admirable patience, and constant support. I am deeply indebted to their highly commendable efforts during my journey in research. I am also grateful to the team at Sidra Medicine, for their persistent support given to me at all times.

I would also like to thank all my professors and friends at Qatar University for their support, encouragement and credible ideas during the entire course of my Masters. Special thanks go to Zeineb, Hisham, George, Hakem and Saad.

TABLE OF CONTENTS

Dedication	iv
Acknowledgements	v
List of Tables	ix
List of Figures	x
List of Abbreviations	xiv
INTRODUCTION	1
Human Genome	1
Genomics Research in Qatar	2
Motivation	2
Problem Statement	3
Thesis Objective	3
Research Contributions	4
Thesis Outline	5
BACKGROUND AND LITERATURE REVIEW	6
Overview of Deep Learning	6
Convolutional Neural Networks	8
Recurrent Neural Networks	9
Autoencoders	10

Architecture Designing in Deep Learning	10
Advancement in Deep Learning	13
Splice Site Recognition Problem	15
Splice Site Detection Methods	16
Comparative Analysis	18
Summary	20
METHODOLOGY	22
DNA Representation	22
DARTS Algorithm	23
Performance Evaluation Method	26
Classification Performance Metrics	27
Computational Performance Metrics	28
IMPLEMENTATION	30
Dataset	30
Experimental Setup	31
Architecture Search	31
Architecture Evaluation	34
RESULTS	36
Classification Performance Results	36
Computational Performance Results	37
Execution Time	37

Precision	37
CONCLUSION AND FUTURE WORK	42
REFERENCES	44
Appendix: Detailed Results	53

LIST OF TABLES

Comparative Analysis of Literature Review	19
Hardware for Benchmarking	31
Hyperparameters for architecture search	34
Comparison of Model Performance	36
Comparison of Execution Time	37
Comparison of Learning and Inference Speed on GPUs	41
Accuracy and Loss for Seed 1	69
Accuracy and Loss for Seed 2	70
Accuracy and Loss for Seed 3	71
Accuracy and Loss for Seed 4	72
Accuracy and Loss for Seed 5	73
Accuracy and Loss for Seed 6	74
Accuracy and Loss for Seed 7	75
Accuracy and Loss for Seed 8	76
Accuracy and Loss for Seed 9	77
Accuracy and Loss for Seed 10	78

LIST OF FIGURES

An illustration of DNA.	1
DL architecture: Multilayer perceptron.	7
Types of neural networks.	8
CNN Architecture.	11
CNN Architectures.	14
Splice-junction sites.	16
DNA representation by one-hot vector.	23
DARTS method.	23
Confusion matrix.	27
Normal and Reduction cell in Epoch 1.	32
Normal and Reduction cell in Epoch 2.	33
Normal and Reduction cell in Epoch 3.	33
Normal and Reduction cell in Epoch 4.	34
Normal and Reduction cell learned on splice dataset.	35
Plots of training accuracy and loss.	38
Plots of validation accuracy and loss.	39
Plot of execution time for DARTS.	40
Plot of comparison of training and inference speed.	40
Normal and Reduction cell in Epoch 6.	53
Normal and Reduction cell in Epoch 7.	53
Normal and Reduction cell in Epoch 8.	54
Normal and Reduction cell in Epoch 9.	54
Normal and Reduction cell in Epoch 10.	54
Normal and Reduction cell in Epoch 11.	55
Normal and Reduction cell in Epoch 12.	55

Normal and Reduction cell in Epoch 13.	55
Normal and Reduction cell in Epoch 14.	56
Normal and Reduction cell in Epoch 15.	56
Normal and Reduction cell in Epoch 16.	56
Normal and Reduction cell in Epoch 17.	57
Normal and Reduction cell in Epoch 18.	57
Normal and Reduction cell in Epoch 19.	57
Normal and Reduction cell in Epoch 20.	58
Normal and Reduction cell in Epoch 21.	58
Normal and Reduction cell in Epoch 22.	59
Normal and Reduction cell in Epoch 23.	59
Normal and Reduction cell in Epoch 24.	59
Normal and Reduction cell in Epoch 25.	60
Normal and Reduction cell in Epoch 26.	60
Normal and Reduction cell in Epoch 27.	60
Normal and Reduction cell in Epoch 28.	61
Normal and Reduction cell in Epoch 29.	61
Normal and Reduction cell in Epoch 30.	61
Normal and Reduction cell in Epoch 31.	62
Normal and Reduction cell in Epoch 32.	62
Normal and Reduction cell in Epoch 33.	62
Normal and Reduction cell in Epoch 34.	63
Normal and Reduction cell in Epoch 35.	63
Normal and Reduction cell in Epoch 36.	63
Normal and Reduction cell in Epoch 37.	64
Normal and Reduction cell in Epoch 38.	64
Normal and Reduction cell in Epoch 39.	64
Normal and Reduction cell in Epoch 40.	65

Normal and Reduction cell in Epoch 41.	65
Normal and Reduction cell in Epoch 42.	65
Normal and Reduction cell in Epoch 43.	66
Normal and Reduction cell in Epoch 44.	66
Normal and Reduction cell in Epoch 45.	66
Normal and Reduction cell in Epoch 46.	67
Normal and Reduction cell in Epoch 47.	67
Normal and Reduction cell in Epoch 48.	67
Normal and Reduction cell in Epoch 49.	68
Normal and Reduction cell in Epoch 50.	68

LIST OF ABBREVIATIONS

- CNN** Convolutional Neural Networks
- DA** Deep Autoencoder
- DARTS** Differential Architecture Search
- DBM** Deep Boltzmann Machine
- DBN** Deep Belief Network
- DL** Deep Learning
- DNA** Deoxyribonucleic Acid
- DNN** Deep Neural Network
- GPU** Graphical Processing Unit
- GRU** Gated Recurrent Units
- LSTM** Long Short Term Memory
- ML** Machine Learning
- NAO** Neural Architecture Optimization
- NAS** Neural Architecture Search
- NGS** Next Generation Sequencing
- PPM** Path Towards Personalized Medicine
- QGP** Qatar Genome Program
- QNRF** Qatar National Research Fund
- RNN** Recurrent Neural Network

SGD Stochastic Gradient Descent

TPU Tensor Processing Unit

CHAPTER 1: INTRODUCTION

Human Genome

The Human Genome consists of a set of nucleic acid sequences encoded as DNA molecule. The DNA forms a double helix shape and is composed of nucleotides. Each nucleotide is composed of a nucleoside and a phosphate group. A nucleoside is a nitrogenous base and a sugar. The nitrogenous bases are of 4 types; Adenine(A), Thymine(T), Guanine(G) and Cytosine(C). The information in a DNA is stored in these four chemical bases. As shown in Figure 1.1, the bases pair up with each other in such a way that A pairs with T and G pairs with C. Almost 99 percent of the bases are same in all humans and their order provides information for building and maintaining the human body. Each DNA strand forms the shape of a spiral ladder with the base pairs attached to the sugar-phosphate backbone.

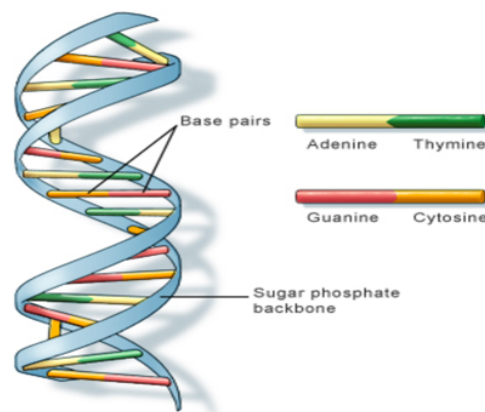


Figure 1.1: An illustration of DNA.

The prosperity of research in genomics started ever since the DNA molecules were interpreted by Watson and Crick [1] as the medium of genetic information. Researchers have been ever since striving to decipher the biological process using this genetic information in order to study the roles assumed by multiple genetics factors under different environmental conditions. Genomic research has become data

intensive with the advancement of Next Generation Sequencing (NGS) techniques with the capability of sequencing an entire human genome in a single day. The vast amount of information generated can be used for conducting scientific study using statistical methods. These methods can be used to recognize patterns in the DNA sequences by identifying various genomic elements like introns, exons, positioned nucleosomes, promoters, enhancers and splice site junctions.

Genomics Research in Qatar

Recently, there has been an increased interest in genomics research in Qatar which could lead to great advancement in the field of precision medicine and personalized healthcare. This section highlights the areas of genomic research initiatives in Qatar. The Qatar Genome Program (QGP), launched in December 2013, is a national initiative that aims to map the genome of Qatari population. The aim of the project is to develop personalized healthcare by incorporating advanced and innovative genomic technologies into research and medicine. Individual research studies have further contributed to the availability of whole genomes and exomes providing insights to the genetic structure of the Qatari population [2]. The Qatar National Research Fund (QNRF) along with the collaboration of QGP launched the initiative of Path Towards Personalized Medicine (PPM) in 2015 to support and advance research activities focused on providing personalized medical treatment tailored to patient's individual characteristics.

Motivation

The genomics community is rapidly embracing DL techniques to solve many of the problems in the field. But DL is yet to revolutionize the genomics domain to resolve many of the pressing challenges [3]. The intersection of genomic research and current state-of-the-art DL methods in computer vision, speech recognition and natural

language processing will lead to an insightful understanding of genomics that will benefit multiple fields like precision medicine and drug discovery. Hence, there is a need to adopt the powerful DL methods utilized in other domains to tackle the challenges in the field of genomics. The research work is carried out at SIDRA and the methods will be evaluated on SIDRA's High Performance Computing platform.

Problem Statement

Deep Learning is shifting the paradigm in Machine Learning (ML) from feature engineering and algorithmic development into a single framework where no hand crafted feature are needed. However, the recent trends in DL have shown that much effort is dedicated to the design of complex neural network architectures. For example in ImageNet competition, through 4 years the winning model went from 8 layers [4] in 2012 to 152 layers [5] in 2016 with various choices on architectures parameters such as skip connections, filter sizes, repeated blocks etc. The problem lifted by DL of feature engineering is creating an additional burden of architecture engineering. Therefore there is a need to automate the process of manual architecture definition to a data-driven approach. Our objective is to explore some recent DL techniques to tackle this issue in genomics domain. Many of the architectures used in genomics have been borrowed from computer vision and natural language processing domains and still undergo the manual tasks of feature and architecture engineering. To the best of our knowledge this is the first attempt to apply and adapt similar techniques for genomics data.

Thesis Objective

The focus of this thesis is to adapt the DARTS technique in [6] for splice site classification in genomics domain. The study was performed on the Splice Site Recognition (SSR) dataset which provides a scope for analysis of the human genome and iden-

tification of unknown regions to understand the biochemical processes involved in building and maintaining a human body. The following are the objectives of the thesis:

- Conduct a literature review on existing DL approaches performed in solving problems in the field of genomics.
- Develop new DL convolutional architectures using DARTS approach for splice site classification.
- Evaluate the performance of the discovered architecture against fixed architectures based on Recurrent Neural Network (RNN) and Convolutional Neural Networks (CNN).
- Benchmark the architecture performance over CPU and multiple GPU models in terms of execution time, learning and inference speed.

Research Contributions

The main contributions of this thesis are shown in the following.

- A literature review is conducted on the application of DL techniques in genomics. For each of the selected studies, the addressed problem, DL architecture used, dataset, and the achieved performance are presented. The methods used to address challenges were discussed, and a comparison between the reviewed studies is presented.
- A recently proposed continuous architecture search method, DARTS, was adapted and applied on DNA data. This was initially implemented on image dataset. This is the first research study that has applied Neural Architecture Search (NAS) to solve a problem in genomics domain.

- A data loader for DNA sequence was written as the model input initially was images. This was adapted to take sequence data represented by a four-dimensional hot-vectors representing the four base pairs (A,C, T,G).
- For model evaluation, the original code written in pytorch worked only for Graphical Processing Unit (GPU). All the necessary adaptation to handle CPU processing were introduced.
- For computational performance analysis, the precision of model weights was changed to experiment with different precision levels of single, half and double floating point operations. The learning and evaluation experiments adapted to benchmark on the three data types in the each of the forward and backward passes.
- Implemented visualization graphs for the experiments and interactive standard and mean deviation plots of the results.

Thesis Outline

The thesis is organized as follows. In Chapter 2, we present the necessary background information of the existing approaches for the splice site classification problem. In Chapter 3, the DARTS algorithm is discussed in detail. Our adapted approach and the materials and methods used are discussed in Chapter 4. Chapter 5 presents an evaluation of the results obtained. Finally, Chapter 6 concludes the thesis and proposes some future work.

CHAPTER 2: BACKGROUND AND LITERATURE REVIEW

We present the background needed for the remainder of the thesis in this chapter. In Section 2.1, we present an overview of DL models. Section 2.3 presents the advancements in the field of DL. Section 2.4 introduces the splice site recognition problem. Section 2.5 presents the literature review of the existing methods used for prediction and classification of splice sites. The literature was specifically searched for studies that applied ML and DL techniques and taking into review the various architectures used. In Section 2.6, we present a table with a comparative analysis of the studies

Overview of Deep Learning

Deep Learning is a class of ML algorithms that combines raw inputs into layers of intermediate features. They take raw features from large datasets and use them to create a predictive tool from the patterns hidden inside the data. They have shown impressive results over existing best-in class ML algorithms across various domains. For the past five years, DL algorithms have revolutionized fields such as high-energy physics [7], computational chemistry [8] and dermatology [9]. The off-the-shelf implementation of these algorithms across different fields have produced comparable or higher accuracies than previous state-of-the art methods that required extensive customization over year.

Deep Learning techniques rely on artificial neural networks which grew from research on artificial neurons that are similar to the neurons in the brain. The history of artificial neural networks dates back to first computational model proposed in [10] where layers of neuron-like nodes mimic how neurons in the brain process information. The neural networks are composed of an input layer, one or more hidden layers and eventually a link to the output layer as shown in Figure 2.1. A layer has a set of nodes which are connected using edges to the immediate earlier and deeper lay-

ers. The nodes in certain neural networks connect to itself with a delay. The input layers consist of the variables measured in the data set of interest. There are multiple hidden layers in neural networks where each layer does feature construction for the layers before it. The increasing levels of abstraction allows efficient learning of data representation. The higher level abstractions are learned by representing them at lower levels as less abstraction. These are very powerful techniques as they help to learn from the raw data directly [11] and can be used in many fields. DL has been successful in areas such as computer vision, image recognition, speech translation, object detection and robotics and is most useful when large datasets are available.

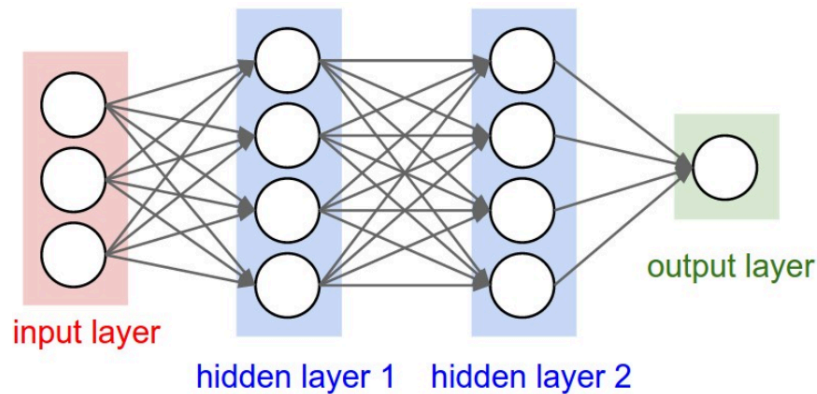


Figure 2.1: DL architecture: Multilayer perceptron.

The intersection of DL and genomic research has led to insightful understanding of the genomics domain. Although DL is still in early stages in genomics, it has influenced many fields such as cancer diagnosis and treatment, population genetics and functional genomics. It has been particularly useful in gene expression and disease analysis [12] [13] for predicting the effects of mutations in non-coding regions in a DNA. It has made effective contributions in the detection of skin cancer [9] and breast cancer [14]. Several DL architectures as shown in Fig. 2.2 have been used in the field of genomics such as RNN, CNN, Deep Neural Network (DNN), Deep Belief Network (DBN), Deep Boltzmann Machine (DBM) and Deep Autoencoder (DA).

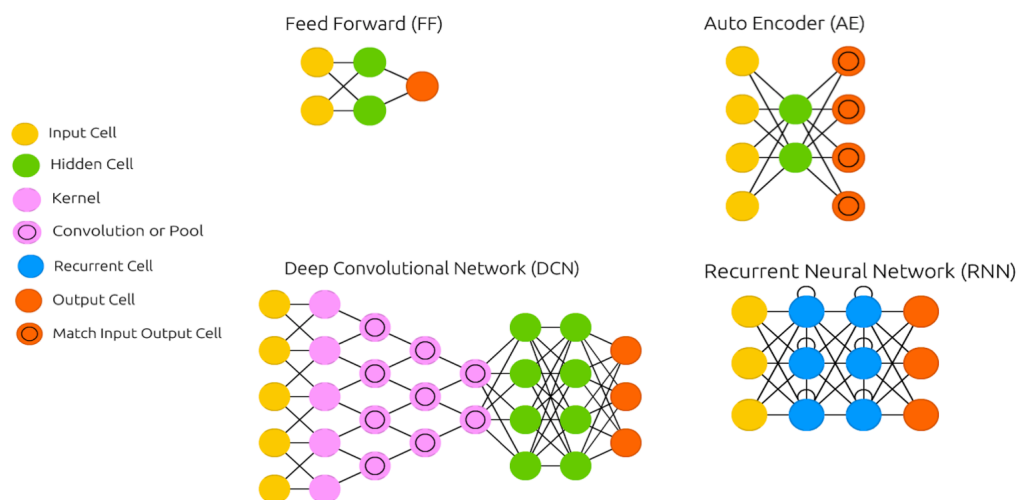


Figure 2.2: Types of neural networks.

These different types of architectures have been used to solve various specific problems in genomics. For example, the CNN architectures have been used for problems in image classification tasks and have been widely adopted in genomics for automatically learning the characteristics of gene data. The RNNs used in speech recognition problems have found to be efficient in handling DNA sequence data. The following section provides a review of how various DL architectures have been used in the perspective of genomics study.

Convolutional Neural Networks

CNN have been most successful in the field of image processing and computer vision due to their ability to examine spatial information. In genomics, the fundamental building blocks of CNN used in computer vision [4] was utilized for feature extraction by representation of the DNA sequence as an image. CNN architectures have been used for discovering sequence patterns of genomic sequence motifs to perform classification tasks [15]. Several three layer CNN models such as DeepSEA [16] have been developed for sequence based prediction and classification problems and a similar architecture in [17] to analyze the functional activities in a genome sequence.

CNN models [18] and DeepBind [19] were proposed to understand sequence specificity involved in binding of proteins.

Though CNN architectures have shown superior performance over existing methods, poor architecture design would result in inefficient performance of these tuned models [18]. Therefore, it is of utmost importance for the researcher to have biological background and an in-depth understanding of the CNN architecture to optimize and choose a particular architecture for a specific task. Increasing the depth of the network will not have much effect, but choice and design of convolutional and pooling operations, window and kernel size could result in much improved performance.

Recurrent Neural Networks

RNN have shown impressive results in the field of natural language processing, speech recognition and translation. They have proven to outperform CNN and other DL models on sequential data due to their memory like property to store information about past inputs. The inputs are scanned in a sequential manner and the output is a function of the the current input and previous hidden layer. A variance of the RNN is the bi-directional model proposed in [20] where the future inputs are considered as well. Subsequently to solve the problem of vanishing gradient Long Short Term Memory (LSTM) was proposed in [21] and later Gated Recurrent Units (GRU) in [22] that truncated the gradient propagation.

The sequential nature of the DNA made RNN models applicable to use in many scenarios. A neural machine translation model based on LSTM was proposed in that converted the protein prediction problem as similar to a language translation task. Many other models based on RNN such as DeepNano [23], a hybrid convolutional LSTM model [24] named DanQ were built for prediction from protein sequences. Recently proposed seq-to-seq model based on RNN [25] has been applied for prediction of secondary protein structure.

Autoencoders

Autoencoders were initially used for the initialization of network weights and have now been used for deeper architectures by stacking a number of autoencoders to be extended to form denoising autoencoder [26], contractive autoencoder [27], sparse autoencoder [28] and variational autoencoder [29]. They have been successful due to their ability to learn from compressed input representation through encode-decode method. Autoencoders have been used for dimension reduction in gene expression and also for gene clustering tasks. The autoencoders do not guarantee improvement in the model with higher reconstruction accuracy. Variational autoencoders are best suited for genomic tasks because of their ability to deal with complex dependencies that exists. A two step model was proposed in [30] for prediction in drug responses with an initial prediction by unsupervised model and then by a semi-supervised prediction model. The unsupervised model called the Perturbation variational autoencoder embedded the data space to a lower dimensional latent space to model the drug-induced effect into a linear function. The semi-supervised prediction model Drug Response variational autoencoder is then trained jointly to model both drug-induced effects as well as the outcome of the treatment. These variational autoencoder frameworks have led to improvement in predicting drug responses.

Architecture Designing in Deep Learning

Designing a DL architecture is one of the key hyperparameters to solve problems using DL techniques. At present, DL architectures are designed manually by human experts by handcrafting through careful experimentation or modifying existing networks. CNN architectures are a class of neural architectures that consist of alternating layers of convolution and pooling operations. The final layers consist of fully connected layers and the last layer is a softmax classifier as shown in Figure 2.3. These architectures are trained using back-propagation via Stochastic Gradient De-

scend (SGD) to determine the network weights and biases to minimize the loss function for mapping the input to outputs. The CNN has a set of learnable kernels or

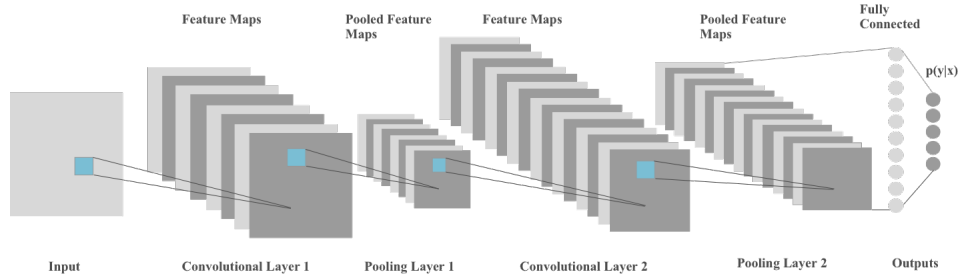


Figure 2.3: CNN Architecture.

features that extracts features from input data to produce feature maps. Feature maps are generated by sliding the filter over input and performing the dot product or the convolution operation, followed by a non-linear activation function such as sigmoid, ReLU, and tanh. The activation function is to introduce non-linearity in the model. ReLU is the preferred activation function as it speeds up training better than others [4]. The weights are shared among feature maps to reduce the number of parameters and also to detect the same feature irrespective of its location in the inputs [31]. The feature map size depends on the size of the filter and the stride, so when convolution on an input image of size (IXI) by a filter of size (FXF) with stride S , the output size will be (OXO) given by:

$$O = \frac{I - F}{S} + 1 \quad (2.1)$$

The pooling operation reduces the resolution of the feature maps from the previous convolution operation. The input is split into disjoint regions and an output of size (RXR) from each region is produced. There are two types of pooling operations: max pooling or average pooling [32]. When a feature map of size (OXO) is fed into pooling layer, the output is given by:

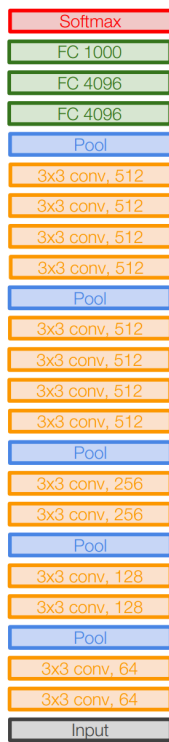
$$P = \frac{I}{R} \quad (2.2)$$

The final layer of CNN consists of a fully connected layer which extracts the global features from the inputs. This layer is similar to a feed-forward neural network where the units are connected to all the hidden units in the previous layer. The last layer is a softmax layer which calculates the probability over K given classes. The availability of more number of classes to predict from implies that more features need to be learned. This requires more filters increasing the total number of parameters in the network. There has been improvements in CNN architectures from the start of 1989 to date in optimization of parameters, regularization, structural changes in designing of new blocks, etc. Many CNN architectures have been introduced over the years that has increased the complexity of architecture and these designs have been based on intuition and engineering. The presence of a number of possible variations and choices makes the design space of CNN architectures extremely hard for a manual exhaustive search. A number of architectures were introduced over the years such as VGGNet [4] as shown in Fig. 2.4a which had used filters with a small receptive field of 3×3 and a convolution stride of 1. The spatial pooling had 5 layers of max pooling operations. The image is padded to preserve the spatial resolution after convolution. The hidden layers were equipped with ReLU operations. This architecture mainly addressed the depth aspect of the convnet architectures design. Another major CNN architecture introduced by google around 2014 in the ImageNet Large-Scale Visual Recognition Challenge was GoogleNet [33] shown in Fig. 2.4c. The simple convolution layer was replaced with an inception module which performed 3×3 , 5×5 convolution operations with 3×3 max pooling and 1×1 convolution operation operating independently on the feature maps from the previous layers. All the feature maps were then concatenated in the end. The ResNet model [5] shown in Fig. 2.4b was proposed in 2015 which introduced a novel architecture with skip connections and batch normalization. The skip connections were gated recurrent units similar to

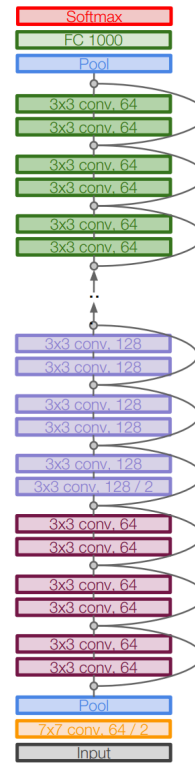
the one in RNN. The architecture had 152 layers and was able to outperform human level performance.

