STUDIES ON PLANT GROWTH REGULATORS AND ENZYMES PRODUCTION BY SOME BACTERIA

By

F. A. MANSOUR, H. S. ALDESUQUY* and H. A. HAMEDO

Department of Botany, Faculty of Science, Mansoura University, Mansoura, Egypt Present address: Ministry of Education, Hail Teacher College, P. O. Box 1818, Kingdom of Saudi Arabia

دراسات على الأنشطة الهرمونية والانزيية لبعض البكتريا

فتحى عواد منصور و حشمت سليمان الدسوقى و هند عبد الحميد حميدو

يهدف هذا البحث نحو دراسة الأنشطة الهرمونية لأربع وعشرين سلالة بكتيرية تابعة لجنس ستربتوميسيس وقدرتها على إنتاج أنزيمي الألفا أميليز والبروتينيز . وقد أسفرت هذه الدراسة عن النتائج التالية :

الكائنات نشاطا من حيث قدرتها على إنتاج منظمات النمو (الأوكسينات والجبريلينات والمبريلينات والسيتوكينينات) وأنزيمات الألفا أميليز والبروتينيز بليها في ذلك:

S. alboviridis, S. griseoviridis, S. phaeochromogenes, Streptomyces sp. No. 20.

٢ ـ أظهرت الكائنات :

S. albidoflavus, S. albus, S. caesius, S. citreus, S. griseinus, S. scabies, Streptomyces sp. No. 21, S. tetranusemus, S. violaceus and S. viridosporus

نشاطا متوسطا عنذ مقارنتها بالكائنات المذكورة أعلاه من حيث قدرتها على إنتاج منظمات النمو والانزيمات موضوع الدراسة ، في حين كانت بقية الكائنات غير ذي نشاط يذكر من حيث انتاج منظمات النمو أو الانزيمات .

وقد وجد أن S. rimosus, S. olivaceoviridis تعطي أعلى قدر من منظمات النمو والنشاط الأنزيمي (أنزيمي الفا أميليز والبروتينيز بعد فترة تحصين ٨٤ ساعة أما بالنسبة S. rochei فكانت بعد ٦٠ ساعة).

Key Words: α-Amylase, Auxins, Cytokinin, Gibberellins, Streptomyces

ABSTRACT

 Streptomyces sp. No. 21, S. tetranusemus, S. violaceus and S. viridosporus. The remaining seven strains produced comparatively low levels of growth regulators and enzymes.

The maximum production of growth regulating substances and enzymes by S. olivaceoviridis and S. rimosus was attained after 84 hrs, but at 60 hrs for S. rochei.

INTRODUCTION

Various *Streptomyces* species are capable of producing extremely valuable biologically active substances; of which the most important are growth promoting substances, enzymes, antibiotics, pigments, amino acids and vitamins [1].

There is much evidence that bacteria produce indolyl-3-acetic acid (IAA) in culture media [2, 3]. Katuznelson and Cole [4] and Sobieszczanki [5] found gibberellin-like substances in culture liquids of several micro-organisms such as Azotobacter chroococcum [6] and Pseudomonas species [7]. Also, Kampert and Strzelczyk, [8] found that several soil bacteria produced substantial amounts of cytokinin - like substances.

The ability of many *Streptomyces* to produce extracellular enzymes has been demonstrated by many investigators. Amylases and proteases are among the most intensively investigated enzymes [9-18]. However, there is no available report (as the authors are aware) concerning the relationship between the capacity of *Streptomyces* to produce growth-promoting substances and their amylases and proteases activities.

The present investigation was undertaken to study the growth promoters, α-amylase and protease producing potential of some bacterial strains isolated from Egyptian soil.

MATERIALS AND METHODS

Microorganisms and cultural conditions:

Twenty four *Streptomyces* strains, formerly isolated and identified by Mansour [19] were used in this experiment.

Each of the experimental strains was cultured in 250 ml Erlenmyer flasks, each containing 50 ml of starch - casein medium composed of (gl⁻¹ tap water): starch, 10.0; Casein, 0.3; NaCl, 2.0; K_2HPO_4 , 2.0; KNO_3 , 2.0; $MgSO_4$. $7H_2O$, 0.5; $CaCO_3$, 0.2 and $FeSO_4$. $7H_2O$, 0.1. The pH was adjusted to 7-7.5. Dense spore suspensions (5 ml each), prepared from 5-day-old cultures, grown on starch casein agar at 28°C, were used for inoculation. Inoculated flasks (3 replicates) were kept on a gyratory shaker (200 rpm) at 28°C for 4 days. At the end of the incubation period, the whole cultures were filtered through Whatman number 50 filter papers, The obtained filtrates were then filtered through millipore (0.450 μ m) filter and the cell-free filtrates were used for determination of growth regulating activity as well as for α -amylase and proteinase producing potential.