Advancement in Deep Learning

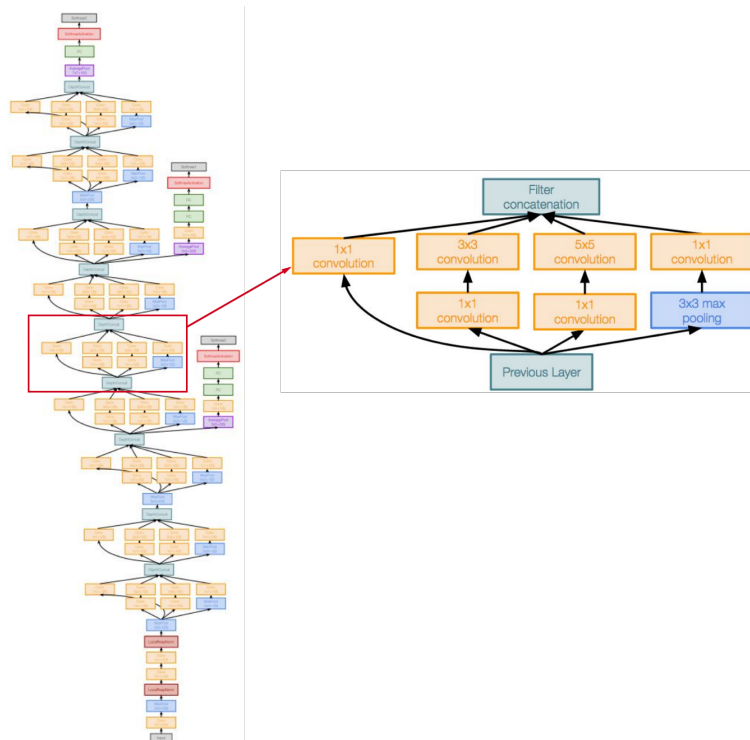
The advancement of neural networks have accomplished major achievements in the field of image classification, object detection and natural language processing. Designing these neural network architectures requires computational resources and significant efforts from human experts in DL through trial and error. Over the recent years, there has been a paradigm shift from feature designing to architecture designing in the field of image classification and natural language processing [34, 35, 36, 37] to develop algorithmic solutions for automating the manual process of architecture design using NAS methods. They have provided promising results in designing models better than human designed ones on some benchmark datasets. Architecture search aims to automate every aspect of ML so that non-experts can apply ML to solve their problems at hand. The goal is to find an optimal architecture from a given search space so that the validation accuracy on a particular task is maximized. NAS has some similarities to program synthesis and inductive programming where a program is searched from examples [38]. Many architecture search algorithms have been proposed such as Reinforcement Learning (RL) [37] which uses a policy gradient algorithm to optimize architecture configurations. This approach is computationally expensive and time consuming as they design and train each network from scratch during the exploration in the search space. Several approaches were proposed for improving the efficiency of NAS such as establishing a particular structure for the search space [34], performance prediction [39] and weight prediction of individual architectures [40] and by a parameter sharing mechanism [41] across multiple architectures. A novel approach of searching the architectures over a continuous domain alternate to searching over a discrete set of child architectures were proposed



(a) VGGNet Architecture.



(b) ResNet Architecture.



(c) GoogleNet Architecture.

Figure 2.4: CNN Architectures.

by DARTS [6] and Neural Architecture Optimization (NAO) [42]. These continuous search mechanisms has outperformed other architecture search algorithms by achieving competitive performance over a rich architecture search space and using less computational resources. Both DARTS and NAO performs continuous optimization using gradient based method to find the best architecture. The difference in both approaches lies in the optimization space and method of deriving the best architecture from the continuous space. The NAO uses a decoder network to derive the architecture whereas in the DARTS approach the discrete architecture is obtained by *argmax* of the mixture of operations in the network.

Splice Site Recognition Problem

Proteins form an essential component in all living organisms and a major biological process that occurs in all living cells is the production of proteins. They play a vital role in the biochemical reactions within cells and in metabolism. The protein coding process called gene expression occurs in two stages: Transcription and Translation. During Transcription, the DNA is copied to produce an mRNA (messenger Ribonucleic Acid). The protein coding process occurs in the translation phase where the mRNA sequence is decoded to produce the proteins.

Eukaryotic organisms are complex organisms with cells in which the genetic material is made up of a membrane-bound nucleus or nuclei. Their genes are composed of alternated segments of introns and exons. Exons form the coding regions in a DNA during translation to proteins. The biological significance of intronic regions are not known yet as they do not participate in the protein building process. During translation stage in eukaryotes, the process of splicing occurs where the introns are spliced out from the mRNA molecule. Splice site classification is a sub-problem of gene prediction to correctly recognize genes from a DNA sequence. This prediction is hard due to the mechanism of splicing to remove the introns and combine exons.

The exons are short in length and has few nucleotides, whereas introns are longer. The number of introns and their length vary among species and even within the same species. The introns vary differently and the ones involved in splicing are called spliceosomal introns. The spliceosomal introns follow the GT-AG rule where the 5' end begins with nucleotides GT and end with AG at the 3' end. These borders are referred to as splice sites. The boundary points where splicing occurs on a gene sequence are called splice junctions or splice sites as shown in Fig. 2.5, where the GT is called the donor and AG is called the acceptor splice site.

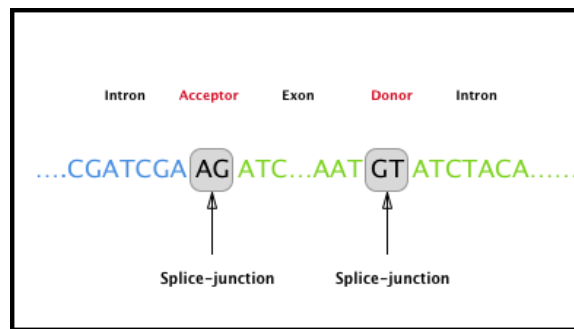


Figure 2.5: Splice-junction sites.

The problem of splice site recognition can be formulated by the following: Given an unclassified DNA sequence, classify it as one having a splice site or a non-splice site sequence. An efficient learning algorithm is required where the objective is to determine a classification rule that can accurately identify a region or pattern to distinguish a genomic DNA sequence.

Splice Site Detection Methods

Precise identification of Exon-Intron(EI) junctions or donors and Intron-Exon junctions(IE) or acceptors from a sequence is beneficial for advancements in transcriptome research and is a crucial step for fully understanding the gene expression. The accurate detection of splice junctions is challenging because of the high rate of false positives caused by the presence of short canonical splicing patterns [43].

There are currently two different techniques used to solve the splice junction prediction problem: Alignment based techniques and ML based. The sequence alignment-based techniques map millions of short RNA sequences produced by RNA-seq to the reference genome and then estimate where splicing occurs by identifying the adjacent exon locations. The existing alignment-based techniques such as SpliceMap [44] and TopHat [45] detect only canonical splice sites while missing the non-canonical sites which are required for accurate junction prediction. The ML based techniques can predict non-canonical sites as well by appropriate training. Different ML approaches have been used such as Support Vector Machines(SVM) [46, 47, 48, 49] Random Forest(RF) [50], Decision Trees (DT) [51], Naïve Bayesian(NB) [52], Markov Model [53] and AdaBoost [54] to identify splice or non-splice sites. Among them SVM models have been used very often due to their capability to handle high-dimensional datasets. However, certain kernel and penalty parameters in SVM require extensive tuning which is time consuming. The effectiveness of all these approaches also depends on the feature engineering technique used, which is a major initial step in solving a classification problem. Many feature engineering techniques have been proposed for feature construction directly from the DNA sequence, such as the MM1 (1-order Markov model) in [46] for feature construction from splice site sequences and using the SVM for prediction. The features were constructed based on different statistical approach [50] [47] with automated feature extraction proposed in [52] for prediction of splice sites. A length-variable Markov Model (LVMM) which produced higher accuracy with low time cost was discussed by [53]. A hybrid algorithm of AdaBoost classifier was proposed in [54] which provided an improvement in performance compared to the other approaches. The efficacy of all these approaches is based on the feature extraction step which is often a tedious task that is performed by domain experts. Manual operation of feature extraction often leads to incomplete representation or one-high dimensional feature space which will cause problems in

the ML process. The challenges involved in performing manual feature extraction and model training has led to a demand for DL based computational methods that performed automated feature representation. Many DL architectures were used and developed for the splice site prediction based on CNN [55, 56, 57, 58, 59, 60], RNN [43, 61, 62, 63, 64, 65, 66], Restricted Boltzmann Machines (RBM) [67], Autoencoders [63, 68] and Deep Belief Networks [67]. The steady performance of DL architectures has led to the design of many complex hybrid architectures [69, 70]. Although these DL architectures have removed the burden of manual feature extraction, they are still time consuming due to the manual effort present in designing efficient architectures. In general, the existing methods still utilise the manual effort in designing architectures which needs a lot of expert domain knowledge and is time consuming.

Comparative Analysis

In this section, the comparison of the reviewed studies is presented in Table 2.1. The criteria for comparison are the following:

1. The referenced study.
2. Datasets used in the study.
3. Type of DL architecture used.
4. Method of DNA representation used: One-hot Encoding, Word Embeddings, Orthogonal Encoding, Label Encoder, Orthonormal Sparse Encoding.
5. Metrics used for evaluation of the proposed approach.
6. Summary of the study.

Table 2.1: Comparative Analysis of Literature Review

Reference	Dataset used	DL method	Input representation	Reported classification metrics	Summary
[43]	UCSC database	RNN	One-hot Encoding	Accuracy	Proposed and tested RNN architectures to model DNA sequences and to detect splice junctions.
[55]	Splice Site Recognition (SSR) dataset	CNN	Label Encoder	Sensitivity, Specificity, Accuracy, AUC	Proposed a shallow version of ResNet CNN architecture for splice site classification.
[57]	Gencode database	Deep CNN	Label Encoder	Accuracy, Recall, Precision, F1-score	Proposed a deep CNN based approach for prediction of splice junctions.
[58]	Histone, Splice, Promoter	CNN	One-hot Encoding	Accuracy	Proposed a new approach for classifying DNA sequences using CNN by representing DNasequences as text data.
[59]	HS3D and GENCODE datasets	CNN	Orthonormal Sparse Encoding	Accuracy, Sensitivity, Specificity	Presented a deep learning based splice junction sequence classifier, named DeepSplice, based on CNN to classify candidate splice junctions.
[60]	CCDS, NCBI, TISdb datasets	CNN	One-hot Encoding	Accuracy, Sensitivity, Specificity, MCC	Proposed a DL architecture based on CNN for prediction of translation initiation sites.
[62]	Human, Cross-species, New pre-miRNAs	RNN	Word Embeddings	Sensitivity, Specificity, F-score, g-mean	Proposed a novel miRNA precursor prediction algorithm that can successfully reflect structural characteristics of precursor miRNAs.
[63]	Public human miRNA-mRNA pairing database	RNN	One-hot Encoding	Accuracy, Sensitivity, Specificity, F-score	Presents a DL framework for miRNA target prediction using deep recurrent neural networks-based auto-encoding.
[64]	MAQC-III	RNN	One-hot Encoding	Pearson, Spearman, r^2	Proposed a novel approach to model nucleotide sequences to calculate sequence-specific bias without pre determining sequence structures.
[65]	GENCODE Release 25	RNN-GRU	Word Embeddings	Accuracy, Sensitivity, Specificity, F-score, PPV, NPV	Implemented a gated RNN architecture that learns complex and long range patterns for predicting protein coding potential.
[66]	Ensembl human genome and UCSC-hg38 dataset	RNN	One-hot Encoding	Accuracy	Proposed a deep RNN based approach to classify a sequence is to be a coding or noncoding sequence.
[69]	HS3D database	CNN-BLSTM	One-hot Encoding	Accuracy, Sensitivity, Specificity, MCC	Proposed a hybrid architecture for splice site classification.
[70]	GENCODE dataset	CNN-RNN	One-hot Encoding	Accuracy, loss, R^2	Proposed a computational model, COSSMO, that accurately predicts competitive alternative splice site selection from raw DNA sequences.

Summary

The review of the literature on studies related to prediction of splice site junctions indicated that DL architectures were varying between RNN and CNN. Complex hybrid architectures were also proposed that required human expertise to design them through trial and error approaches. Observations guided the choice of methods, experiments and metrics used in this thesis.

In the reviewed studies, it was observed that selection of an appropriate DL architecture according to the capabilities and characteristics of the input data is crucial for success. This required further extensive tuning of the architecture parameters to better suit the problems in hand. Many architectures were adapted from computer vision to genomics by human experts by means of parameter sharing, multi-modal, multi-task and transfer learning approaches. The success of these architectures in computer vision is based on the notion that the object to be classified occupies a considerable part of the input image. However, in genomics the classification feature occupies only a tiny fraction the input. Also DL architectures are over-parametrized and the performance can be conditional if the models are not appropriately designed.

Despite the successful adaptation of models from other domains for solving problems in genomics, transfer learning approaches still pose challenges. The best practices for training and fine tuning the models are not concrete and depend on additional hyper-parameters for specific DL architectures. The task relatedness in multi-task learning and the similarity in data and domain in transfer learning methods are difficult to assess. The differences in domains requires fine tuning or pre-training the model from scratch and it does not guarantee expected results. In genomics domain, there is a compelling need to adopt methods to discover domain-specific architectures to address specific problems.

In the context of the literature review, the following choices were made:

1. The DARTS method was adapted to discover new high performance DL architecture for DNA splice site classification. The continuous approach based on DARTS approach was preferred as the NAO was proposed very recently.
2. The search space will be performed on CNN architectures as it was most used and performed well in recent studies.
3. The discovered model will be compared against fixed baseline architectures of CNN and RNN.
4. The following classification metrics are selected from the literature review to be computed: Accuracy, Sensitivity, Specificity, F score, AUC score
5. The discovered model will be investigated further to benchmark on computational performance.

CHAPTER 3: METHODOLOGY

In this chapter, we describe our representation of the input DNA sequence in Section 3.1. We then present the computational procedure for finding the best architecture cell in Section 3.2. Section 3.3 presents the methods and metrics used for the evaluation of the discovered architecture.

DNA Representation

The sequence data is biologically described using four types of nucleotides, adenine (A), cytosine (C), guanine (G) and thymine (T). Each of these sequences are converted into numerical representation using 1-dimensional orthogonal encoding or one-hot encoding for downstream analysis. However, to shape the input appropriately for the DARTS CNN model, the DNA sequences are represented as a 3-dimensional tensor, which is similar in shape to an image tensor input to image classification networks. Firstly, one-hot encoding is applied which converts each nucleotide in the DNA sequence of length n_d into a four-dimensional vector and then concatenates each of them to form the complete sequence. The next step is to transform the list of one-hot vectors to a 3-dimensional tensor.

let $s \in S$ where $S = \{A, T, C, G\}$, then, a sequence (A,C,G,T,A,C) will be encoded into a tuple of 4-D binary vectors as shown in Fig. 3.1.

$$([1,0,0,0],[0,0,1,0],[0,0,0,1],[0,1,0,0],[1,0,0,0],[0,0,1,0])$$

The encoded sequence is then represented as a three-dimensional matrix of shape $(3 \times n_d \times 4)$. The final representation of the input to the model will be in the form (batch size $\times 3 \times n_d \times 4$)

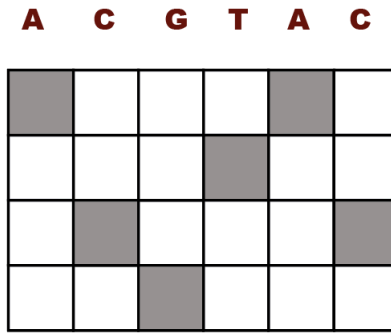


Figure 3.1: DNA representation by one-hot vector.

DARTS Algorithm

The DARTS technique discovers state-of-the-art network architectures by formulating the task in a differential manner. The interesting part in this method is that the search space is treated as continuous rather than searching over a discrete set of architectures in the search space.

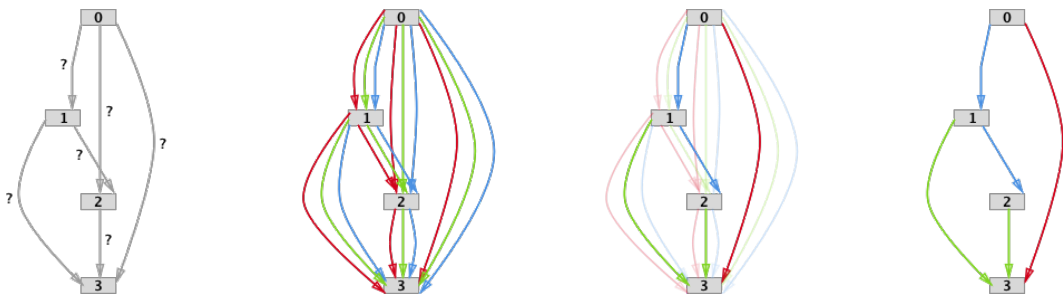


Figure 3.2: DARTS method.

The cell in the architecture is considered as a Direct Acyclic Graph (DAG) consisting of a set of nodes and edges. Each cell has one output node and two input nodes. Let N be the number of nodes and each node represented by x^i . Each edge (i, j) performs an operation represented by $o^{(i,j)}$ that transforms x^i . The intermediate nodes are computed based on its predecessors.

$$x^i = \sum_{j < i} (o^{i,j} x^j) \quad (3.1)$$

The learning of the cell involves learning the operations that transform the input. Fig. 3.2 shows an overview of the DARTS method [6]. The goal of the method is to find a cell that forms the building block of the final architecture. Initially the operations on the edges are unknown. Let O be the set of operations where each operation is represented as $o(\cdot)$ to be applied to x^i . The choice of the operation is made in a continuous manner by performing a softmax on all possible operations.

$$o^{-(i,j)}(x) = \sum_{o \in O} \frac{\exp(\alpha_0^{(i,j)})}{\sum_{o' \in O} \exp(\alpha_0^{(i,j)})} o(x) \quad (3.2)$$

Here $\alpha_0^{(i,j)}$ is a vector with dimension $|O|$ that indicates the mixing of operation between a pair of nodes. The architecture search phase jointly performs learning on a set of continuous variables $\alpha = \{\alpha^{(i,j)}\}$ and the weights (ω) within each operation in O . The value of α and ω is obtained through a bi-level optimization algorithm where α becomes the higher level variable and ω acts as the lower level variable. The search finds a value of α that minimizes the validation loss L_{val} for that value of ω that minimizes the training loss L_{train} .

$$\min_{\alpha} L_{valid}(\omega^*(\alpha), \alpha) \quad (3.3)$$

$$s.t \quad \omega^*(\alpha) = \operatorname{argmin}_{\omega} L_{train}(\omega, \alpha) \quad (3.4)$$

This bi-level optimization algorithm 1 shows the optimization of ω and α in the respective search spaces through a gradient-based approach. The operation at each edge is replaced by the operation that had the maximum value of α .

The optimization is performed in the architecture search algorithm 2 during the training phase and the best discrete architecture $arch_{final}$ is saved to be evaluated on the

Algorithm 1: Optimization algorithm of ω w.r.t α

- 1 Let $o^{-(i,j)}$ be the set of operations parameterized by $\alpha^{(i,j)}$ for an edge *pair*(i, j)
 - 2 **while not converged do**
 - 3 compute ω by decreasing $\nabla_{\omega} L_{train}(\omega, \alpha)$;
 - 4 compute α by decreasing $\nabla_{\alpha} L_{valid}(\omega - \xi \nabla_{\omega} L_{train}(\omega, \alpha), \alpha)$;
 - 5 Replace $o^{-(i,j)}$ with $o^{(i,j)} = \operatorname{argmax}_{o \in O} \alpha_o^{i,j}$ for each pair of edge(i, j)
-

architecture evaluation algorithm 3. The training phase in the architecture evaluation phase has a fixed architecture $arch_{final}$ and is then trained to obtain the optimal architecture weights. The trained model is evaluated on unseen data in the architecture evaluation phase.

Algorithm 2: Architecture Search Algorithm

Phase1: Architecture Search

Input : $x_s = (x_{s,1}, x_{s,2}, x_{s,3}, \dots, x_{s,|x_s|})$ $x_s \in X_s$
where X : a set of sequences with $|X|=N$
 x_s : a single sequence with length $|x_s|$
 $x_{(s,i)} \in \{A, T, C, G\}$ for $i = 1, 2, 3, \dots, |x_s|$
 y : label for x_s . $y \in \{0, 1\}$
where 0 means non-splice and 1 represents splice sequence

Output: $arch_{final}$

where $arch_{final}$ is the final architecture cell

```
/* Data Preprocessing and Loading */
1  $X_T \leftarrow transform(X_s)$ ; where  $X_T$ : Transformed dataset (Section 3.1)
2  $(x_t, x_v) \leftarrow split(X_T)$ ; where  $x_t$ : Training set and  $x_v$ : Validation set
3 initialize best accuracy  $acc_b$ ;
4 for each epoch do
   /* Training */
5   for  $m, n$  training data randomly selected from  $x_t, x_v$  do
6     initialize new architecture  $arch(\omega, \alpha)$  using Algorithm 1;
7     train( $arch(\omega, \alpha)$ ); // Training the model
   /* Validation */
8   for  $p$  validation data randomly selected from  $x_v$  do
9      $acc_{val} = valid(arch(\omega, \alpha))$ ; // Validating the model
10  if  $acc_{val} > acc_b$  then
11     $acc_b = acc_{val}$ ;
12     $arch_{final} = arch(\omega, \alpha)$ ;
13 save  $arch_{final}$  for architecture evaluation phase;
```

Algorithm 3: Architecture Evaluation Algorithm

Phase2: Architecture Evaluation

Input : $x_s = (x_{s,1}, x_{s,2}, x_{s,3}, \dots, x_{s,|x_s|})$ $x_s \in X_s$
where X : a set of sequences with $|X|=N$
 x_s : a single sequence with length $|x_s|$
 $x_{(s,i)} \in \{A, T, C, G\}$ for $i = 1, 2, 3, \dots, |x_s|$
 y : label for x_s . $y \in \{0, 1\}$
where 0 means non-splice and 1 represents splice sequence

Output: acc_{final}

where acc_{final} is the final accuracy

```
/* Data Preprocessing and Loading */
1  $X_T \leftarrow transform(X_s)$ ; where  $X_T$ : Transformed dataset (Section 3.1)
2  $(x_t, x_v, x_f) \leftarrow split(X_T)$ ; where  $x_t$ : Training set,  $x_v$ : Validation set and  $x_f$ : Test set
3 initialize best accuracy  $acc_b$ ;
4 initialize weights  $\omega$ ;
5 initialize architecture  $arch$  to  $arch_{final}$  obtained from Algorithm 2;
6 for each epoch do
    /* Training */
7     for  $m$  training data randomly selected from  $x_t$  do
8         train( $arch(\omega, \alpha)$ ); // Training the model
9         update weights  $\omega$  using SGD with momentum;
    /* Validation */
10    for  $p$  validation data randomly selected from  $x_v$  do
11         $acc_{val} = valid(arch(\omega, \alpha))$ ; // Validating the trained model
12    if  $acc_{val} > acc_b$  then
13         $acc_b = acc_{val}$ ;
14        save best model  $arch(\omega, \alpha)$ ;
15  $acc_{final} = test(arch(\omega, \alpha))$ ; // Testing the final model
```

Performance Evaluation Method

The performance of DARTS model was compared using the metrics described in the Section 3.31 and Section 3.3.2. The results proved that DARTS model outperformed other fixed DL architectures. The benchmarking metrics were used to evaluate the model on multiple GPU and CPU architectures.

Classification Performance Metrics

There are several metrics used for assessing the classification performance. These metrics have been derived from the confusion matrix as shown in Fig. 3.3. The terminologies used in the confusion matrix are described in the following

		Actual	
		Positive	Negative
Predicted	Positive	True Positive	False Positive
	Negative	False Negative	True Negative

Figure 3.3: Confusion matrix.

- **Condition Positives (P):** The total identified number of samples that have the feature.
- **Condition Negatives (N):** The total identified number of samples that does not have the feature.
- **True Positives (TP):** The total identified number of samples predicted to have the feature, and in fact they have the feature.
- **True Negatives (TN):** The total identified number of samples predicted to not have the feature, and in fact they do not have the feature.
- **False Positives (FP):** The total identified number of samples predicted to have the feature, and in fact they do not have the feature.
- **False Negatives (FN):** The total identified number of samples predicted to not have the feature, and in fact they have the feature.

- **Accuracy:** The percentage of samples that were correctly classified by the model

$$Accuracy = \frac{TP + TN}{TP + TN + FP + FN} \quad (3.5)$$

- **Sensitivity:** The sensitivity is the measure of the model to correctly classify samples with the feature.

$$Sensitivity = \frac{TP}{TP + FN} \quad (3.6)$$

- **Specificity:** The specificity is the measure of the model to correctly classify samples without the feature.

$$Specificity = \frac{TN}{TN + FP} \quad (3.7)$$

- **F-score:** The weighted average of the measure of precision and recall.
- **Area Under the ROC Curve(AUC):** The measure of the performance of the model across all possible classification thresholds.

Computational Performance Metrics

The resources in high-performance computing (HPC) are formed by a heterogeneous hardware. The GPU is a parallel computing coprocessor that are specialized in accelerating vector computations. Training of DL models are mostly performed on GPU due to large number of vector computations that take place and CPU is used for scalar operations. However, the CPU-based benchmarking is included for users that run DL workloads in a CPU environment. The performance of DARTS model was benchmarked on multiple GPU and CPU architectures and the time taken for architecture search and architecture evaluation on each were noted. The learning

and inference speed of the trained model was compared on the different GPU architectures over three types of datatype: single precision, double precision and half precision.

CHAPTER 4: IMPLEMENTATION

The methodology makes use of deep CNN to distinguish features between true and false splice junctions. CNN architectures have shown better performance in learning features that classify actual splice sites from false ones. The method consists of two stages: Architecture Search and Architecture Evaluation. In the first stage, architecture search using DARTS was performed to discover the best model and the second stage validates the discovered model on a held out unseen data. The model gains from the information present in the genomic sequence of the candidate splice junction to accurately classify whether the sequence corresponds to a splice junction or not.

Dataset

The experiments were performed using Splice Site Recognition (SSR) dataset that is publicly available at [71]. The underlying problem posed in this dataset is to classify, given a sequence of DNA, as a splice or a non-splice sequence. The splice junctions are locations in a DNA sequence where ‘superfluous’ DNA is removed during protein creation process. The beginning and end of an exon is determined by the splice-donor and splice-acceptor sequences present. In this study, the prediction of splice junctions are performed using the given annotated DNA sequences with true acceptor splice site sequences. The original dataset has 159,771 true acceptor splice site sequences and 14,868,555 non-acceptor sequences. To avoid the class imbalance problem we used all the true acceptor splice site sequences and equal number of non-acceptor sequences chosen randomly. The dataset used in our study has a total of 319542 DNA sequence samples with a sequence length of 141 base pair. The experiments were repeated 10 times for better performance validation and robustness of the trained model. In each of the experiment, the dataset was randomly split

into three sets from the complete dataset using customized data loaders into train, validation and test datasets respectively of size 0.6, 0.2, 0.2.

Experimental Setup

The architecture search and evaluation experiments were performed on Sidra HPC environment using NVIDIA Tesla K40M. The implementation was done using Pytorch which is an open-source ML library in python based on Torch that supports strong GPU acceleration.

The computational benchmarking experiments were performed on Intel(R) Xeon(R) x86_64, Quadro K4100M and Tesla V100-SXM2 architectures. The specification of the hardware architectures used is listed in Table 4.1. The GPUs and CPUs were configured with the same environment as the original experiments. The CPU evaluation was performed by submitting the task as a job to the cluster environment.

Table 4.1: Hardware for Benchmarking

Device Model	Number of available cores	Memory size(in GB)
Tesla V100-SXM2	5120	16
Tesla K40m	2880	12
Quadro K4100M	1152	4
Intel(R) Xeon(R) CPU E5-2670	32	256

Architecture Search

The following operations were only included from a rich primitive space used in [6] for our search space O : 3×3 separable convolutions, 3×3 dilated separable convolutions, 3×3 max pooling, 3×3 average pooling and zero. A building block of a convo-

lution operation is formed by three steps: Firstly an activation function is Rectifier Linear Unit (RELU) applied and then a convolution operation (CN) is executed and finally, a batch normalization (BN) is performed. This is denoted as ReLU-Conv-BN and we used the same ReLU-Conv-BN order in [6] for performing convolutional operations. Our discovered convolutional cell consisted of 4 nodes, where the output node is result of the depth-wise concatenation of the convolution and pooling layers excluding the input node. The final architecture network was formed by stacking multiple cells together. The architecture consists of two types of convolutional cells called normal cell and reduction cell to make it scalable for any input size. When a feature map is taken as input, the normal cell returns a feature map of same dimension. The reduction cell returns a feature map where the height and width are reduced by a factor of two. The reduction cells are located at $1/3$ and $2/3$ of the total depth of the architecture. The architecture has a reduction cell in every third cell of the complete architecture. The first and second input nodes of the cell k are set to the $k-2$ and $k-1$ cells respectively. The Figures 4.1 to 4.4 shows the discovered normal and reduction cells during the first 4 epochs in the architecture search experiment.

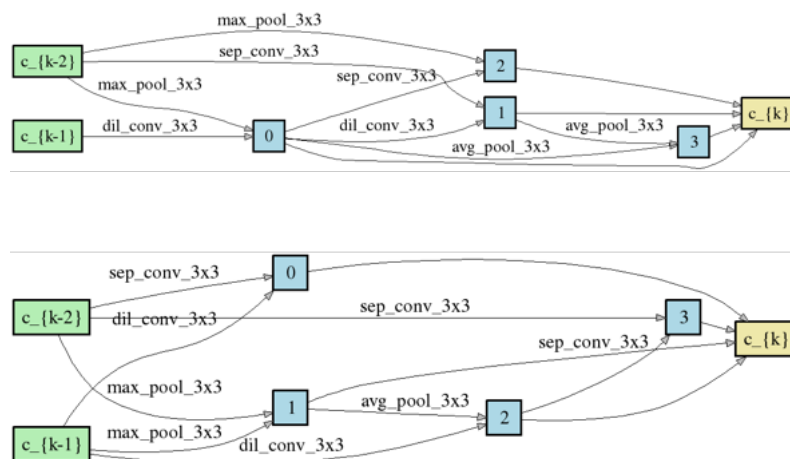


Figure 4.1: Normal and Reduction cell in Epoch 1.

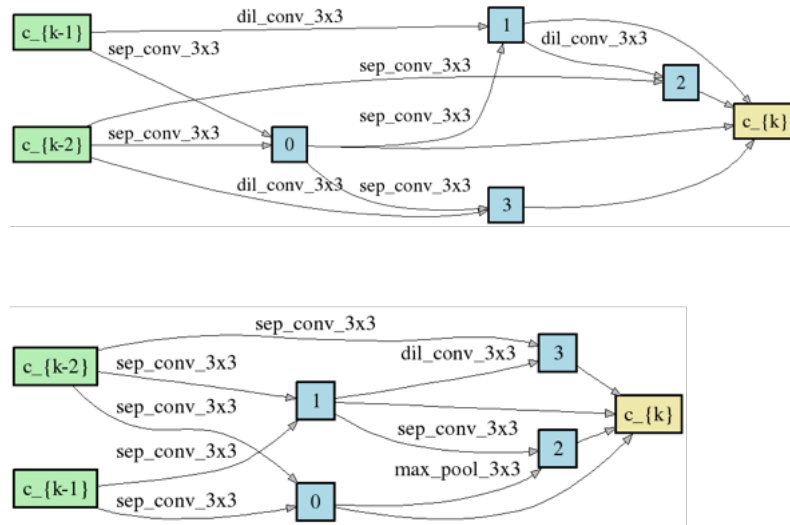


Figure 4.2: Normal and Reduction cell in Epoch 2.

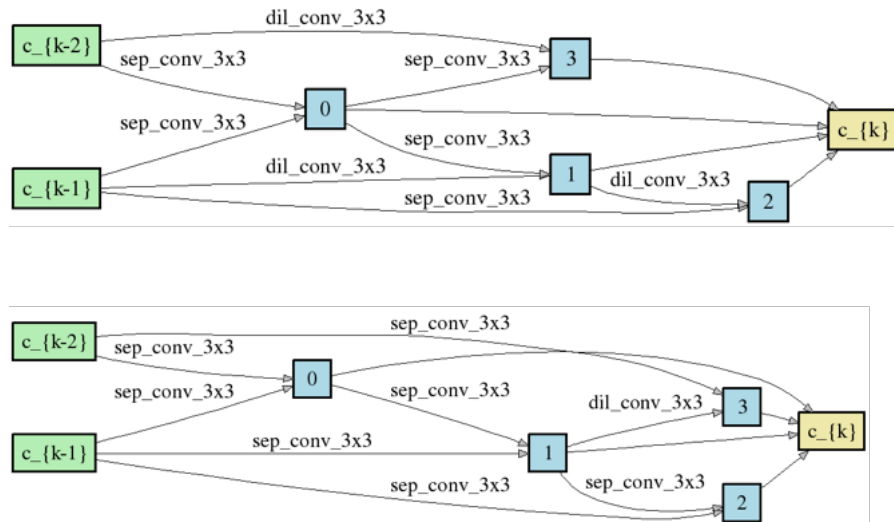


Figure 4.3: Normal and Reduction cell in Epoch 3.

The architecture search was performed on train and validation datasets. A network composed of 8 cells were trained for 50 epochs using DARTS with batch size 500 set for both training and validation. The weights ω were optimized using SGD with momentum and Adam as the optimizer for architecture variables. The initial learning rate was set as 0.0025 and was gradually decreased to a minimum of 0.001. The rest

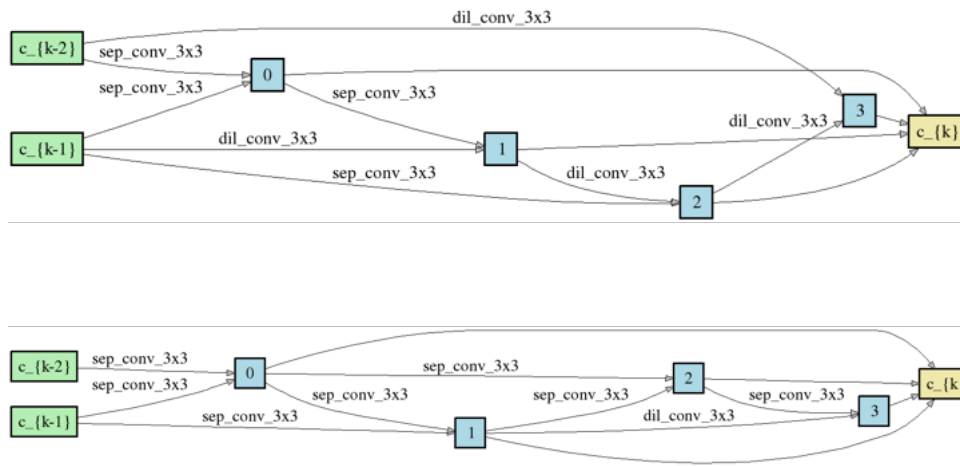


Figure 4.4: Normal and Reduction cell in Epoch 4.

of the hyperparameters were chosen similar to the original implementation in [6] as shown in Table 4.2.

Table 4.2: Hyperparameters for architecture search

Hyperparameter	Definition	Value
Batch size	Number of samples propagated through the network	500
Initial learning rate	The initial rate which influences the current value of weights of the model during training	0.0003
Architecture learning rate	The learning rate for architecture encoding	0.0025
Minimum learning rate	The minimal value to which the learning rate will decrease during model training	0.001
Epochs	Number of training iterations of the model	50
Weight decay rate	A regularization rate to improve the generalization performance of neural networks	0.0003
Loss function	The function used to evaluate the set of weights	Cross Entropy
Update strategy	The optimization technique used to update the model parameters	SGD with momentum

Architecture Evaluation

The best architecture cells shown in Fig. 4.5 were selected based on the performance on validation dataset. In order to select the optimal architecture for evaluation, the search experiment was run ten times with different seeds. The best performing cell was recorded during epoch 29 in the ninth iteration. A network of 3 cells were trained

for 40 epochs with batch size 500. The rest of the hyperparameters were similar to the ones used for the architecture search process. The selected best architecture was evaluated using a held out test dataset. It is important to note that the test set was never used during the architecture search process. As the results were subject to variance even for same configurations, the mean and standard deviation of 10 independent runs was reported.

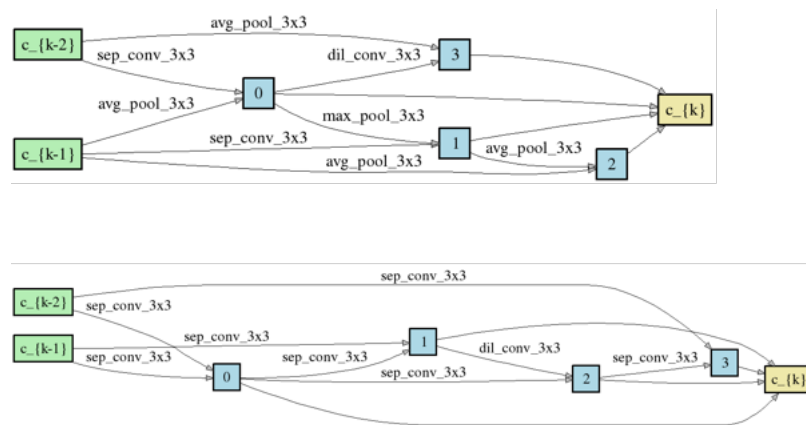


Figure 4.5: Normal and Reduction cell learned on splice dataset.

CHAPTER 5: RESULTS

In this chapter we present the results of the benchmarking experiments conducted. Section 5.1 presents the classification performance of DARTS model with five established models in DL. In Section 5.2, the computational performance of the model is benchmarked on GPU and CPU.

Classification Performance Results

The splice dataset classification results using various DL techniques are presented in Table 5.1. The average and standard deviation metrics were calculated over 10 repetitions of each experiment. The DARTS experiment was repeated by running the experiments on 10 different seeds. The experiments on the baseline models were repeated by k-fold cross-validation. The performance of the automatically discovered architecture was compared against fixed baseline CNN and RNN models. The baseline models were implemented using keras framework to benchmark against known models from the literature. Notably, DARTS outperformed over the widely known LSTM and baseline CNN with and without embedding in terms of test accuracy, sensitivity, specificity, F-score and AUC score. The model achieved better accuracy results over the hybrid and LSTM model with embedding.

Table 5.1: Comparison of Model Performance

Model	Accuracy	Sensitivity	Specificity	F-score	AUC score
DARTS	94.15 ± 0.12	94.00 ± 0.10	95.20 ± 0.30	94 ± 0.10	94.8 ± 0.01
Baseline LSTM with embeddings	93.98 ± 0.11	95 ± 1.09	92.32 ± 1.25	93.74 ± 0.25	93.66 ± 0.27
Hybrid(RNN+CNN)	93.66 ± 0.27	94.63 ± 0.78	93.33 ± 0.63	94.02 ± 0.14	93.66 ± 0.27
Baseline LSTM	86.99 ± 14.65	85.26 ± 19.81	88.71 ± 9.80	85.87 ± 16.93	86.99 ± 14.65
Baseline CNN	64.15 ± 0.63	65.27 ± 6.00	63.04 ± 6.22	64.43 ± 2.16	64.10 ± 0.50
Baseline CNN with embeddings	53.63 ± 3.65	95.62 ± 6.61	11.63 ± 13.77	67.35 ± 0.51	53.63 ± 3.65

Fig. 5.1 and Fig. 5.2 shows the mean and standard deviation plots of training and validation metrics over 40 epochs for all the baseline models against our proposed model. Notably, the fixed CNN baseline architectures performed poorly compared to other architectures.

Computational Performance Results

Execution Time

In addition, the proposed model was benchmarked on GPU systems and CPU. The execution time for performing architecture search and architecture evaluation were calculated on different device architectures and the results are presented in Table 5.2. Notably, the most advanced Tesla V100 GPU completed the search in less than 11 hours and the evaluation in half an hour. Fig. 5.3 shows the plot of search and evaluation execution time of DARTS model on each of the devices.

Table 5.2: Comparison of Execution Time

Device Model	Architecture Search(hours)	Architecture Evaluation(hours)
Tesla V100-SXM2	10.75	0.5
Tesla K40m	38.5	2
Quadro K4100M	101	50
Intel(R) Xeon(R) CPU E5-2670	526	74

Precision

The learning and inference speed of the trained model were compared on different GPU architectures. The experiments were performed on single precision, half precision, double precision data types. The model was fed with a single batch input of 500 sequences. For training, the time required for 20 forward and backward passes

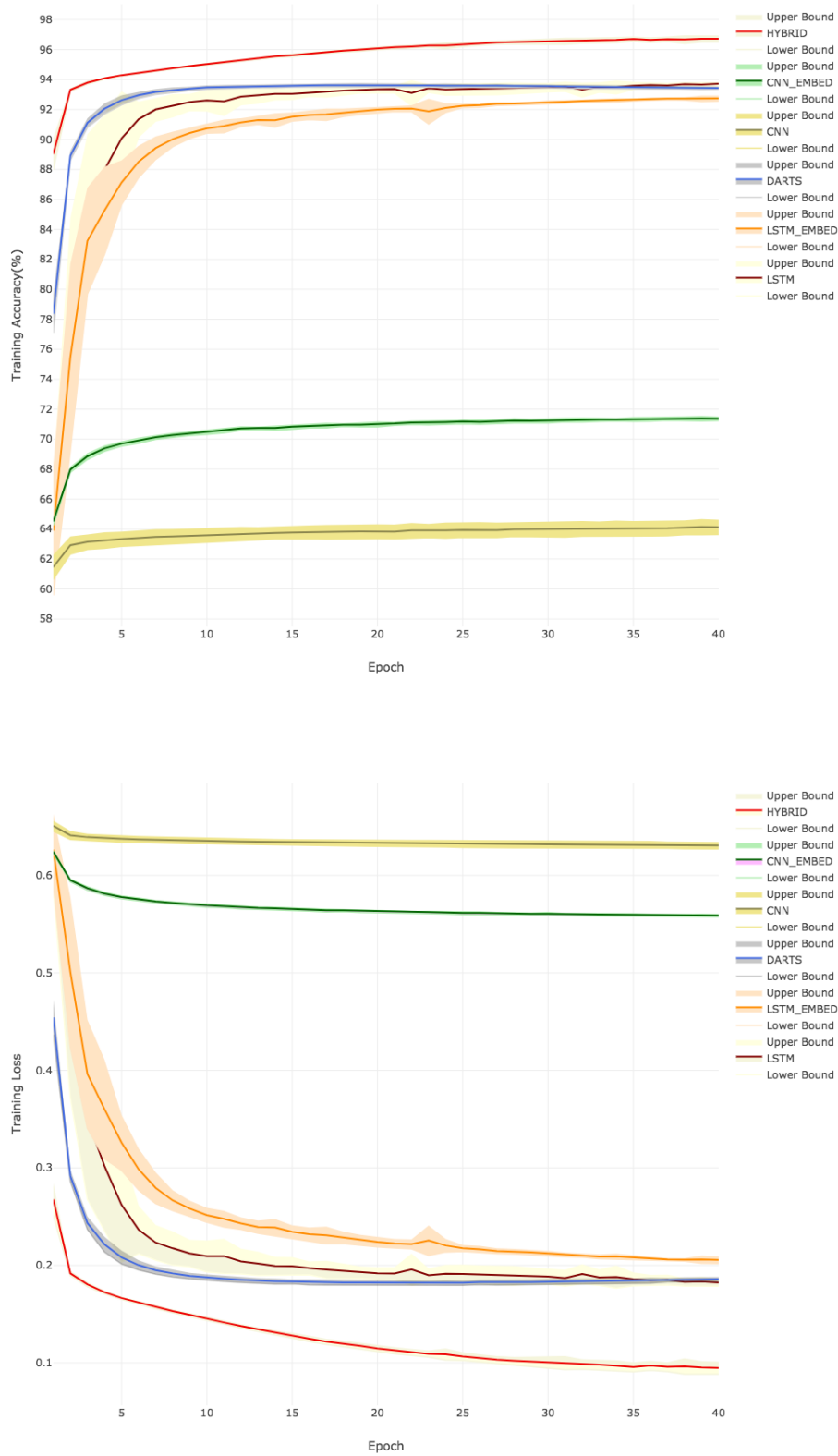


Figure 5.1: Plots of training accuracy and loss.

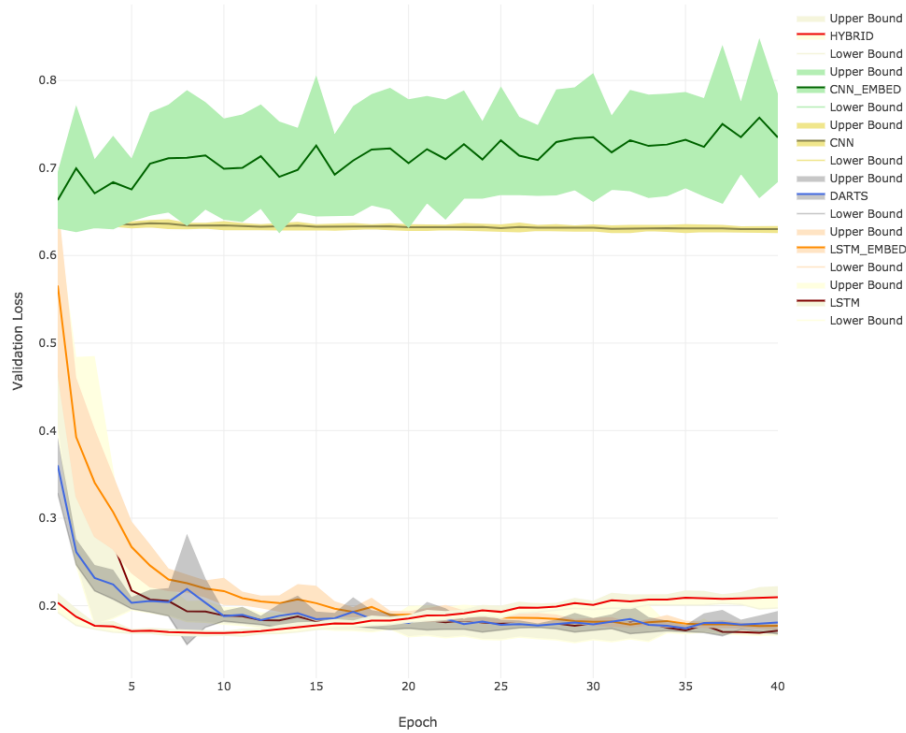
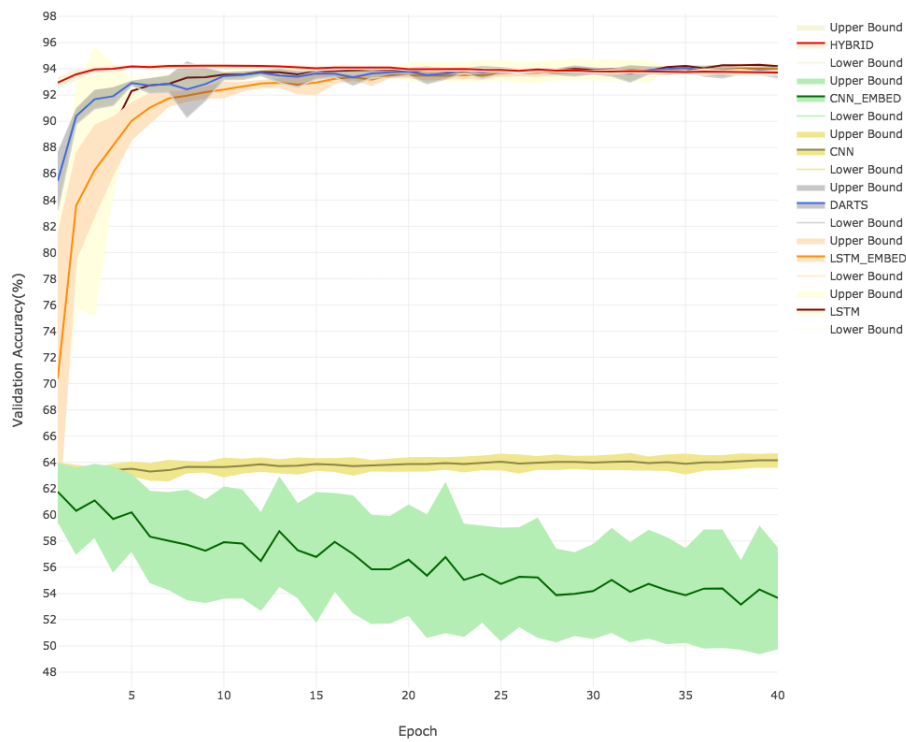


Figure 5.2: Plots of validation accuracy and loss.

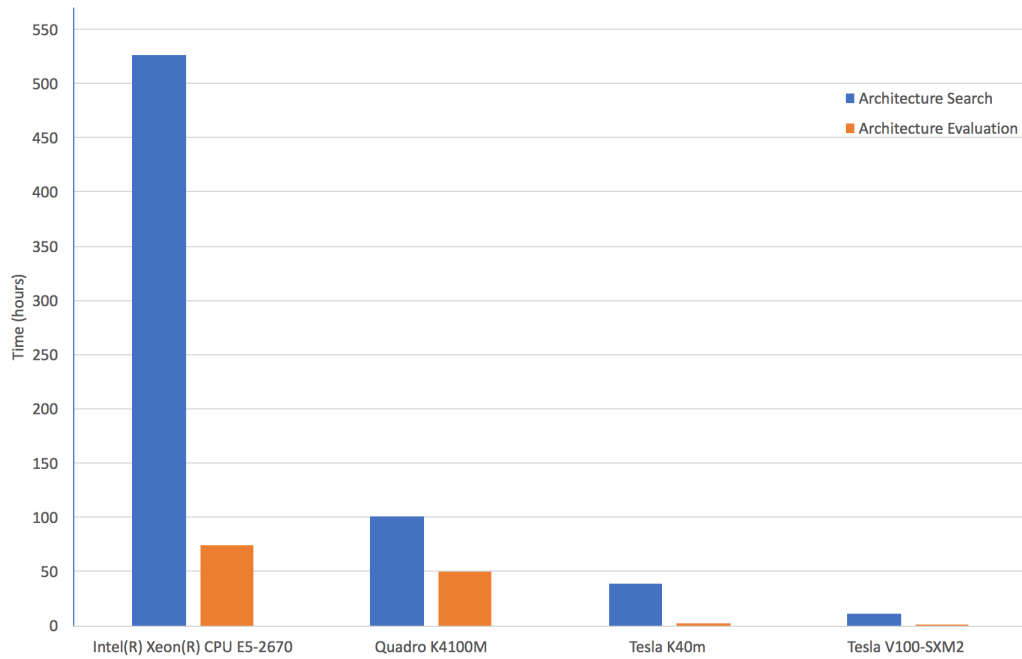


Figure 5.3: Plot of execution time for DARTS.

were averaged. In inference, time duration of 20 forward passes were averaged. Five warm up steps were included that was not calculated towards the final results. Fig. 5.4 shows the plot of training and inference speed of DARTS model on each of the devices.

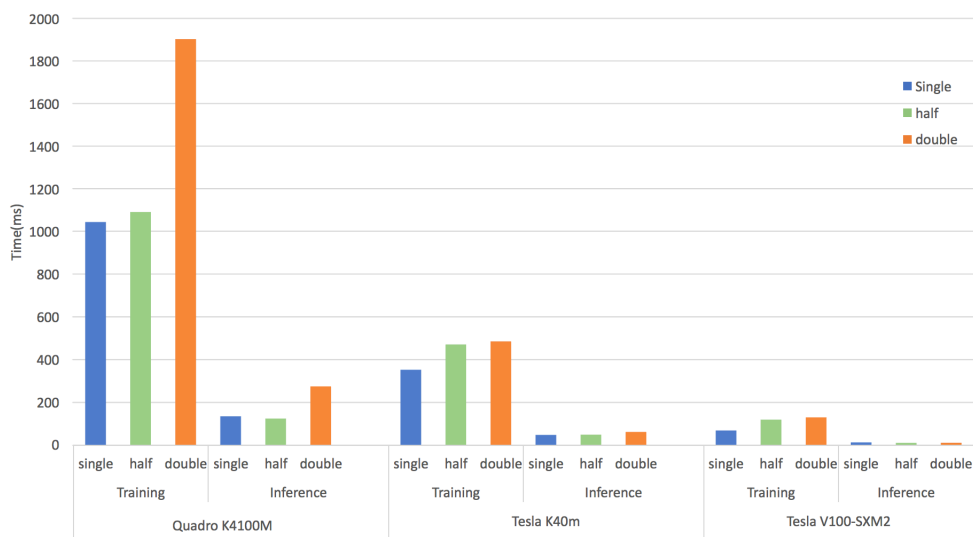


Figure 5.4: Plot of comparison of training and inference speed.

Table 5.3: Comparison of Learning and Inference Speed on GPUs

GPU Model	Training			Inference		
	Single	Half	Double	Single	Half	Double
Quadro K4100M	1041.24	1090.38	1902.38	134.97	124.59	274.78
Tesla K40m	352.53	470.72	485.17	48.022	49.38	62.13
Tesla V100-SXM2	68.46	119.55	130.09	13.21	11.02	11.07

CHAPTER 6: CONCLUSION AND FUTURE WORK

Deep Learning is an emerging research topic among the genomics community. Its applications can be revolutionized by introducing high-performance computing methods to analyze datasets in the field of gene therapies, molecular diagnostics and personalized medicine. In the scope of this thesis, an advanced DL approach based on differential architecture search was implemented to solve the splice site classification problem in genomics and to discover new high performance CNN architectures. It was observed in the literature review that most of the work was focused on using manually designed architectures in ML and DL. This study has aided in bridging the gap between the state-of-the art in DL and its application to genomics. The evaluation results showed that the newly discovered architecture outperformed the fixed baseline DL architectures using the same dataset. The architecture was compared alongside the well-known LSTM model and complex hybrid architectures. Furthermore, the discovered architecture was evaluated on multiple CPU and GPU architectures. The total time taken for performing the architecture search and evaluation were determined as well as the floating point instructions per second for single, double and half precision were compared. The computational benchmarking results obtained proved that there is significant improvement in execution time when using advanced GPU architectures.

For all its promises, DL in genomics still possess a number of challenges. The results largely depend on the quality of the data input that are well annotated so that the model can learn to distinguish features and identify patterns. Another challenge is the lack of judgement capability where the technique is able to distinguish from a biologically relevant variation and normal variations. This would require applying further experimental design and controls. The advancements in the field of DL in the field of computer vision and speech recognition has led to new methods being constantly proposed that awaits its application in genomics domain. Furthermore, the

availability of quasi-unlimited storage at a reasonable price, the surge in computing power and the lower computational costs will allow these advanced DL techniques to reshape the capabilities of machines to completely understand and interpret the human genome.

As future steps, the plan is to further improve the performance of DARTS based on CNN by including more primitives such as skip connect and higher convolutional operations, thereby widening the architecture search space. This will help to traverse more information to lower layers. The DARTS approach will further be evaluated against the recent parallel work of NAO which also performs continuous optimization of architecture space. The study showed that fixed RNN architectures have better results than CNN. Genomics data are sequential data similar to speech recognition, natural language processing and language translation. It would be interesting to implement DARTS to search for a recurrent cell that can be recursively connected to form a RNN that can be applied for tasks of protein function prediction. As part of performance benchmarking, the plan is to evaluate the models further on GPU clusters and Google's Tensor Processing Unit (TPU) which are specialized hardware architectures developed to accelerate AI workloads. In addition, this approach will be tested on future genomics classification tasks, as it will be highly useful to uncover new insights from the vast available sequencing data.

REFERENCES

- [1] J. D. Watson, F. H. Crick, *et al.*, “Molecular structure of nucleic acids,” *Nature*, vol. 171, no. 4356, pp. 737–738, 1953.
- [2] P. V. Jithesh and V. Scaria, “From genomes to genomic medicine: enabling personalized and precision medicine in the middle east,” *Personalized Medicine*, vol. 14, no. 5, pp. 377–382, 2017. PMID: 29754564.
- [3] T. Ching, D. S. Himmelstein, B. K. Beaulieu-Jones, A. A. Kalinin, B. T. Do, G. P. Way, E. Ferrero, P.-M. Agapow, M. Zietz, M. M. Hoffman, *et al.*, “Opportunities and obstacles for deep learning in biology and medicine,” *Journal of The Royal Society Interface*, vol. 15, no. 141, p. 20170387, 2018.
- [4] A. Krizhevsky, I. Sutskever, and G. E. Hinton, “Imagenet classification with deep convolutional neural networks,” in *Advances in neural information processing systems*, pp. 1097–1105, 2012.
- [5] K. He, X. Zhang, S. Ren, and J. Sun, “Deep residual learning for image recognition,” in *Proceedings of the IEEE conference on computer vision and pattern recognition*, pp. 770–778, 2016.
- [6] H. Liu, K. Simonyan, and Y. Yang, “Darts: Differentiable architecture search,” *arXiv preprint arXiv:1806.09055*, 2018.
- [7] P. Baldi, P. Sadowski, and D. Whiteson, “Searching for exotic particles in high-energy physics with deep learning,” *Nature communications*, vol. 5, p. 4308, 2014.
- [8] G. B. Goh, N. O. Hodas, and A. Vishnu, “Deep learning for computational chemistry,” *Journal of computational chemistry*, vol. 38, no. 16, pp. 1291–1307, 2017.

- [9] A. Esteva, B. Kuprel, R. A. Novoa, J. Ko, S. M. Swetter, H. M. Blau, and S. Thrun, “Dermatologist-level classification of skin cancer with deep neural networks,” *Nature*, vol. 542, no. 7639, p. 115, 2017.
- [10] W. S. McCulloch and W. Pitts, “A logical calculus of the ideas immanent in nervous activity,” *The bulletin of mathematical biophysics*, vol. 5, no. 4, pp. 115–133, 1943.
- [11] Y. Bengio *et al.*, “Learning deep architectures for ai,” *Foundations and trends® in Machine Learning*, vol. 2, no. 1, pp. 1–127, 2009.
- [12] M. K. Leung, H. Y. Xiong, L. J. Lee, and B. J. Frey, “Deep learning of the tissue-regulated splicing code,” *Bioinformatics*, vol. 30, no. 12, pp. i121–i129, 2014.
- [13] H. Y. Xiong, B. Alipanahi, L. J. Lee, H. Bretschneider, D. Merico, R. K. Yuen, Y. Hua, S. Gueroussov, H. S. Najafabadi, T. R. Hughes, *et al.*, “The human splicing code reveals new insights into the genetic determinants of disease,” *Science*, vol. 347, no. 6218, p. 1254806, 2015.
- [14] B. E. Bejnordi, M. Veta, P. J. Van Diest, B. Van Ginneken, N. Karssemeijer, G. Litjens, J. A. Van Der Laak, M. Hermsen, Q. F. Manson, M. Balkenhol, *et al.*, “Diagnostic assessment of deep learning algorithms for detection of lymph node metastases in women with breast cancer,” *Jama*, vol. 318, no. 22, pp. 2199–2210, 2017.
- [15] J. Lanchantin, R. Singh, Z. Lin, and Y. Qi, “Deep motif: Visualizing genomic sequence classifications,” *arXiv preprint arXiv:1605.01133*, 2016.
- [16] J. Zhou and O. G. Troyanskaya, “Deep supervised and convolutional generative stochastic network for protein secondary structure prediction,” *arXiv preprint arXiv:1403.1347*, 2014.

- [17] D. R. Kelley, J. Snoek, and J. L. Rinn, “Basset: learning the regulatory code of the accessible genome with deep convolutional neural networks,” *Genome research*, vol. 26, no. 7, pp. 990–999, 2016.
- [18] H. Zeng, M. D. Edwards, G. Liu, and D. K. Gifford, “Convolutional neural network architectures for predicting dna–protein binding,” *Bioinformatics*, vol. 32, no. 12, pp. i121–i127, 2016.
- [19] B. Alipanahi, A. DeLong, M. T. Weirauch, and B. J. Frey, “Predicting the sequence specificities of dna-and rna-binding proteins by deep learning,” *Nature biotechnology*, vol. 33, no. 8, p. 831, 2015.
- [20] M. Schuster and K. K. Paliwal, “Bidirectional recurrent neural networks,” *IEEE Transactions on Signal Processing*, vol. 45, no. 11, pp. 2673–2681, 1997.
- [21] S. Hochreiter and J. Schmidhuber, “Long short-term memory,” *Neural computation*, vol. 9, no. 8, pp. 1735–1780, 1997.
- [22] K. Cho, B. Van Merriënboer, C. Gulcehre, D. Bahdanau, F. Bougares, H. Schwenk, and Y. Bengio, “Learning phrase representations using rnn encoder-decoder for statistical machine translation,” *arXiv preprint arXiv:1406.1078*, 2014.
- [23] V. Boža, B. Brejová, and T. Vinař, “Deepnano: deep recurrent neural networks for base calling in minion nanopore reads,” *PloS one*, vol. 12, no. 6, p. e0178751, 2017.
- [24] D. Quang and X. Xie, “Danq: a hybrid convolutional and recurrent deep neural network for quantifying the function of dna sequences,” *Nucleic acids research*, vol. 44, no. 11, pp. e107–e107, 2016.

- [25] A. Busia, J. Collins, and N. Jaitly, “Protein secondary structure prediction using deep multi-scale convolutional neural networks and next-step conditioning,” *arXiv preprint arXiv:1611.01503*, 2016.
- [26] P. Vincent, H. Larochelle, I. Lajoie, Y. Bengio, and P.-A. Manzagol, “Stacked denoising autoencoders: Learning useful representations in a deep network with a local denoising criterion,” *Journal of machine learning research*, vol. 11, no. Dec, pp. 3371–3408, 2010.
- [27] S. Rifai, P. Vincent, X. Muller, X. Glorot, and Y. Bengio, “Contractive autoencoders: Explicit invariance during feature extraction,” in *Proceedings of the 28th International Conference on International Conference on Machine Learning*, pp. 833–840, Omnipress, 2011.
- [28] C. Poultney, S. Chopra, Y. L. Cun, *et al.*, “Efficient learning of sparse representations with an energy-based model,” in *Advances in neural information processing systems*, pp. 1137–1144, 2007.
- [29] D. P. Kingma and M. Welling, “Auto-encoding variational bayes,” *arXiv preprint arXiv:1312.6114*, 2013.
- [30] L. Rampasek, D. Hidru, P. Smirnov, B. Haibe-Kains, and A. Goldenberg, “Dr. vae: Drug response variational autoencoder,” *arXiv preprint arXiv:1706.08203*, 2017.
- [31] T. Chen, R. Xu, Y. He, and X. Wang, “A gloss composition and context clustering based distributed word sense representation model,” *Entropy*, vol. 17, no. 9, pp. 6007–6024, 2015.
- [32] S. Hijazi, R. Kumar, and C. Rowen, “Using convolutional neural networks for image recognition,” *Cadence Design Systems Inc.: San Jose, CA, USA*, 2015.

- [33] C. Szegedy, W. Liu, Y. Jia, P. Sermanet, S. Reed, D. Anguelov, D. Erhan, V. Vanhoucke, and A. Rabinovich, “Going deeper with convolutions,” in *Proceedings of the IEEE conference on computer vision and pattern recognition*, pp. 1–9, 2015.
- [34] H. Liu, K. Simonyan, O. Vinyals, C. Fernando, and K. Kavukcuoglu, “Hierarchical representations for efficient architecture search,” *arXiv preprint arXiv:1711.00436*, 2017.
- [35] E. Real, A. Aggarwal, Y. Huang, and Q. V. Le, “Regularized evolution for image classifier architecture search,” *arXiv preprint arXiv:1802.01548*, 2018.
- [36] B. Zoph and Q. V. Le, “Neural architecture search with reinforcement learning,” *arXiv preprint arXiv:1611.01578*, 2016.
- [37] B. Zoph, V. Vasudevan, J. Shlens, and Q. V. Le, “Learning transferable architectures for scalable image recognition,” *arXiv preprint arXiv:1707.07012*, vol. 2, no. 6, 2017.
- [38] P. D. Summers, “A methodology for lisp program construction from examples,” *Journal of the ACM (JACM)*, vol. 24, no. 1, pp. 161–175, 1977.
- [39] B. Baker, O. Gupta, R. Raskar, and N. Naik, “Accelerating neural architecture search using performance prediction,” *arXiv preprint arXiv:1705.10823*, 2017.
- [40] A. Brock, T. Lim, J. M. Ritchie, and N. Weston, “Smash: one-shot model architecture search through hypernetworks,” *arXiv preprint arXiv:1708.05344*, 2017.
- [41] H. Pham, M. Y. Guan, B. Zoph, Q. V. Le, and J. Dean, “Efficient neural architecture search via parameter sharing,” *arXiv preprint arXiv:1802.03268*, 2018.

- [42] R. Luo, F. Tian, T. Qin, E. Chen, and T.-Y. Liu, “Neural architecture optimization,” in *Advances in neural information processing systems*, pp. 7816–7827, 2018.
- [43] B. Lee, T. Lee, B. Na, and S. Yoon, “Dna-level splice junction prediction using deep recurrent neural networks,” *arXiv preprint arXiv:1512.05135*, 2015.
- [44] K. F. Au, H. Jiang, L. Lin, Y. Xing, and W. H. Wong, “Detection of splice junctions from paired-end rna-seq data by splicemap,” *Nucleic acids research*, vol. 38, no. 14, pp. 4570–4578, 2010.
- [45] C. Trapnell, L. Pachter, and S. L. Salzberg, “Tophat: discovering splice junctions with rna-seq,” *Bioinformatics*, vol. 25, no. 9, pp. 1105–1111, 2009.
- [46] A. K. Baten, B. C. Chang, S. K. Halgamuge, and J. Li, “Splice site identification using probabilistic parameters and svm classification,” in *BMC bioinformatics*, vol. 7, p. S15, BioMed Central, 2006.
- [47] P. K. Meher, T. K. Sahu, A. Rao, and S. Wahi, “Identification of donor splice sites using support vector machine: a computational approach based on positional, compositional and dependency features,” *Algorithms for molecular biology*, vol. 11, no. 1, p. 16, 2016.
- [48] Y. Zhang, C.-H. Chu, Y. Chen, H. Zha, and X. Ji, “Splice site prediction using support vector machines with a bayes kernel,” *Expert Systems with Applications*, vol. 30, no. 1, pp. 73–81, 2006.
- [49] D. Wei, W. Zhuang, Q. Jiang, and Y. Wei, “A new classification method for human gene splice site prediction,” in *International Conference on Health Information Science*, pp. 121–130, Springer, 2012.

- [50] P. K. Meher, T. K. Sahu, and A. R. Rao, "Prediction of donor splice sites using random forest with a new sequence encoding approach," *BioData mining*, vol. 9, no. 1, p. 4, 2016.
- [51] H. S. Lopes, C. R. Erig Lima, and N. J. Murata, "A configware approach for high-speed parallel analysis of genomic data," *Journal of Circuits, Systems, and Computers*, vol. 16, no. 04, pp. 527–540, 2007.
- [52] U. Kamath, K. De Jong, and A. Shehu, "Effective automated feature construction and selection for classification of biological sequences," *PloS one*, vol. 9, no. 7, p. e99982, 2014.
- [53] Q. Zhang, Q. Peng, Q. Zhang, Y. Yan, K. Li, and J. Li, "Splice sites prediction of human genome using length-variable markov model and feature selection," *Expert Systems with Applications*, vol. 37, no. 4, pp. 2771–2782, 2010.
- [54] E. Pashaei, A. Yilmaz, M. Ozen, and N. Aydin, "Prediction of splice site using adaboost with a new sequence encoding approach," in *Systems, Man, and Cybernetics (SMC), 2016 IEEE International Conference on*, pp. 003853–003858, IEEE, 2016.
- [55] R. Elsousy, N. Kathiresan, and S. Boughorbel, "On the depth of deep learning models for splice site identification," *bioRxiv*, p. 380667, 2018.
- [56] X. Du, Y. Yao, Y. Diao, H. Zhu, Y. Zhang, and S. Li, "Deepss: Exploring splice site motif through convolutional neural network directly from dna sequence," *IEEE Access*, vol. 6, pp. 32958–32978, 2018.
- [57] Y. Zhang, X. Liu, J. N. MacLeod, and J. Liu, "Deepsplice: Deep classification of novel splice junctions revealed by rna-seq," in *2016 IEEE International Conference on Bioinformatics and Biomedicine (BIBM)*, pp. 330–333, IEEE, 2016.

- [58] N. G. Nguyen, V. A. Tran, D. L. Ngo, D. Phan, F. R. Lumbanraja, M. R. Faisal, B. Abapihi, M. Kubo, and K. Satou, “Dna sequence classification by convolutional neural network,” *Journal of Biomedical Science and Engineering*, vol. 9, no. 05, p. 280, 2016.
- [59] Y. Zhang, X. Liu, J. MacLeod, and J. Liu, “Discerning novel splice junctions derived from rna-seq alignment: a deep learning approach,” *BMC Genomics*, vol. 19, no. 1, p. 971, 2018.
- [60] J. Zuallaert, M. Kim, Y. Saeys, and W. De Neve, “Interpretable convolutional neural networks for effective translation initiation site prediction,” in *2017 IEEE International Conference on Bioinformatics and Biomedicine (BIBM)*, pp. 1233–1237, IEEE, 2017.
- [61] S. T. Kothen-Hill, A. Zviran, R. C. Schulman, S. Deochand, F. Gaiti, D. Maloney, K. Y. Huang, W. Liao, N. Robine, N. D. Omans, *et al.*, “Deep learning mutation prediction enables early stage lung cancer detection in liquid biopsy,” 2018.
- [62] S. Park, S. Min, H. Choi, and S. Yoon, “deepmirgene: deep neural network based precursor microrna prediction,” *arXiv preprint arXiv:1605.00017*, 2016.
- [63] B. Lee, J. Baek, S. Park, and S. Yoon, “deeptarget: end-to-end learning framework for microrna target prediction using deep recurrent neural networks,” in *Proceedings of the 7th ACM International Conference on Bioinformatics, Computational Biology, and Health Informatics*, pp. 434–442, ACM, 2016.
- [64] Y.-z. Zhang, R. Yamaguchi, S. Imoto, and S. Miyano, “Sequence-specific bias correction for rna-seq data using recurrent neural networks,” *BMC genomics*, vol. 18, no. 1, p. 1044, 2017.

- [65] S. T. Hill, R. Kuintzle, A. Teegarden, E. Merrill III, P. Danaee, and D. A. Hendrix, “A deep recurrent neural network discovers complex biological rules to decipher rna protein-coding potential,” *Nucleic acids research*, vol. 46, no. 16, pp. 8105–8113, 2018.
- [66] H. Bae, B. Lee, S. Kwon, and S. Yoon, “Dna steganalysis using deep recurrent neural networks,” *arXiv preprint arXiv:1704.08443*, 2017.
- [67] T. Lee and S. Yoon, “Boosted categorical restricted boltzmann machine for computational prediction of splice junctions,” in *International Conference on Machine Learning*, pp. 2483–2492, 2015.
- [68] Z.-C. Xu, P. Wang, W.-R. Qiu, and X. Xiao, “iss-pc: Identifying splicing sites via physical-chemical properties using deep sparse auto-encoder,” *Scientific Reports*, vol. 7, no. 1, p. 8222, 2017.
- [69] T. Naito, “Human splice-site prediction with deep neural networks,” *Journal of Computational Biology*, vol. 25, no. 8, pp. 954–961, 2018.
- [70] H. Bretschneider, S. Gandhi, A. G. Deshwar, K. Zuberi, and B. J. Frey, “Cossmo: predicting competitive alternative splice site selection using deep learning,” *Bioinformatics*, vol. 34, no. 13, pp. i429–i437, 2018.
- [71] S. Sonnenburg, G. Rätsch, C. Schäfer, and B. Schölkopf, “Large scale multiple kernel learning,” *Journal of Machine Learning Research*, vol. 7, no. Jul, pp. 1531–1565, 2006.

APPENDIX: DETAILED RESULTS

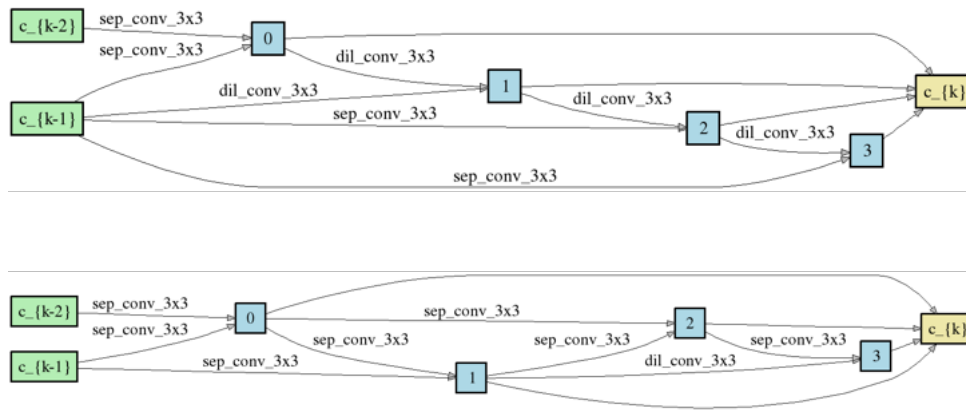


Figure A.1: Normal and Reduction cell in Epoch 6.

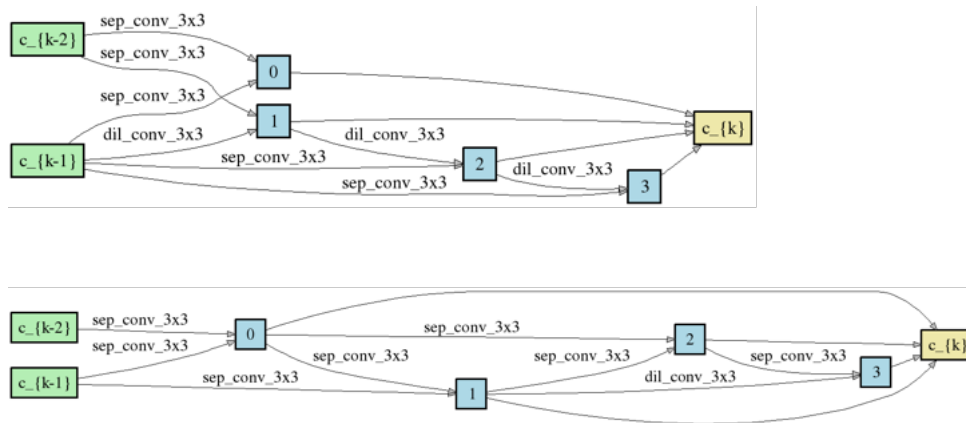


Figure A.2: Normal and Reduction cell in Epoch 7.

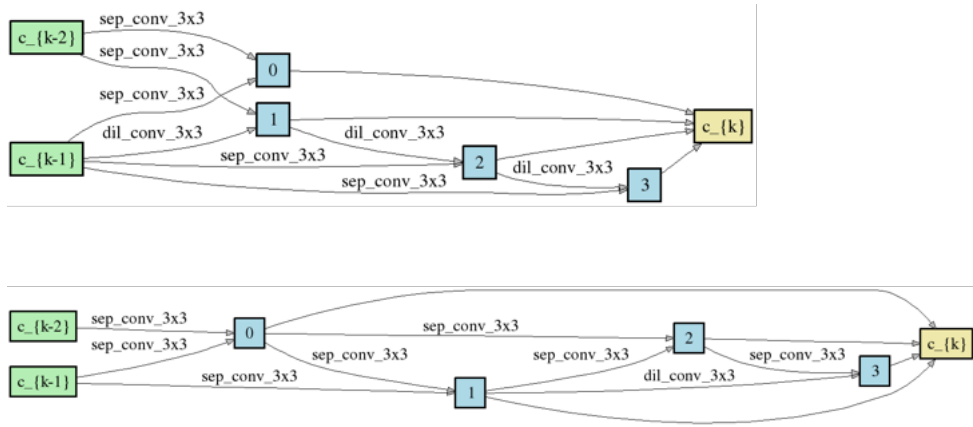


Figure A.3: Normal and Reduction cell in Epoch 8.

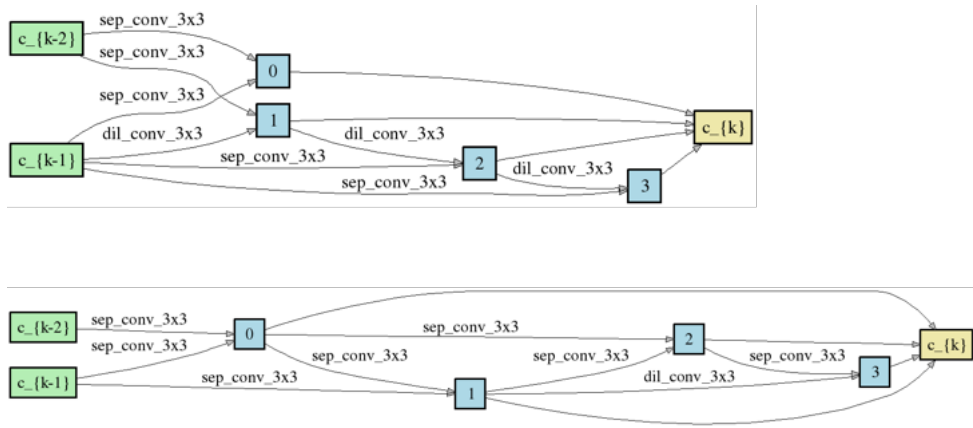


Figure A.4: Normal and Reduction cell in Epoch 9.

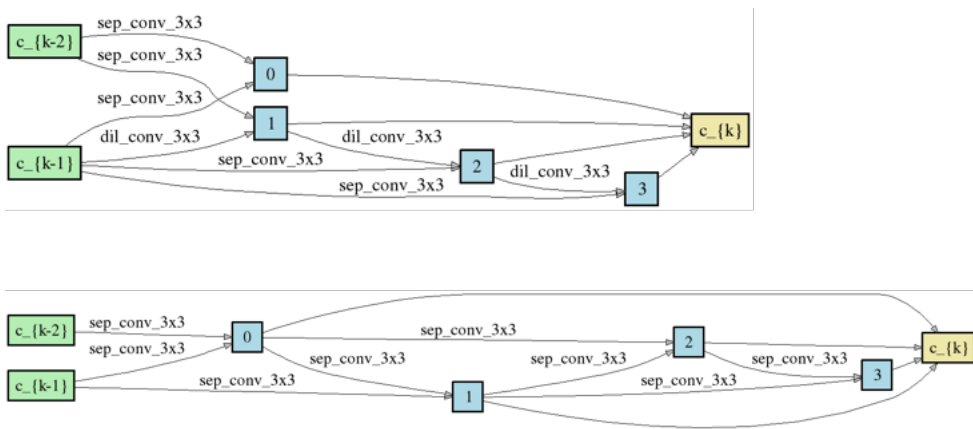


Figure A.5: Normal and Reduction cell in Epoch 10.

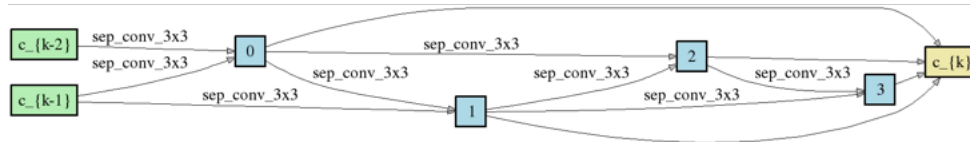
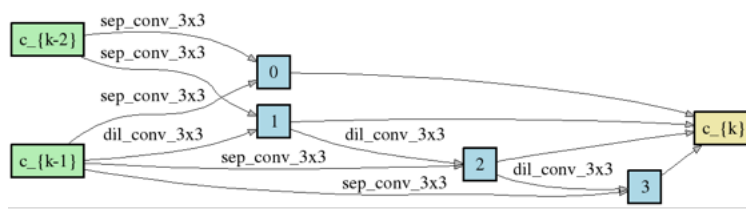


Figure A.6: Normal and Reduction cell in Epoch 11.

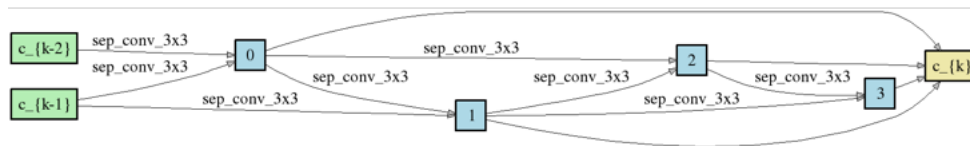
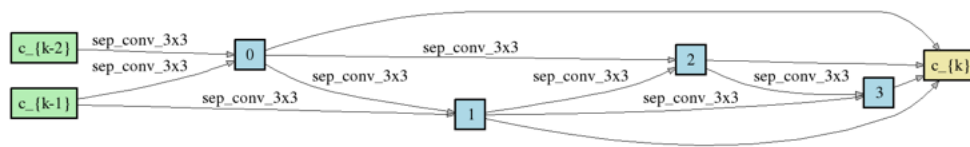


Figure A.7: Normal and Reduction cell in Epoch 12.

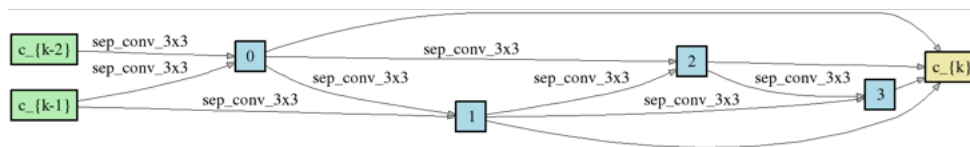
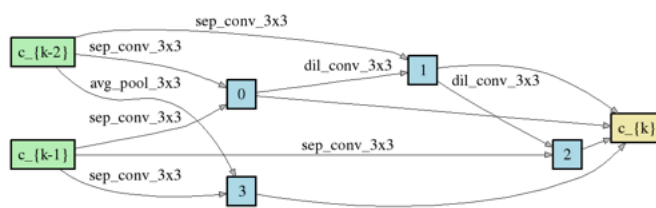


Figure A.8: Normal and Reduction cell in Epoch 13.

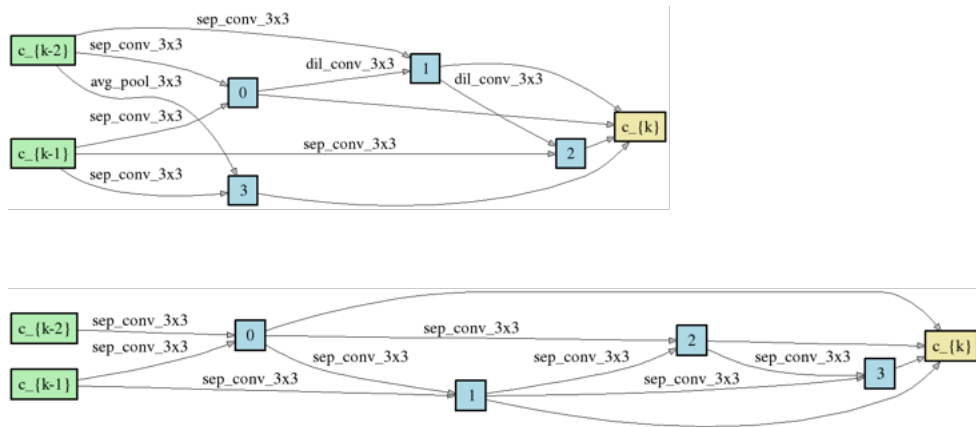


Figure A.9: Normal and Reduction cell in Epoch 14.

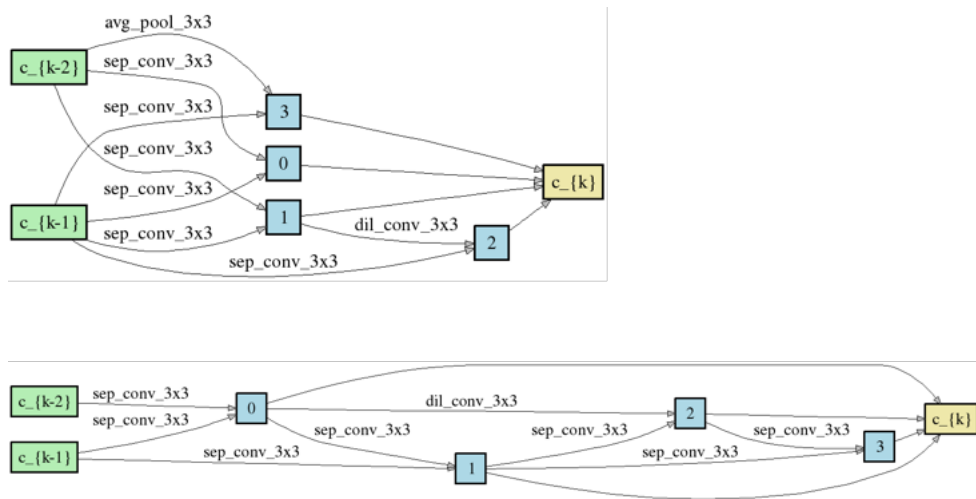


Figure A.10: Normal and Reduction cell in Epoch 15.

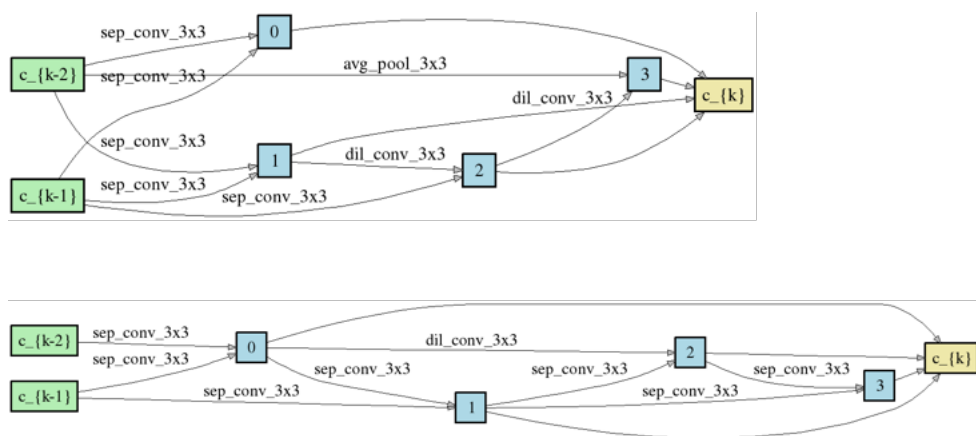


Figure A.11: Normal and Reduction cell in Epoch 16.

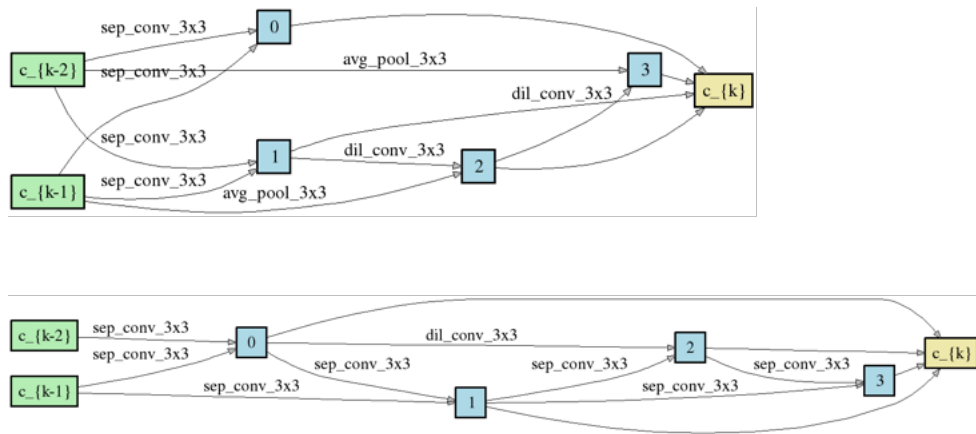


Figure A.12: Normal and Reduction cell in Epoch 17.

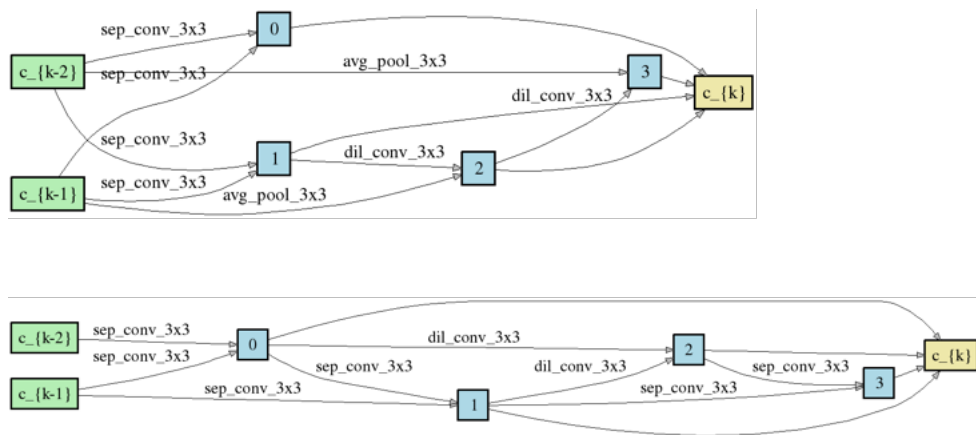


Figure A.13: Normal and Reduction cell in Epoch 18.

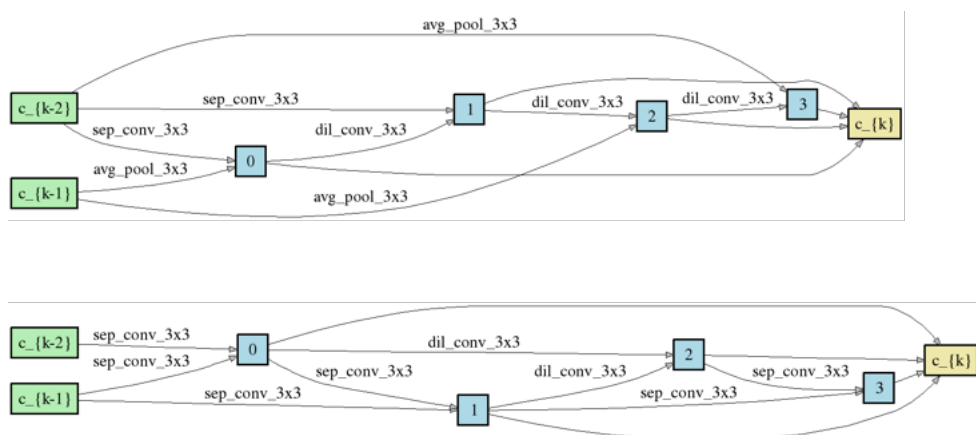


Figure A.14: Normal and Reduction cell in Epoch 19.

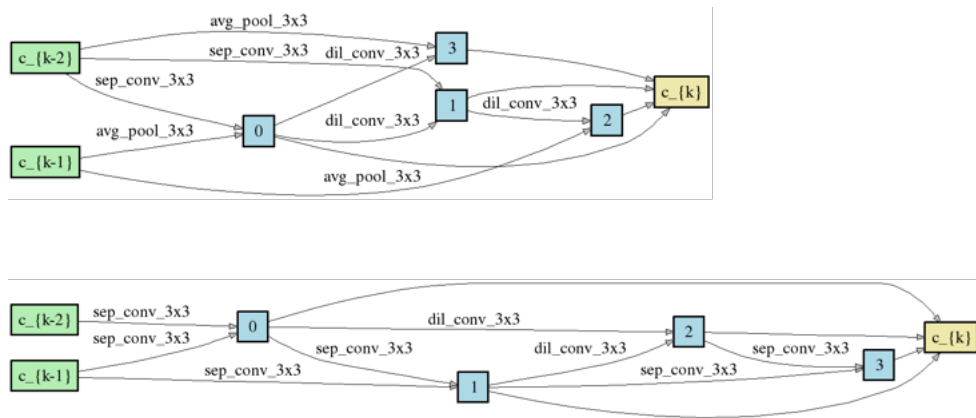


Figure A.15: Normal and Reduction cell in Epoch 20.

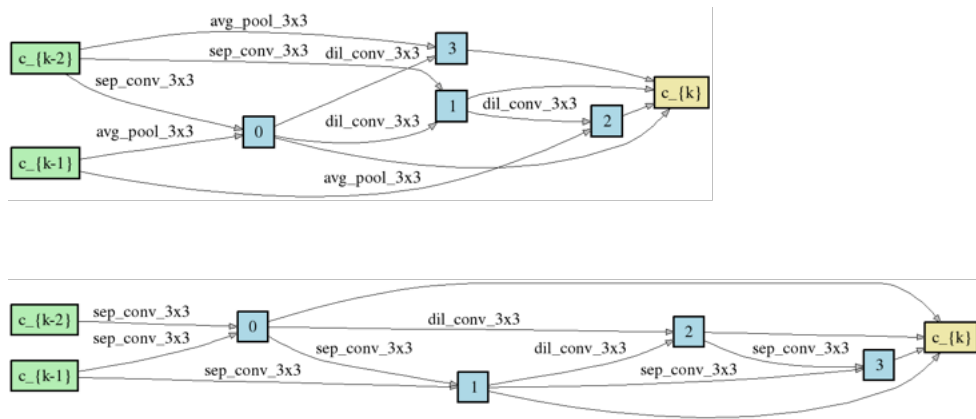


Figure A.16: Normal and Reduction cell in Epoch 21.

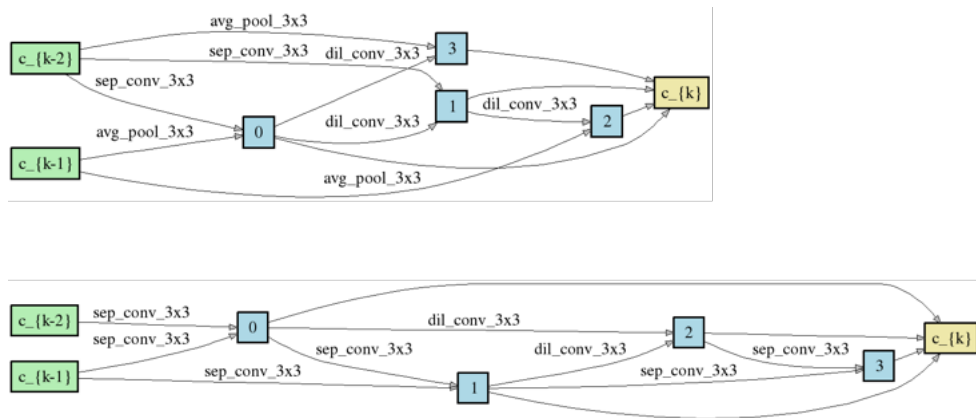


Figure A.17: Normal and Reduction cell in Epoch 22.

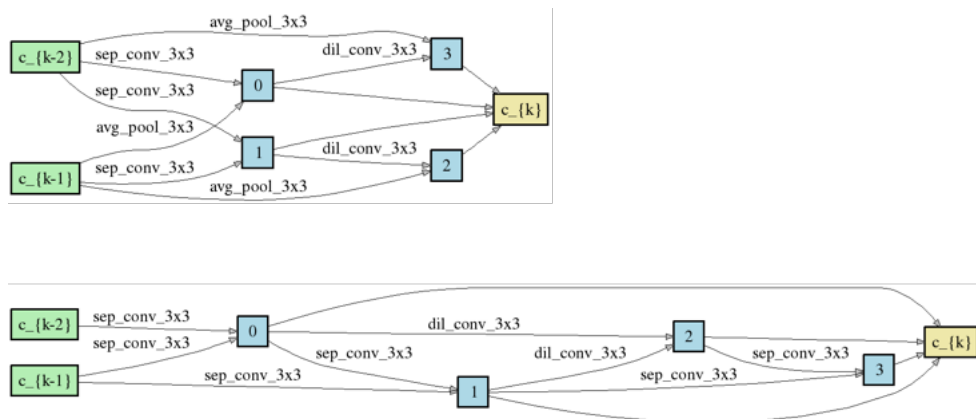


Figure A.18: Normal and Reduction cell in Epoch 23.

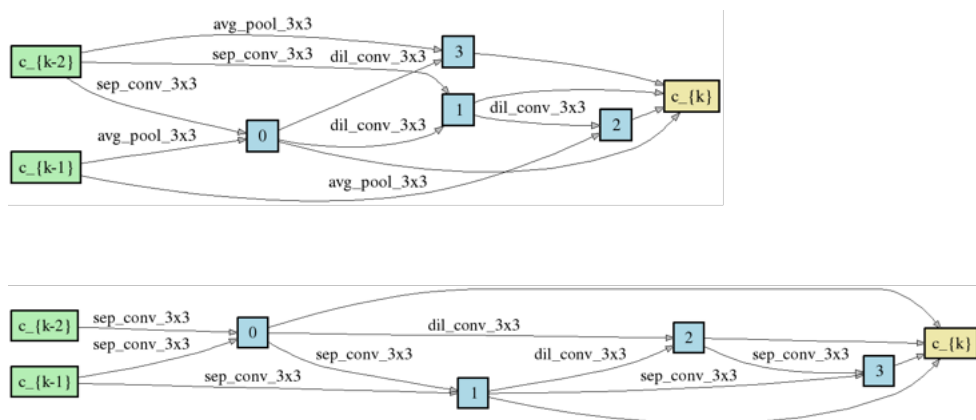


Figure A.19: Normal and Reduction cell in Epoch 24.

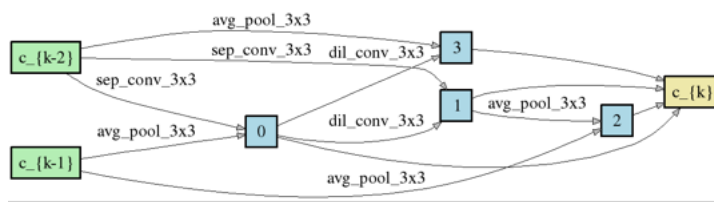


Figure A.20: Normal and Reduction cell in Epoch 25.

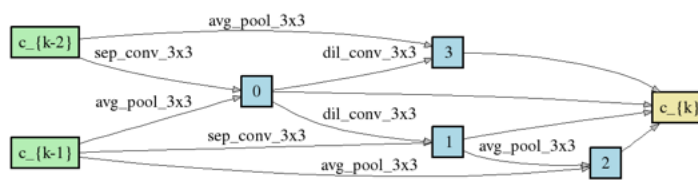


Figure A.21: Normal and Reduction cell in Epoch 26.

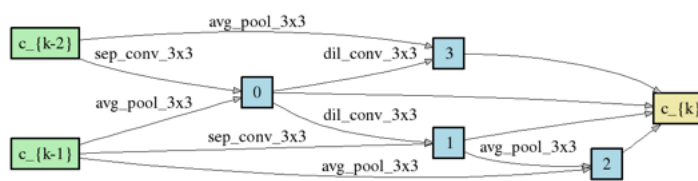


Figure A.22: Normal and Reduction cell in Epoch 27.

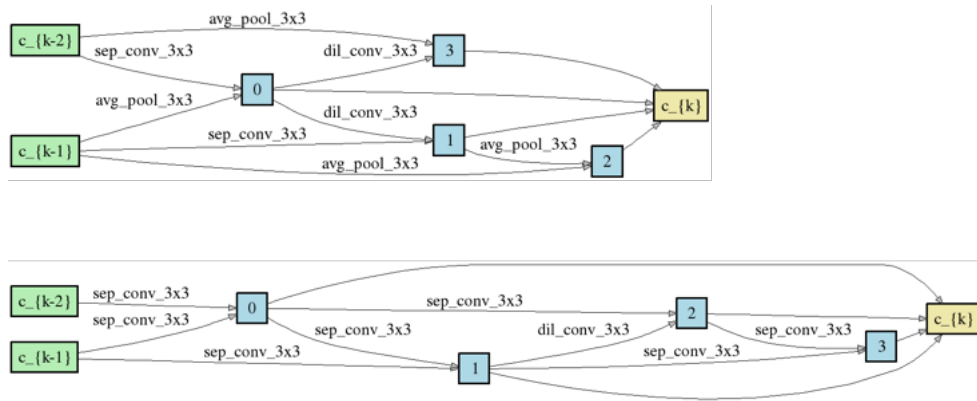


Figure A.23: Normal and Reduction cell in Epoch 28.

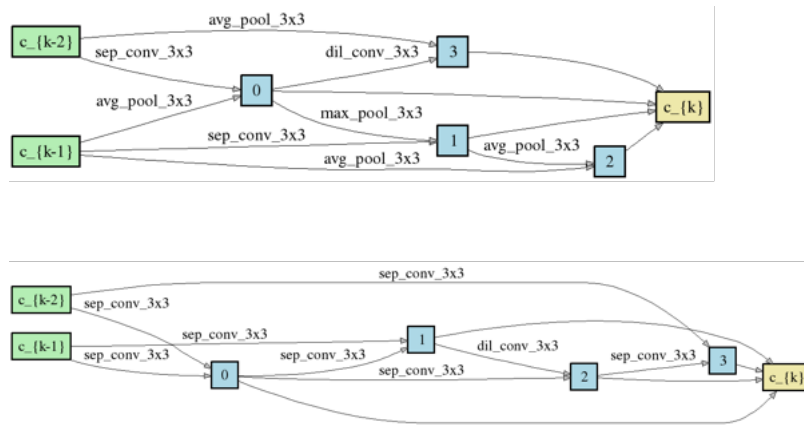


Figure A.24: Normal and Reduction cell in Epoch 29.

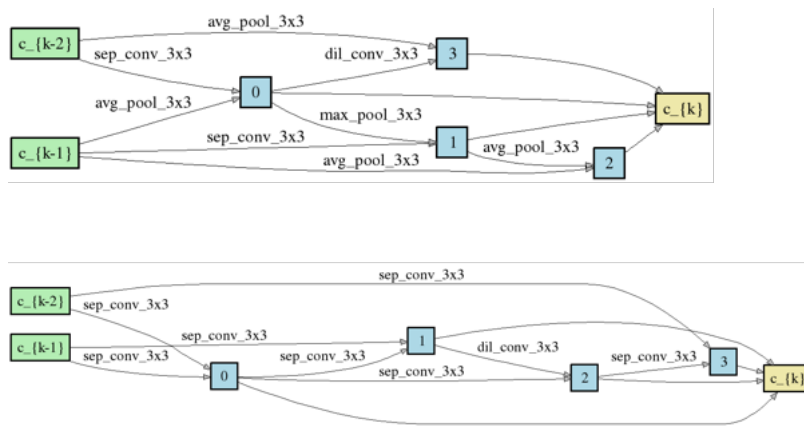


Figure A.25: Normal and Reduction cell in Epoch 30.

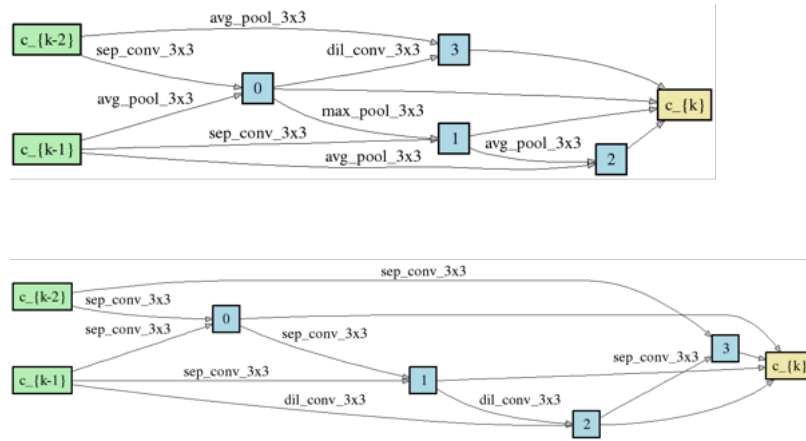


Figure A.26: Normal and Reduction cell in Epoch 31.

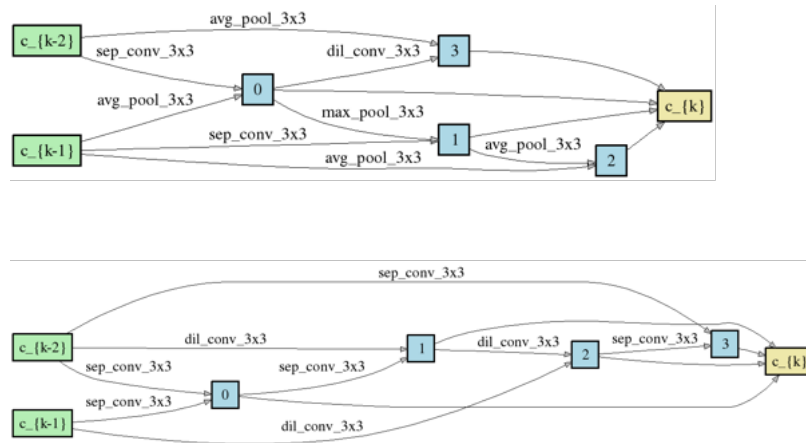


Figure A.27: Normal and Reduction cell in Epoch 32.

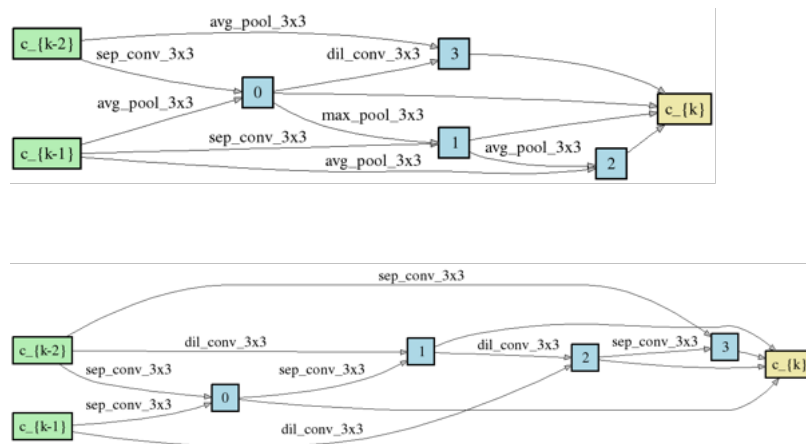


Figure A.28: Normal and Reduction cell in Epoch 33.

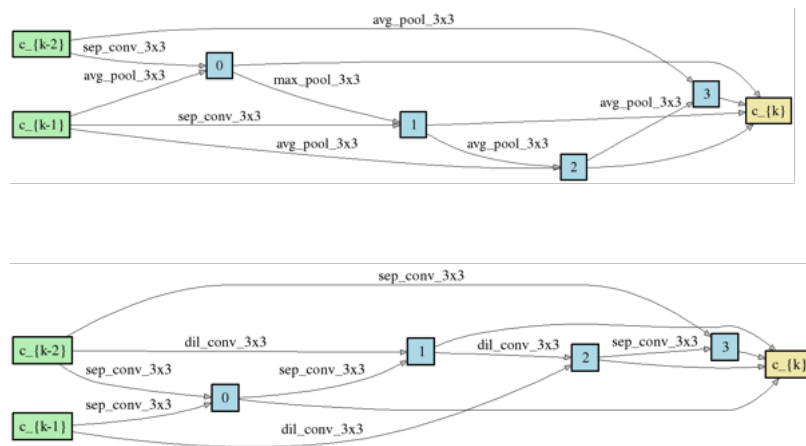


Figure A.29: Normal and Reduction cell in Epoch 34.

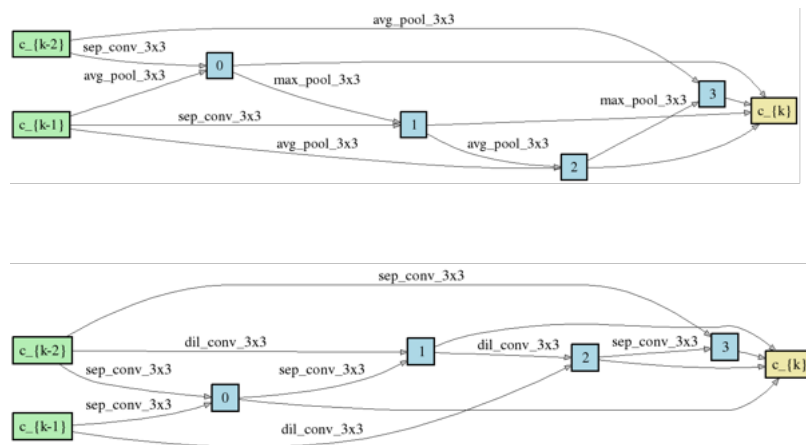


Figure A.30: Normal and Reduction cell in Epoch 35.

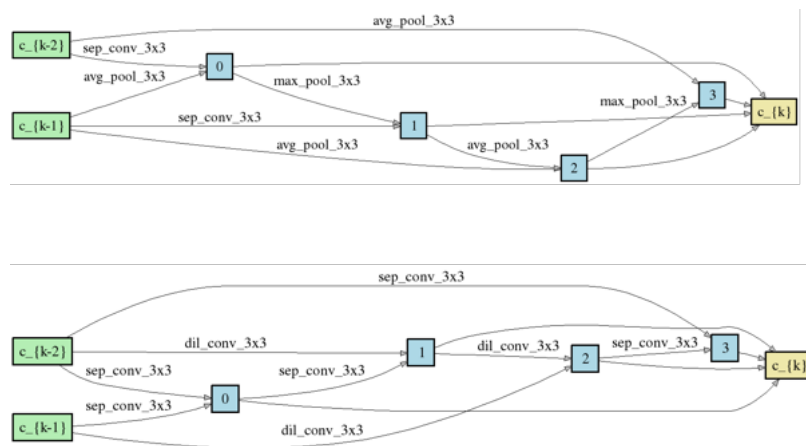


Figure A.31: Normal and Reduction cell in Epoch 36.

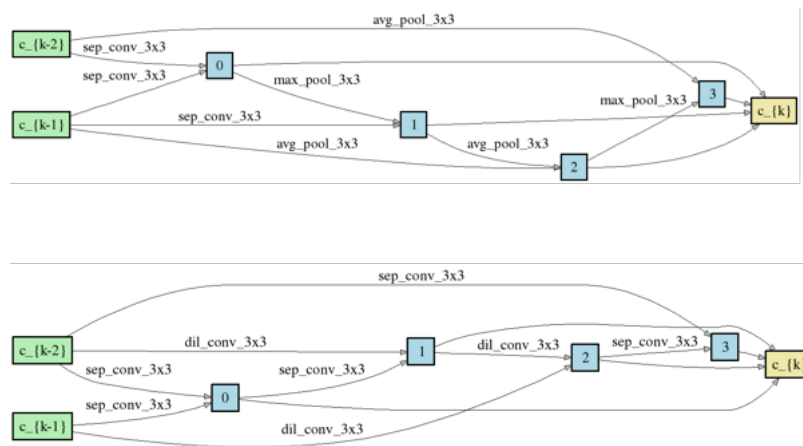


Figure A.32: Normal and Reduction cell in Epoch 37.

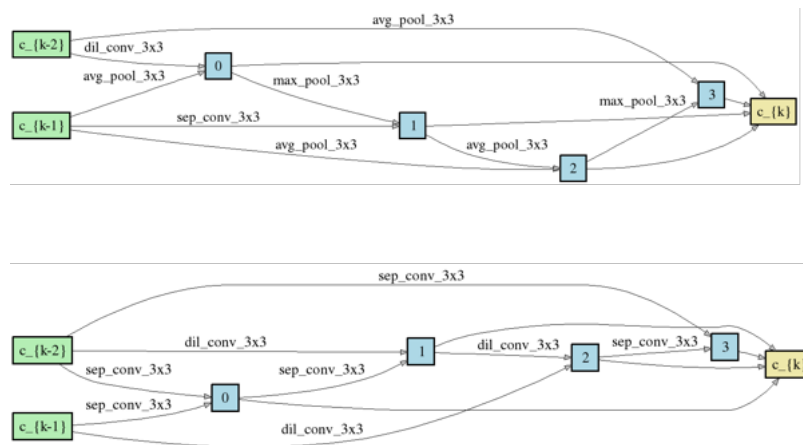


Figure A.33: Normal and Reduction cell in Epoch 38.

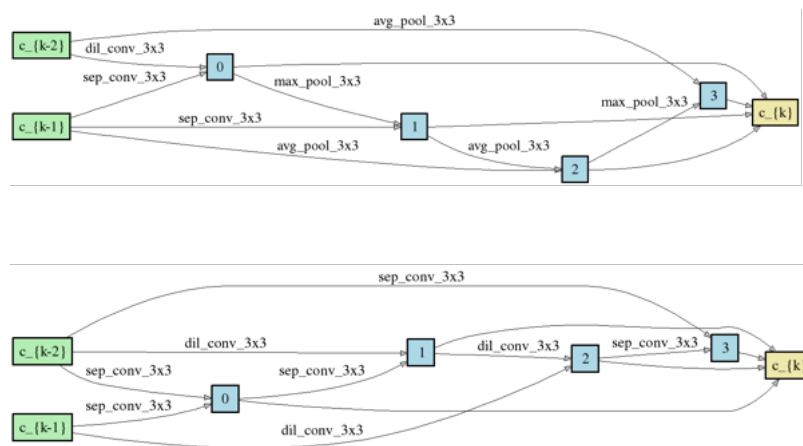


Figure A.34: Normal and Reduction cell in Epoch 39.

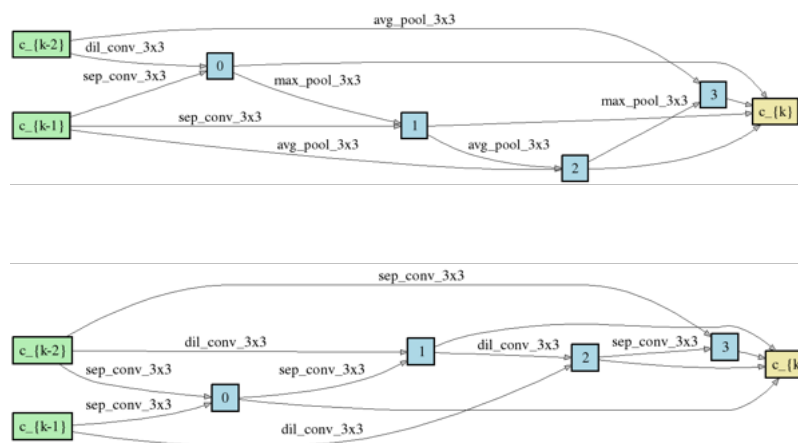


Figure A.35: Normal and Reduction cell in Epoch 40.

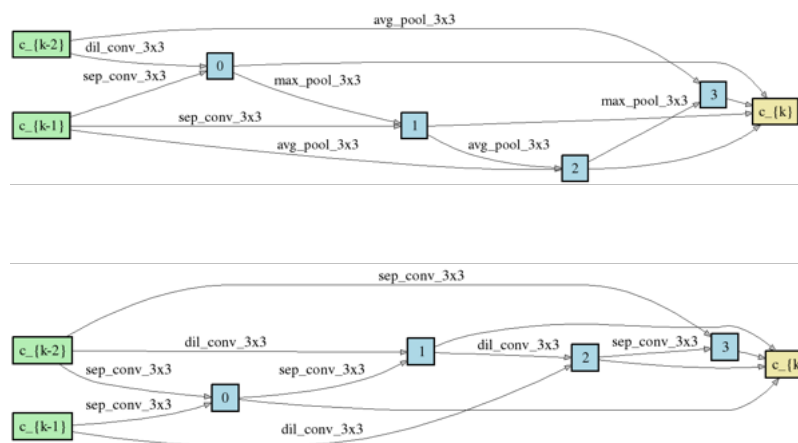


Figure A.36: Normal and Reduction cell in Epoch 41.

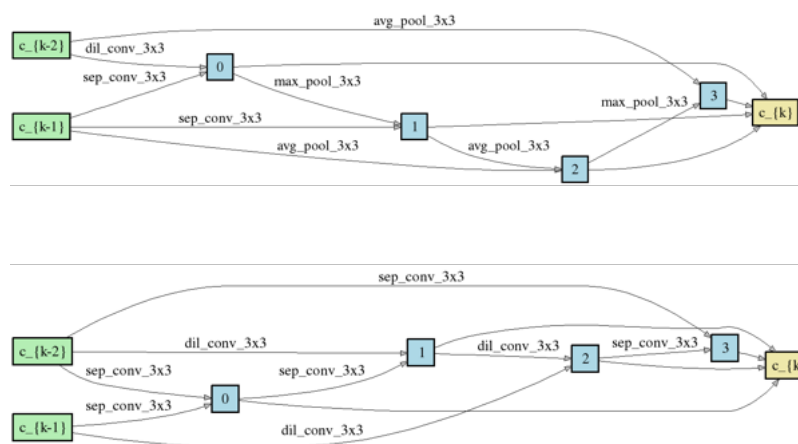


Figure A.37: Normal and Reduction cell in Epoch 42.

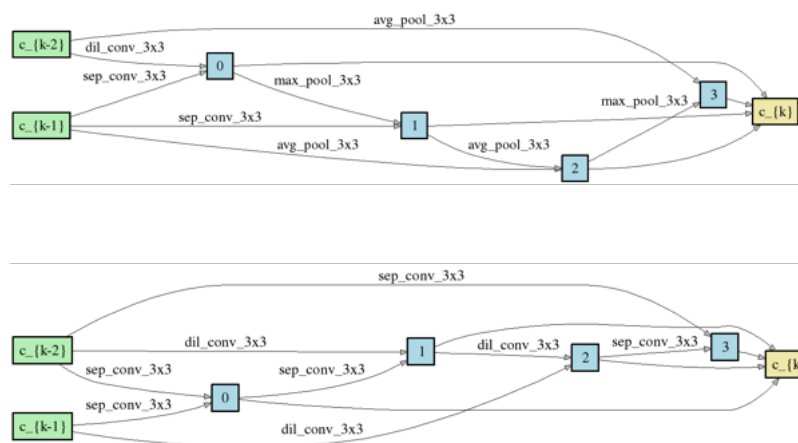


Figure A.38: Normal and Reduction cell in Epoch 43.

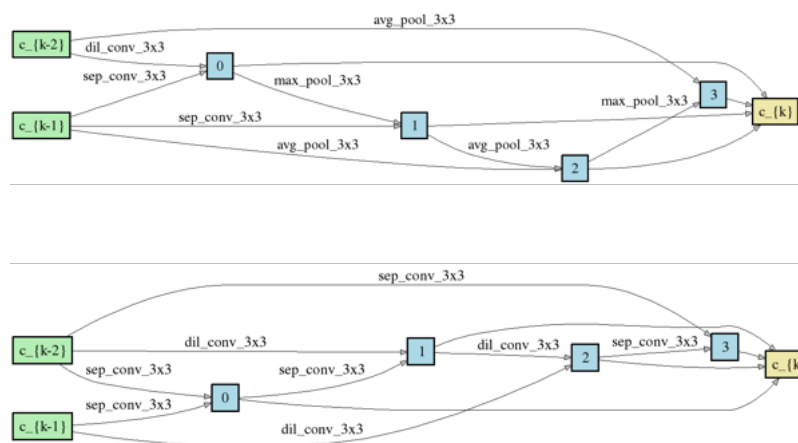


Figure A.39: Normal and Reduction cell in Epoch 44.

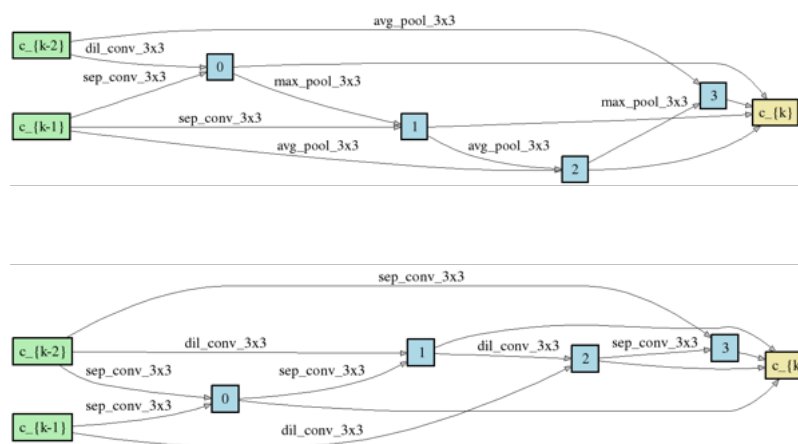


Figure A.40: Normal and Reduction cell in Epoch 45.

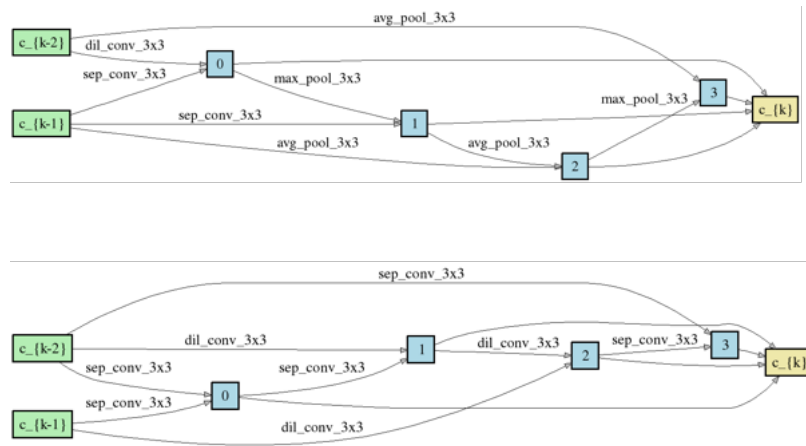


Figure A.41: Normal and Reduction cell in Epoch 46.

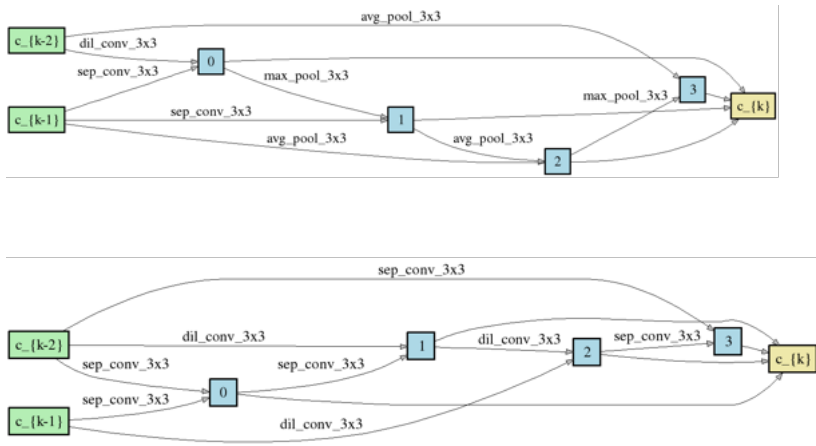


Figure A.42: Normal and Reduction cell in Epoch 47.

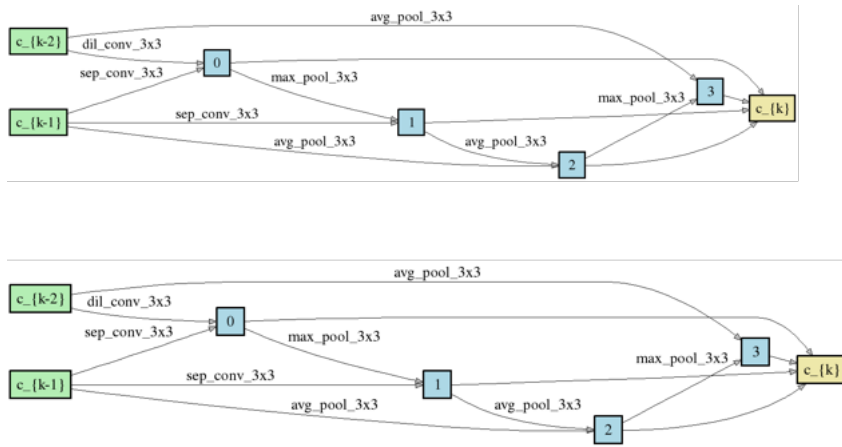


Figure A.43: Normal and Reduction cell in Epoch 48.

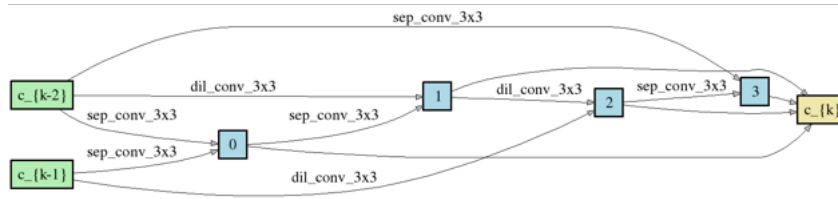
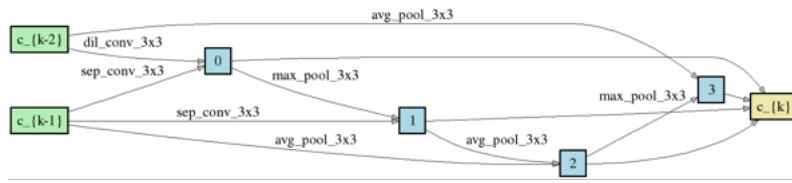


Figure A.44: Normal and Reduction cell in Epoch 49.

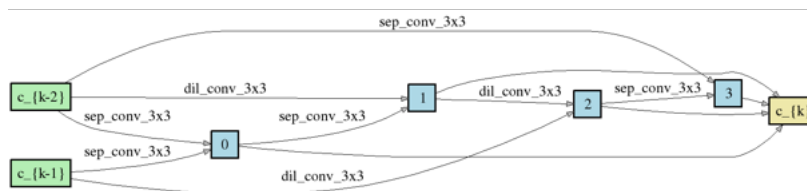
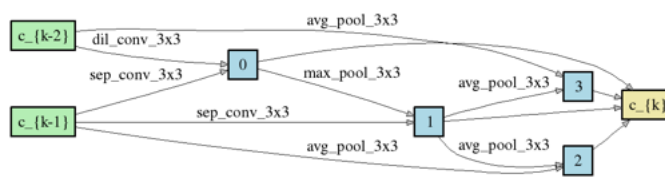


Figure A.45: Normal and Reduction cell in Epoch 50.

Table A.7: Accuracy and Loss for Seed 7

Epoch	trainaccdarts	trainlossdarts	validaccdarts	validlossdarts	trainaccstm	trainlossstm	validaccstm	validlossstm	trainaccstmembed	trainlossstmembed	validaccstmembed	validlossstmembed	trainaccnn	trainlossnn	validaccnn	validlossnn	trainaccnmembed	trainlossnmembed	validaccnmembed	validlossnmembed	trainaccybrid	trainlossybrid	validaccybrid	validlossybrid
1	79.857088	0.432998	82.459162	0.396583	61.84	0.6446	55.67	0.6781	61.09	0.6541	60.2	0.6676	61.64	0.6502	62.89	0.6395	64.37	0.6255	63.45	0.6438	88.97	0.2677	92.65	0.211
2	89.125572	0.288577	91.032423	0.249139	68.56	0.5843	81.92	0.4286	67.14	0.6013	72.84	0.5609	62.83	0.6422	63.37	0.6375	67.85	0.5959	62.91	0.6401	93.25	0.1929	93.4	0.1912
3	91.513367	0.234229	92.567442	0.211397	84.29	0.3767	90.07	0.2658	73.94	0.5388	76.74	0.5094	62.98	0.6403	63.03	0.6396	68.52	0.5899	63.28	0.6449	93.75	0.1808	93.93	0.1775
4	92.759944	0.205604	91.298431	0.240893	89.4	0.2811	90.62	0.2551	77.25	0.4979	82.1	0.4192	63.03	0.6397	63.31	0.6367	69.23	0.5826	61.49	0.6555	94.11	0.1728	93.92	0.1789
5	93.233539	0.195141	93.213684	0.196601	91.17	0.2431	92.88	0.2033	83.46	0.3971	86.62	0.3354	63.06	0.6397	63.06	0.6387	69.5	0.5802	62.47	0.6497	94.26	0.1663	94.21	0.1716
6	93.367064	0.191085	93.251238	0.191462	91.8	0.2298	93.15	0.196	86.03	0.3481	88.14	0.3058	63.11	0.6389	63.54	0.6377	69.64	0.5782	59.99	0.6748	94.5	0.1606	94.23	0.1697
7	93.444779	0.18853	92.565877	0.212354	92.24	0.2194	92.33	0.2197	87.71	0.3155	90.38	0.2588	63.2	0.6387	63.19	0.6388	69.96	0.5751	57.4	0.7493	94.66	0.1563	94.52	0.1621
8	93.577261	0.185977	93.902173	0.17925	92.52	0.2141	93.66	0.1882	89.13	0.2847	91.42	0.2382	63.34	0.637	63.63	0.6326	70.1	0.5731	59.94	0.6644	94.82	0.1526	94.4	0.1645
9	93.64298	0.184177	93.521939	0.186481	92.54	0.2121	93.67	0.187	90.03	0.2671	91.7	0.2311	63.36	0.6366	63.33	0.6366	70.26	0.5713	58.39	0.6766	94.97	0.148	94.26	0.17
10	93.592386	0.183532	93.180824	0.191828	92.69	0.2087	93.42	0.1918	90.49	0.2583	91.86	0.2273	63.36	0.6363	63.26	0.6384	70.29	0.5715	55.65	0.7291	95.06	0.1447	94.47	0.1648
11	93.685228	0.182198	93.439008	0.194949	92.8	0.2052	93.62	0.1908	90.82	0.2503	92.38	0.2155	63.36	0.6361	63.71	0.6329	70.46	0.5697	62.82	0.6436	95.21	0.1411	94.41	0.1652
12	93.662278	0.182396	93.880267	0.178551	92.96	0.2025	93.72	0.1842	91.1	0.2446	92.38	0.2151	63.44	0.6364	63.69	0.6339	70.51	0.5694	56.09	0.7029	95.31	0.1372	94.34	0.1687
13	93.666451	0.181824	93.886526	0.178167	92.96	0.2023	93.86	0.182	91.33	0.239	92.4	0.2135	63.44	0.6358	63.67	0.6316	70.76	0.5675	60.41	0.6571	95.44	0.1336	94.33	0.1684
14	93.706613	0.181363	93.528199	0.182657	93.13	0.1991	93.14	0.202	91.47	0.2354	92.96	0.2021	63.49	0.6357	63.77	0.6335	70.69	0.567	54.41	0.7125	95.56	0.1305	93.7	0.1859
15	93.718087	0.181132	94.035177	0.176198	93.12	0.1983	93.81	0.1824	91.64	0.2322	93.11	0.1988	63.42	0.6354	63.77	0.6323	70.86	0.5661	61.55	0.653	95.63	0.1276	93.73	0.1828
16	93.657584	0.181294	93.942857	0.175756	93.16	0.1969	94.02	0.1762	91.78	0.2292	93.11	0.1998	63.61	0.6354	63.67	0.634	70.77	0.5664	61.95	0.6454	95.79	0.1239	94.19	0.1735
17	93.660713	0.181239	93.944422	0.176104	93.26	0.195	93.93	0.1797	91.96	0.2263	93.29	0.1955	63.53	0.6351	63.54	0.6339	70.83	0.5651	52.2	0.7684	95.87	0.1212	94.22	0.1807
18	93.707134	0.181076	93.636166	0.181253	93.31	0.1944	94.06	0.1752	91.97	0.2256	93.24	0.1951	63.63	0.635	63.64	0.6351	70.9	0.5652	53.04	0.7219	95.97	0.1179	94.32	0.1741
19	93.664886	0.181536	94.150969	0.171554	93.37	0.1923	93.93	0.1796	92.1	0.2219	93.22	0.1937	63.54	0.6351	63.61	0.6354	70.92	0.5648	53.94	0.7143	96.02	0.1159	93.96	0.1829
20	93.748339	0.180346	93.302874	0.188884	93.45	0.1905	94.11	0.1745	92.18	0.2201	93.5	0.1925	63.59	0.6348	63.53	0.6345	70.87	0.5649	58.21	0.6744	96.09	0.1137	94.06	0.1844
21	93.707656	0.180209	92.210679	0.215832	93.46	0.191	94.13	0.1773	92.2	0.2196	93.4	0.1949	63.56	0.6346	63.24	0.636	70.96	0.564	52.34	0.7709	96.2	0.1099	94.1	0.1835
22	93.681055	0.180321	93.334169	0.195041	93.46	0.1898	94.08	0.1742	92.3	0.2176	93.48	0.1917	63.65	0.6344	63.62	0.6323	71.05	0.563	57.13	0.6872	96.25	0.1092	94.14	0.1896
23	93.664364	0.181336	94.04926	0.173562	93.5	0.1887	94.16	0.1733	92.31	0.2171	93.22	0.198	63.77	0.6337	63.74	0.632	71.08	0.5636	52.97	0.7264	96.29	0.1075	93.91	0.1919
24	93.739993	0.180453	94.160357	0.171141	92.58	0.2073	94.16	0.1733	92.39	0.2147	93.68	0.1873	63.71	0.6329	63.7	0.631	71.19	0.5629	54.48	0.7028	96.38	0.1052	94.07	0.1946
25	93.667494	0.180036	93.844278	0.17851	92.58	0.2073	94.16	0.1733	92.43	0.2143	93.38	0.1949	63.77	0.6322	63.87	0.6326	71	0.5629	53.23	0.7034	96.46	0.1026	93.91	0.2035
26	93.628897	0.181744	93.781688	0.176755	92.58	0.2073	94.16	0.1733	92.45	0.2129	93.16	0.1946	63.77	0.6326	63.57	0.6321	71.08	0.5625	55.27	0.6892	96.24	0.1093	93.65	0.2017
27	93.650282	0.180985	93.705015	0.181529	92.58	0.2073	92.84	0.2079	92.51	0.2116	93.26	0.1979	63.77	0.6323	63.95	0.6296	71.08	0.5615	51.95	0.749	96.49	0.1014	93.97	0.2011
28	93.658105	0.18153	93.850537	0.176452	92.72	0.2068	93.96	0.1784	92.55	0.2113	93.4	0.1923	63.77	0.6323	63.89	0.631	71.03	0.5628	56.6	0.6725	96.41	0.1028	94.04	0.2007
29	93.612206	0.182099	94.147839	0.171847	93.17	0.1971	94.15	0.1745	92.52	0.2111	93.78	0.1835	63.78	0.6323	63.93	0.6296	71.09	0.5615	53.41	0.6996	96.57	0.099	93.92	0.2037
30	93.572566	0.182407	93.606436	0.190582	93.42	0.1914	94.15	0.1774	92.61	0.2093	93.46	0.1911	63.82	0.6323	63.66	0.6319	71.24	0.5618	52.65	0.6906	96.58	0.0984	93.75	0.2023
31	93.592387	0.181737	93.742569	0.178083	93.42	0.1914	94.15	0.1774	92.6	0.2092	93.76	0.1822	63.83	0.6321	63.88	0.629	71.19	0.5604	50.15	0.7619	96.67	0.096	93.97	0.2069
32	93.509455	0.1833	94.252767	0.169539	93.42	0.1914	94.15	0.1774	92.64	0.209	93.91	0.1812	63.86	0.6319	63.92	0.6289	71.3	0.5605	53.11	0.7115	96.6	0.0977	93.98	0.2081
33	93.574131	0.182047	94.161922	0.170434	93.42	0.1914	93.12	0.2039	92.76	0.2062	93.69	0.1864	63.78	0.6322	63.83	0.6302	71.3	0.5605	56.68	0.6771	96.58	0.0966	93.83	0.2082
34	93.549095	0.183366	94.238594	0.170864	92.68	0.2094	94.01	0.1788	92.75	0.2064	93.82	0.1827	63.82	0.6318	63.91	0.6302	71.32	0.5613	51.89	0.7172	96.58	0.0966	93.83	0.2082
35	93.508934	0.183756	94.208864	0.173306	93.14	0.1971	94.16	0.175	92.72	0.2056	93.92	0.1807	63.87	0.6314	63.94	0.6291	71.28	0.5605	51.19	0.6984	96.58	0.0966	93.83	0.2082
36	93.480768	0.184407	93.834889	0.176817	93.35	0.193	93.82	0.1856	92.85	0.2047	93.87	0.1809	63.99	0.6314	64.25	0.6285	71.25	0.5606	50.35	0.7181	96.58	0.0966	93.83	0.2082
37	93.516757	0.184486	94.199476	0.17042	93.41	0.1911	94.26	0.1711	92.75	0.2055	93.83	0.1812	63.97	0.6311	63.48	0.6327	71.38	0.5597	52.63	0.6963	96.58	0.0966	93.83	0.2082
38	93.48807	0.184232	94.183828	0.170086	93.53	0.18828	94.21	0.1709	92.82	0.2041	93.98	0.181	63.96	0.6307	64.21	0.6281	71.33	0.5601	50.5	0.741	96.58	0.0966	93.83	0.2082
39	93.516757	0.184194	94.251113	0.171066	93.44	0.1888	94.29	0.1702	92.81	0.2045	93.95	0.1788	64.06	0.6309	63.96	0.6313	71.34	0.5603	57.91	0.6728	96.58	0.0966	93.83	0.2082
40	93.479204	0.18491	92.606561	0.214524	93.58	0.1866	94.18	0.1748	92.87	0.2036	93.86	0.1796	64.14	0.6305	64.05	0.6286	71.29	0.5605	50.56	0.7353	96.58	0.0966	93.83	0.2082

Table A.8: Accuracy and Loss for Seed 8

Epoch	trainaccdarts	trainlossdarts	validaccdarts	validlossdarts	trainaccstm	trainlossstm	validaccstm	validlossstm	trainaccstmembed	trainlossstmembed	validaccstmembed	validlossstmembed	trainaccnn	trainlossnn	validaccnn	validlossnn	trainaccnmembed	trainlossnmembed	validaccnmembed	validlossnmembed	trainaccybrid	trainlossybrid	validaccybrid	validlossybrid
1	77.287262	0.468698	86.602618	0.350734	62.38	0.6415	63.3	0.636	63.13	0.6356	76.45	0.5156	60.51	0.6563	62.87	0.6436	64.64	0.6222	58.19	0.7337	86.73	0.3046	93.15	0.1979
2	88.671797	0.295135	90.667837	0.252849	81.61	0.4132	89.96	0.2711	74.54	0.5116	84.21	0.3869	62.17	0.6457	62.69	0.6418	68.33	0.5915	56.91	0.7343	93.28	0.1927	92.64	0.2098
3	90.843136	0.248033	90.558304	0.250005	89.81	0.2704	92.28	0.2157	84.16	0.3823	86.72	0.3368	62.26	0.645	62.99	0.6415	69.19	0.5829	55.1	0.7456	93.83	0.1801	94.03	0.1764
4	91.723564	0.229763	92.277963	0.215048	91.48	0.2339	91.95	0.225	85.89	0.3504	88.09	0.3103	62.25	0.6446	62.87	0.6409	69.63	0.5786	51.82	0.7774	94.07	0.1724	93.88	0.1791
5	92.233669	0.21628	92.89604	0.202019	92.12	0.2196	92.74	0.2087	87.06	0.3271	89.42	0.279	62.5	0.6436	62.63	0.6403	69.98	0.5753	54.25	0.7364	94.31	0.1654	94.27	0.1678
6	92.684837	0.205788	93.165176	0.198509	92.52	0.211	93.36	0.1916	88.69	0.2961	91.73	0.2354	62.44	0.6432	62.84	0.6391	69.99	0.5739	51.18	0.8131	94.49	0.16	93.78	0.1744
7	92.982137	0.198907	93.356076	0.191213	92.75	0.2052	93.1	0.1959	89.76	0.2732	92.25	0.2227	62.59	0.643	61.75	0.6445	70.23	0.5718	52.31	0.8086	94.66	0.1554	94.37	0.1662
8	93.124007	0.194097	93.306004	0.194116	92.93	0.2009	93.64	0.1862	90.32	0.2614	91.74	0.2286	62.56	0.643	62.96	0.6378	70.44	0.5702	51.08	0.8855	94.81	0.151	94.34	0.1653
9	93.303952	0.191306	92.963324	0.198313	93.05	0.1977	93.94	0.1798	90.43	0.257	92.49	0.2129	62.52	0.6422	62.9	0.6398	70.44	0.5686	51.37	0.8275	94.92	0.1477	94.44	0.1651
10	93.47138	0.189515	93.751957	0.183264	93.13	0.1955	93.98	0.1764	90.87	0.2485	92.54	0.2116	62.57	0.642	61.86	0.6443	70.63	0.5674	52.32	0.7973	95.09	0.143	94.27	0.1681
11	93.472423	0.186859	93.845842	0.179066	93.22	0.1944	93.96	0.1774	91.17	0.2424	92.93	0.2048	62.74	0.6407	63.06	0.6378	70.65	0.5671	51.54	0.8391	95.24	0.1393	94.42	0.164
12	93.4912	0.186382	93.70658	0.184754	93.3	0.1917	94.19	0.1724	91.17	0.2424	92.93	0.2048	62.8	0.64	62.83	0.6382	70.83	0.5652	51.98	0.8081	95.34	0.1359	94.39	0.1648
13	93.621073	0.184027	93.894349	0.178959	93.38	0.1897	94.06	0.1755	91.17	0.2424	92.93	0.2048	62.94	0.6399	62.92	0.6373	70.83	0.5651	51.14	0.8457	95.49	0.1327	94.31	0.1663
14	93.573609	0.183417	93.867749	0.178083	93.46	0.1884	94.06	0.1736	91.17	0.2424	91.21	0.2447	63	0.64	63.3	0.6368	70.78	0.5649	51.23	0.7657	95.64	0.1283	94.25	0.168
15	93.601253	0.182592	94.025788	0.17804	93.42	0.1879	94.05	0.1734	91.17	0.2424	90.45	0.2574	63.05	0.6394	63.28	0.6369	70.98	0.5639	50.7	0.8244	95.74	0.1247	94.09	0.1733
16	93.607512	0.183645	93.398324	0.193946	93.18	0.194	94.31	0.1703	91.17	0.2424	92.54	0.2141	62.98	0.6394	63.22	0.636	70.92	0.5638	52.62	0.7712	95.86	0.1212	94.15	0.1767
17	93.678969	0.182005	93.935033	0.177541	93.58	0.1857	94.05	0.1737	90.72	0.2516	93.04	0.2011	62.97	0.6389	62.49	0.6391	70.97	0.5633	53.04	0.786	95.99	0.1177	94.17	0.1789
18	93.659149	0.181747	92.983666	0.207564	93.57	0.1851	94.29	0.1681	91.29	0.2398	92.16	0.2242	62.93	0.6394	63.04	0.6367	71.02	0.5626	52.01	0.7686	96.08	0.1154	94.03	0.1881
19	93.683141	0.181188	94.10872	0.173099	93.59	0.1849	93.97	0.1781	91.51	0.2338	93.2	0.1972	63.08	0.6395	63.19	0.6358	71.09	0.562	51.77	0.7914	96.2	0.1123	94.23	0.1784
20	93.680012	0.180932	94.025789	0.172937	93.63	0.1837	94.13	0.1762	91.73	0.2292	93.28	0.1969	63.01	0.6389	63.02	0.6371	71.15	0.5621	53.67	0.72	96.3	0.1092	93.87	0.1877
21	93.606469	0.183077	93.046255	0.206639	93.35	0.1894	94.01	0.1754	91.91	0.2255	93.27	0.1935	62.96	0.6391	62.96	0.6379	71.07	0.562	51.86	0.7993	96.38	0.1068	93.94	0.1971
22	93.666451	0.181488	94.085249	0.171157	93.5	0.1858	94.19	0.1713	91.84	0.2259	93.45	0.1905	63.01	0.6389	63.1	0.6379	71.18	0.5606	53.45	0.7447	96.42	0.105	94.06	0.1884
23	93.69253	0.181737	94.152533	0.171804	93.62	0.1831	93.97	0.1756	91.87	0.2261	93.6	0.1869	63.12	0.6388	62.96	0.6389	71.14	0.5614	51.12	0.8201	96.49	0.1031	93.9	0.1917
24	93.645066	0.181519	93.579835	0.18648	93.62	0.1829	94.39	0.1666	92.1	0.2217	93.35	0.1917	63.08	0.639	63.25	0.6355	71.29	0.5601	53.24	0.7854	96.6	0.0997	93.71	0.2077
25	93.620552	0.181308	94.086814	0.170836	93.68	0.1821	94.22	0.1704	92.19	0.2197	93.41	0.1944	63.13	0.6387	63.23	0.636	71.17	0.5606	51.52	0.7866	96.6	0.0996	93.84	0.198
26	93.693573	0.18125	94.085249	0.171601	93.71	0.1824	93.36	0.1902	92.22	0.2172	93.77	0.1843	63.1	0.6387	62.72	0.6393	71.26	0.5593	53.48	0.7684	96.71	0.0972	93.89	0.2029
27	93.636721	0.182073	94.119674	0.172214	93.72	0.1807	94.15	0.1714	92.3	0.2166	93.67	0.1847	63.09	0.6386	63.06	0.6366	71.27	0.5601	53.86	0.7412	96.74	0.0951	93.95	0.2016
28	93.694095	0.180939	93.751957	0.179908	93.7	0.1809	94.43	0.1653	92.29	0.2161	93.6	0.1865	63.18	0.6383	62.91	0.6397	71.34	0.5591	50.38	0.8269	96.82	0.0935	93.76	0.2032
29	93.684706	0.181302	94.105591	0.172901	93.76	0.1804	94.43	0.1641	92.35	0.2152	93.81	0.1817	63.19	0.6386	63.23	0.6354	71.29	0.5596	50.57	0.8366	96.87	0.0913	93.77	0.2159
30	93.636721	0.182049	94.044565	0.171991	93.8	0.1799	94.27	0.1664	92.49	0.2125	93.74	0.1808	63.15	0.6383	62.92	0.6388	71.4	0.5591	53.07	0.7348	96.91	0.0909	93.76	0.2051
31	93.618987	0.181941	93.543846	0.185457	93.78	0.1799	94.4	0.1672	92.41	0.2124	93.8	0.182	63.11	0.6383	63.21	0.6358	71.3	0.5594	52.3	0.7469	96.92	0.0901	93.61	0.2138
32	93.58352	0.182793	94.053954	0.177002	92.51	0.202	94.37	0.1667	92.57	0.2102	93.85	0.1793	63.16	0.6383	62.9	0.6388	71.31	0.559	51.54	0.7748	97.01	0.0877	93.61	0.2221
33	93.589257	0.181845	94.130627	0.170773	93.21	0.19	94.25	0.169	92.5	0.2104	93.92	0.1785	63.16	0.6382	62.98	0.6371	71.38	0.5589	52.31	0.7508	97.03	0.0865	93.73	0.2181
34	93.54075	0.184131	94.083685	0.170783	93.75	0.179	94.02	0.1759	92.67	0.2077	94.06	0.1745	63.16	0.6383	63.13	0.636	71.35	0.5588	51.53	0.835	97.03	0.0869	93.7	0.2109
35	93.606469	0.181815	94.20417	0.16834	93.83	0.178	94.33	0.1658	92.7	0.2073	93.97	0.1764	63.11	0.638	63	0.6371	71.34	0.5585	51.85	0.7635	97.09	0.0843	93.6	0.2269
36	93.555354	0.182901	94.140015	0.171587	93.86	0.1781	94.3	0.1713	92.7	0.2058	93.92	0.1806	63.21	0.638	63.11	0.6359	71.34	0.5584	52.26	0.7506	97	0.088	93.29	0.227
37	93.512585	0.184413	94.211994	0.167123	93.72	0.1805	94.37	0.1654	92.77	0.2053	93.55	0.1849	63.17	0.6377	63.08	0.6378	71.36	0.5587	50.15	0.7913	97.07	0.0854	93.71	0.2158
38	93.525624	0.183614	94.119673	0.174041	93.9	0.1769	94.3	0.167	92.67	0.2069	94.02	0.1755	63.16	0.6383	63.18	0.6363	71.34	0.5586	53.23	0.7348	97.18	0.0823	93.91	0.2228
39	93.484419	0.184368	93.819242	0.176738	93.86	0.1776	94.43	0.1653	92.73	0.2044	94.07	0.1748	63.11	0.6382	63.06	0.6375	71.41	0.5587	51.3	0.8654	97.17	0.0829	93.57	0.2204
40	93.515714	0.184484	93.503162	0.186589	93.89	0.1774	94.34	0.167	92.62	0.2073	94.12	0.174	63.11	0.6381	63.2	0.6365	71.51	0.5578	52.15	0.7566	97.16	0.0822	93.77	0.2337

Table A.9: Accuracy and Loss for Seed 9

Epoch	trainaccdarts	trainlossdarts	validaccdarts	validlossdarts	trainaccstm	trainlossstm	validaccstm	validlossstm	trainaccstmembed	trainlossstmembed	validaccstmembed	validlossstmembed	trainaccnn	trainlossnn	validaccnn	validlossnn	trainaccnmembed	trainlossnmembed	validaccnmembed	validlossnmembed	trainaccybrid	trainlossybrid	validaccybrid	validlossybrid
1	79.229627	0.443306	87.391251	0.32724	67.57	0.5888	83.53	0.3948	69.25	0.5835	79.27	0.502	62.05	0.647	63.19	0.6371	64.61	0.6239	59.47	0.6986	89	0.2682	92.32	0.2229
2	88.697876	0.298189	90.33298	0.263447	85.18	0.3598	89.87	0.2691	82.29	0.42	85.36	0.3545	63.24	0.6387	63	0.6393	67.72	0.5971	57.18	0.7375	93.4	0.19	93.78	0.1832
3	90.796715	0.251103	91.179509	0.241918	89.38	0.2795	92.06	0.2247	84.82	0.3719	87.74	0.3154	63.44	0.6373	63.43	0.6365	68.79	0.588	59.08	0.7064	93.9	0.1781	94.04	0.1743
4	91.698006	0.229128	91.107531	0.242301	90.83	0.2499	91.81	0.2287	86.4	0.3426	89.31	0.2841	63.44	0.6372	63.88	0.6349	69.49	0.5808	56.27	0.7361	94.14	0.171	94.27	0.1699
5	92.274353	0.215855	92.842838	0.20314	91.67	0.2317	92.99	0.2027	87.81	0.3151	90.9	0.2494	63.53	0.6364	63.65	0.6338	69.77	0.5767	58.38	0.6957	94.33	0.1656	94.11	0.1729
6	92.795933	0.203728	91.744384	0.228584	92.04	0.2225	93.25	0.1931	88.96	0.2904	91	0.2479	63.46	0.6367	63.41	0.6387	70.1	0.5741	55.57	0.7287	94.48	0.1606	94.31	0.1699
7	93.094799	0.196957	93.3639	0.189392	92.32	0.2165	93.34	0.1968	89.61	0.2763	91.63	0.2311	63.56	0.6359	63.46	0.6344	70.29	0.5724	52.08	0.7873	94.65	0.1564	94.25	0.1687
8	93.271093	0.193419	93.443702	0.188563	92.44	0.2137	93.5	0.1915	90.02	0.2674	91.59	0.2378	63.61	0.6361	63.22	0.6367	70.33	0.5709	55.38	0.7448	94.83	0.152	94.34	0.171
9	93.317514	0.190784	93.701885	0.18234	92.64	0.2095	93.39	0.1924	90.44	0.26	92.28	0.2213	63.62	0.6358	63.73	0.6344	70.54	0.5698	52.9	0.7603	94.96	0.1477	94.33	0.1679
10	93.415049	0.189658	93.262191	0.193359	92.7	0.2072	93.71	0.1878	90.63	0.255	92.51	0.2179	63.65	0.6356	63.63	0.6362	70.61	0.5674	53.4	0.7297	95.11	0.1432	94.09	0.1705
11	93.436434	0.187653	92.989924	0.200032	92.91	0.2035	93.86	0.1811	90.86	0.2492	92.45	0.2134	63.64	0.6353	63.48	0.6354	70.77	0.5669	53.48	0.7431	95.25	0.1395	94.29	0.1704
12	93.487027	0.186814	93.841148	0.182084	92.97	0.2018	93.82	0.1815	91.04	0.2461	92.64	0.2109	63.79	0.6347	63.97	0.6335	70.84	0.5661	51.49	0.8143	95.4	0.1356	94.06	0.1746
13	93.519365	0.1855	92.173125	0.223379	93.04	0.2001	94	0.1787	91.18	0.2422	92.59	0.2146	63.81	0.6341	63	0.6409	70.84	0.5659	57.11	0.7236	95.47	0.1326	94.01	0.1797
14	93.500067	0.185409	93.562623	0.183145	93.08	0.1992	93.77	0.1846	91.29	0.2396	92.39	0.2212	63.74	0.6339	63.86	0.6342	70.89	0.5652	57.83	0.6813	95.61	0.1288	94	0.1777
15	93.502153	0.184149	93.811418	0.181465	93.15	0.1973	93.91	0.1789	91.37	0.1992	93.14	0.1992	63.85	0.6337	63.7	0.6343	71	0.5643	53.24	0.8233	95.73	0.1252	93.95	0.185
16	93.542836	0.184468	93.406147	0.191241	93.17	0.1962	93.93	0.1779	91.52	0.2348	93.32	0.1959	63.83	0.634	64.06	0.6333	71.06	0.5633	55.5	0.7232	95.81	0.1223	93.82	0.1868
17	93.587692	0.183759	93.928774	0.176739	93.19	0.1964	93.85	0.1847	91.65	0.2317	93.4	0.1925	63.89	0.6334	62.99	0.6375	70.95	0.5638	56.95	0.6942	95.9	0.1204	94.17	0.1822
18	93.58039	0.183932	93.906868	0.178311	93.18	0.1965	93.97	0.178	91.8	0.2288	93.32	0.1966	63.74	0.6335	63.85	0.6334	71.07	0.5636	51.73	0.821	95.97	0.1181	94.07	0.1831
19	93.569959	0.183616	93.969458	0.17561	93.3	0.1936	94.13	0.1741	91.86	0.227	93.59	0.1895	63.82	0.6333	63.92	0.6328	71.11	0.5634	52.45	0.8647	96.06	0.1159	94.02	0.1866
20	93.589257	0.183198	93.892785	0.182432	93.17	0.1963	93.92	0.1779	91.98	0.2257	93.51	0.1898	63.84	0.6332	63.72	0.6322	71.07	0.5628	51.41	0.8843	96.15	0.1128	94.05	0.1815
21	93.554833	0.18363	94.043001	0.173656	93.32	0.1917	94.04	0.1786	92.02	0.2242	93.61	0.1894	63.8	0.6334	64.11	0.6312	71.08	0.5627	52.98	0.7374	96.18	0.1114	93.7	0.1944
22	93.523538	0.183489	92.750518	0.210519	93.34	0.1919	94.12	0.1767	92.06	0.2226	93.06	0.2066	64	0.6332	63.77	0.6343	71.1	0.5621	53	0.731	96.1	0.1132	93.93	0.1846
23	93.591343	0.183306	94.032048	0.175847	93.38	0.1901	93.86	0.1808	92.1	0.222	93.48	0.1889	63.86	0.6332	63.96	0.634	71.23	0.562	53.03	0.7263	96.15	0.1128	94.02	0.1905
24	93.488592	0.184362	94.005447	0.173318	93.41	0.1914	93.98	0.176	92.17	0.2185	93.68	0.1841	63.9	0.6332	64.09	0.6332	71.18	0.563	54.1	0.7342	96.22	0.1108	94.06	0.1961
25	93.503196	0.183768	94.0164	0.175689	93.42	0.1906	93.94	0.179	92.29	0.2169	93.68	0.1865	63.9	0.6334	64.07	0.6327	71.28	0.561	53.55	0.7826	96.35	0.1064	94.04	0.1912
26	93.500067	0.183796	93.770734	0.179341	93.49	0.1891	93.85	0.1807	92.21	0.2198	93.64	0.1848	63.9	0.6328	62.99	0.6416	71.26	0.5614	53.47	0.7701	96.46	0.1036	93.53	0.2062
27	93.472944	0.184522	93.853666	0.176812	93.5	0.1883	94.16	0.1727	92.38	0.2146	93.78	0.1824	63.88	0.633	63.94	0.6323	71.37	0.5596	52.08	0.7311	96.42	0.1053	94.05	0.1954
28	93.529275	0.184065	93.371723	0.187311	93.55	0.1874	94.16	0.1735	92.35	0.2161	93.72	0.1878	63.95	0.6326	63.59	0.6348	71.45	0.5594	52.33	0.7985	96.55	0.1008	93.69	0.2019
29	93.512585	0.184983	93.938163	0.177341	93.46	0.1892	94.3	0.1731	92.44	0.2129	93.78	0.1846	63.9	0.6329	63.65	0.6334	71.44	0.5593	52.24	0.7529	96.5	0.1026	93.92	0.2047
30	93.450516	0.185307	94.122803	0.171759	93.5	0.1881	94.16	0.1737	92.5	0.2124	93.93	0.1791	63.9	0.6329	63.83	0.634	71.41	0.5592	51.2	0.9064	96.62	0.0988	93.85	0.2015
31	93.428089	0.185929	93.772299	0.180778	93.53	0.1867	94.25	0.1713	92.56	0.2101	93.89	0.1806	63.95	0.6323	63.98	0.6315	71.45	0.5594	55.5	0.7199	96.57	0.1	93.85	0.2003
32	93.469815	0.185852	93.992929	0.173117	93.6	0.1866	94.31	0.1696	92.61	0.2096	94.03	0.176	64.01	0.6326	64.07	0.6338	71.52	0.5591	50.29	0.8299	96.59	0.0986	93.74	0.1998
33	93.497459	0.185305	93.823936	0.179986	93.61	0.1849	94.15	0.1725	92.7	0.2075	93.73	0.183	63.93	0.6325	64.09	0.6322	71.34	0.5599	50.43	0.8268	96.63	0.0977	93.72	0.2045
34	93.467207	0.185575	94.017965	0.176261	93.55	0.1863	94.16	0.1772	92.68	0.2075	93.7	0.1886	63.97	0.6323	64.09	0.632	71.33	0.5591	50.81	0.7827	96.64	0.0972	93.76	0.2084
35	93.465642	0.185657	93.653378	0.182188	93.65	0.1838	94.32	0.1711	92.69	0.2069	93.7	0.1853	63.94	0.6325	63.52	0.636	71.52	0.5589	52.49	0.7419	96.64	0.0972	93.76	0.2084
36	93.434348	0.185823	94.088378	0.17241	93.65	0.1837	93.87	0.1781	92.74	0.2059	93.97	0.1778	63.96	0.6326	63.49	0.6355	71.4	0.5592	51.41	0.7602	96.64	0.0972	93.76	0.2084
37	93.457819	0.186059	93.850536	0.17532	93.63	0.1845	94.25	0.1703	92.79	0.2045	94.11	0.1757	63.81	0.6327	64.24	0.6302	71.44	0.5585	52.05	0.8522	96.64	0.0972	93.76	0.2084
38	93.449473	0.187431	93.870878	0.179082	93.66	0.1838	94.31	0.1691	92.81	0.2042	93.93	0.1804	64.09	0.6309	64.14	0.6293	71.46	0.5583	51.31	0.7879	96.64	0.0972	93.04	0.1996
39	93.361326	0.187609	94.086814	0.172516	93.67	0.1836	94.31	0.1694	92.88	0.203	94.11	0.1772	64.15	0.6306	64.37	0.6287	71.65	0.5582	50.23	0.8727	96.64	0.0972	93.78	0.1839
40	93.395229	0.187452	94.086814	0.170581	93.72	0.1826	93.85	0.1788	92.95	0.2013	94.16	0.1733	64.06	0.6309	64.37	0.6288	71.45	0.5585	52.46	0.7743	96.64	0.0972	93.35	0.1922

Table A.10: Accuracy and Loss for Seed 10

Epoch	trainaccdarts	trainlossdarts	validaccdarts	validlossdarts	trainaccstm	trainlossstm	validaccstm	validlossstm	trainaccstmembed	trainlossstmembed	validaccstmembed	validlossstmembed	trainaccnn	trainlossnn	validaccnn	validlossnn	trainaccnmembed	trainlossnmembed	validaccnmembed	validlossnmembed	trainaccybrid	trainlossybrid	validaccybrid	validlossybrid
1	78.983963	0.445655	85.573012	0.368654	61.05	0.6554	64.18	0.6367	59.06	0.6592	59.68	0.6621	59.55	0.6634	60.31	0.657	64.03	0.6277	59.11	0.6688	89.46	0.2619	93.61	0.1875
2	88.987353	0.291626	91.032423	0.245106	75.52	0.5064	83.55	0.3963	67.44	0.592	84.68	0.38	61.74	0.6499	62.81	0.6434	67.79	0.5966	60.38	0.6942	93.49	0.1884	93.96	0.1787
3	91.217631	0.241718	91.758466	0.231211	81.19	0.4193	67.55	0.5996	84.05	0.3843	86.66	0.328	62.45	0.644	62.86	0.6414	68.6	0.589	63.08	0.6387	93.91	0.1773	94.14	0.1728
4	91.921765	0.224546	92.484511	0.209213	82.79	0.388	90.22	0.2603	86.07	0.3448	88.21	0.3086	62.68	0.642	62.81	0.64	69.01	0.5846	63.58	0.6394	94.15	0.1703	93.97	0.1803
5	92.448039	0.212674	92.83345	0.202568	90.1	0.2633	92.2	0.2232	87.5	0.3166	90.22	0.2606	62.91	0.64	63.44	0.6357	69.55	0.5797	61.66	0.6492	94.36	0.1647	94.18	0.1696
6	92.740124	0.206488	92.119924	0.217502	91.52	0.2338	92.84	0.2046	89.07	0.2879	91.85	0.2295	63.09	0.6392	63.22	0.6398	69.66	0.5781	60.51	0.6752	94.54	0.1601	94.13	0.1731
7	92.872084	0.201154	93.302874	0.191272	92.04	0.2217	93.37	0.1937	90.01	0.2672	92.29	0.2164	63.04	0.6389	63.48	0.6365	69.91	0.5754	61.08	0.6593	94.71	0.155	93.99	0.1754
8	92.999349	0.198758	92.245104	0.217804	92.5	0.2123	93.45	0.1905	90.52	0.2554	92.64	0.2124	62.99	0.6389	63.51	0.6351	70.06	0.5738	62.06	0.6462	94.87	0.151	94.26	0.1697
9	93.174079	0.19512	92.939852	0.20298	92.71	0.2073	93.6	0.1863	90.93	0.2469	93.03	0.2042	63.19	0.6383	63.49	0.6362	70.14	0.5737	62.1	0.6499	95.01	0.1471	94.31	0.1668
10	93.166255	0.194514	93.637731	0.185088	92.9	0.2024	93.69	0.1855	91.15	0.2413	92.93	0.2087	63.13	0.6383	63.45	0.6363	70.21	0.5726	62.63	0.6431	95.17	0.1428	94.14	0.1694
11	93.179295	0.193286	93.79577	0.183082	92.99	0.2001	93.63	0.1837	91.44	0.2358	93.08	0.1984	63.22	0.638	63.59	0.6348	70.21	0.5713	62.64	0.643	95.29	0.1389	93.91	0.1782
12	93.27005	0.191878	93.409277	0.190483	92.99	0.2001	93.63	0.1837	91.56	0.2344	93.37	0.1945	63.28	0.6375	63.32	0.6354	70.54	0.5694	60.54	0.6578	95.38	0.1353	94.18	0.1717
13	93.300301	0.191137	93.069727	0.197159	92.99	0.2001	93.63	0.1837	91.67	0.2302	93.44	0.1922	63.35	0.637	63.66	0.6337	70.54	0.5688	62.8	0.6439	95.55	0.1313	94.22	0.1708
14	93.258054	0.191268	93.809853	0.188743	92.99	0.2001	93.36	0.2126	91.79	0.2283	93.6	0.1892	63.36	0.637	63.44	0.6379	70.42	0.5688	63.52	0.6401	95.66	0.1279	94.16	0.1781
15	93.367585	0.189385	93.04782	0.19631	92.41	0.2134	93.49	0.1881	91.94	0.2252	93.55	0.1901	63.3	0.6366	63.74	0.635	70.45	0.5688	62.94	0.6439	95.74	0.1249	94.2	0.1734
16	93.337334	0.189849	93.285662	0.192345	93.04	0.1998	93.23	0.1936	92.07	0.2221	93.73	0.1851	63.4	0.6366	63.27	0.6347	70.68	0.567	62.76	0.6421	95.82	0.1222	93.86	0.1865
17	93.303952	0.189906	92.67228	0.204503	93.2	0.1954	93.86	0.1791	92.14	0.2209	93.59	0.1869	63.39	0.6364	63.69	0.6336	70.64	0.5665	62.3	0.6461	95.96	0.1185	94.26	0.1774
18	93.334726	0.188254	93.656508	0.184786	93.33	0.1931	93.99	0.1786	92.14	0.2207	93.54	0.1905	63.45	0.636	63.74	0.6348	70.75	0.5664	59.26	0.6662	96.03	0.1169	93.86	0.1906
19	93.263269	0.189704	93.290357	0.192046	93.38	0.1914	94.02	0.1797	92.22	0.2186	93.61	0.1862	63.41	0.636	63.61	0.6355	70.66	0.5671	63.08	0.6441	96.18	0.1136	94.1	0.1859
20	93.323773	0.189758	93.356076	0.187434	93.4	0.1904	94.1	0.1737	92.26	0.2178	93.76	0.186	63.45	0.6359	63.48	0.636	70.72	0.5649	62.5	0.6435	96.23	0.1109	94	0.1917
21	93.350895	0.188391	92.731741	0.20802	93.48	0.1883	93.74	0.1827	92.32	0.2158	93.89	0.1808	63.42	0.6358	63.89	0.6333	70.87	0.5643	62.89	0.6404	96.32	0.1092	94	0.1918
22	93.352459	0.188398	93.903738	0.176525	93.52	0.1869	93.97	0.178	92.3	0.216	93.59	0.1858	63.45	0.6359	63.76	0.6326	70.87	0.5648	63.62	0.6411	96.4	0.106	93.84	0.1967
23	93.317513	0.188262	93.608001	0.187275	93.58	0.1858	94.25	0.1716	89.54	0.268	93.17	0.1987	63.43	0.6357	63.37	0.6365	70.76	0.5652	56.56	0.7028	96.41	0.1057	93.69	0.1977
24	93.295607	0.188684	93.963198	0.17614	93.56	0.1856	94.22	0.1723	91.33	0.2365	93.18	0.1959	63.33	0.6359	63.84	0.6383	70.97	0.564	63.37	0.6425	96.47	0.1041	94.02	0.1966
25	93.357675	0.187943	93.753523	0.182915	93.6	0.185	94.17	0.1704	92.17	0.2194	93.78	0.1836	63.51	0.6357	63.12	0.6372	70.92	0.5642	62.9	0.6429	96.49	0.1023	94.01	0.1977
26	93.293521	0.188805	93.980411	0.175092	93.54	0.186	93.61	0.1896	92.34	0.2158	93.73	0.1872	63.43	0.6353	63.67	0.6351	70.9	0.5647	63.22	0.6439	96.57	0.1003	93.75	0.206
27	93.285176	0.189898	93.86149	0.17697	93.66	0.1836	94.32	0.1699	92.43	0.2136	93.81	0.1838	63.52	0.6352	63.53	0.6354	70.94	0.5632	57.59	0.6719	96.57	0.1	93.94	0.1941
28	93.35924	0.189168	93.944422	0.175131	93.67	0.1829	94.13	0.1765	92.44	0.2129	93.89	0.181	63.56	0.6354	63.77	0.6344	71.04	0.5626	60.17	0.6569	96.65	0.0987	93.86	0.2038
29	93.327424	0.188806	93.052514	0.199541	93.68	0.182	94.26	0.1709	92.46	0.2126	93.9	0.1799	63.49	0.6352	63.78	0.6336	71.07	0.5626	60.35	0.6562	96.66	0.0976	93.83	0.2066
30	93.2471	0.190023	94.104026	0.173485	93.6	0.1845	94.33	0.1688	92.6	0.2106	93.91	0.1802	63.54	0.6352	63.74	0.6354	71.02	0.5632	61.4	0.6522	96.78	0.0956	93.85	0.2049
31	93.2471	0.19004	94.063342	0.173329	93.72	0.1825	94.26	0.1683	92.6	0.2099	93.94	0.1797	63.41	0.6352	63.26	0.6364	71.11	0.5621	61.62	0.6499	96.75	0.0943	93.81	0.2091
32	93.230931	0.189578	93.595483	0.186601	93.72	0.1816	93.87	0.1804	92.65	0.2097	93.98	0.1794	63.47	0.6353	63.79	0.6329	71.1	0.5621	59.68	0.662	96.81	0.0944	93.72	0.2119
33	93.248665	0.190243	93.927209	0.175719	93.69	0.1817	94.39	0.1648	92.63	0.2084	93.94	0.1777	63.53	0.6351	63.57	0.6355	71	0.5627	62.25	0.6494	96.8	0.0941	93.98	0.2121
34	93.280481	0.189962	93.908433	0.17595	93.82	0.1802	94.22	0.1693	92.7	0.2081	93.98	0.1778	63.56	0.6349	63.02	0.6384	71.13	0.5624	61.91	0.6493	96.84	0.0913	93.78	0.2168
35	93.246057	0.189721	94.043001	0.174542	93.81	0.1803	94.38	0.1664	92.71	0.2077	94.03	0.1771	63.5	0.6349	62.47	0.638	71.14	0.5619	60.66	0.6552	96.88	0.0913	93.72	0.2161
36	93.172514	0.192022	93.950681	0.17568	93.77	0.1806	94.19	0.1711	92.78	0.2061	94.05	0.1759	63.49	0.635	63.46	0.6362	71.11	0.5614	61.61	0.653	96.91	0.0912	94.02	0.2082
37	93.249708	0.190662	93.517245	0.188194	93.78	0.1796	94.22	0.1701	92.71	0.2073	93.86	0.1847	63.81	0.6327	64.24	0.6302	71.2	0.5615	60.27	0.657	96.88	0.0909	93.87	0.2104
38	93.204331	0.191052	93.590788	0.183568	93.83	0.1793	94.38	0.1658	92.81	0.2058	94.1	0.1801	64.09	0.6309	64.14	0.6293	71.19	0.5613	60.86	0.6556	96.93	0.09	93.74	0.2104
39	93.188162	0.190624	93.939727	0.176433	93.87	0.1788	94.45	0.1657	92.81	0.2056	93.92	0.1773	64.15	0.6306	64.37	0.6287	71.15	0.5617	61.44	0.6498	96.96	0.0894	93.41	0.2265
40	93.195464	0.19164	93.789511	0.180775	93.85	0.1787	94.41	0.1642	92.83	0.2046	94.16	0.1746	64.06	0.6309	64.37	0.6288	71.13	0.5612	62.52	0.6506	96.98	0.088	93.77	0.2141