The most active strains (i.e. S. olivaceovirids, S. rimosus and S. rochei) were selected and cultured as above mentioned and left to grow for 96 hrs, meanwhile their growth-regulating production as well as their amylase and proteinase producing-

potential of their culture filtrates were assayed every 12 hrs intervals; so that the suitable time for harvesting and the most active organism(s) can be determined.

Hormone extraction, purification and bioassay

The method of extraction was that originally described by Shindy and Smith [20]. The amount of either acidic or neutral auxins were estimated according to straight growth test of barley coleoptile adopted by Foda and Radwan [21]. Gibberellic acid in extracts was determined by the lettuce hypocotyl bioassay developed and adopted by Frankland and Wareing [22]. Cytokinin content of the cell free extracts was estimated according to the method described by Esashi and Leopold [23].

α-amylase assay:

The reaction mixture composed of: 0.1 ml cell-free filtrate plus 5 ml of 0.5% starch plus 0.1 ml acetate buffer at pH 4.5. Incubation was at 37°C for 30 minutes. The direct reducing value (D.R.V) which was considered to be equivalent to reducing sugars was determined in the filtrates following the procedure of Nelson [24]. One unit of α -amylase was chosen to equal the amount of enzymes which produce 10 μ g glucose from starch hydrolysis.

Proteinase assay

Anson's casein assay method [25], with slight modification [26], was used.

RESULTS

Production of growth regulators

The results presented in Table 1 showed that the experimental organisms possess variable potentialities for production of plant growth regulators.

Total auxins

The total auxin content was measured as µg IAA equivalent ml⁻¹ of culture filtrate. It was obvious from Table 1 that S. alboviridis, S. griseoviridis, S. olivaceoviridis, S. phaeschromogenes, S. rimosus, S. rochei and Streptomyces sp. No. 20 had the ability to produce substantial amounts of auxins. Lesser amounts of IAA-like substances were detected in the culture filtrate of S. albus, S. albidoflavus, S. citreus, S. scabies. Streptomyces sp. No. 21, S. tetranusemus, S. viridosporus and S. violaceus. The remaining other species of Streptomyces seemed to produce very slight amount of auxins.

Gibberellins

The gibberellin content was measured as µg GA₃ equivalent ml⁻¹ of culture filtrate. The data presented in table 1 showed that, 6 species, namely S. albviridis, S. citreus, S. griseoviridis, S. prasinus, S. rimosus and S. tetranusemus

proved to be the most active in production of gibberellins. These were followed by S. albus, S. minoensis, S. olivaceovirids, S. olivaceus, S. rochei and S. sp. which produced detectable amounts of gibberellins. On the other hand, S. albidoflavus, S. atroolivaceus, S. caesius, S. coelicolor, S. griseinus, S. gougerotii, S. matensis and S. viridosporus showed mush lesser capacity for production of gibberellins.

S. olivaceoviridis, S. violaceus and S. viridosporus showed comparatively the highest biomass growth. These are followed by: S. albus, S. alboviridis, S. caesius, S. coelicolor, S. matensis, S. olivaceus, S. phaeochromogenes, S. prasinus, S. rochei, S. rimosus and Streptomyces spp. No. 20 & 21, whereas, the remaining species, namely S. citreus, S. griseinus, S. minoensis, S. scabies and S. tetranusemus produced slight

Table 1

Preliminary screening for growth-regulating substances produced by some thallobacteria grown in starch-casein shaken cultures for 4 days at 28°C. Each value is the mean of duplicate determinations ± standard errors.

Organisms Names	Strain Nos.	Total Auxins Equivalent To IAA µg ml ⁻¹	Gibberellins Equivalent To GA ₃ µg ml ⁻¹	Cytokinins Equivalent To Kinetin µg ml ⁻¹
Streptomyces albidoflavus	1	24.01±1.23	2.76≌0.22	3.93±0.13
Streptomyces alboviridis	2	35.87±1.25	9.74±0.41	8.62±0.29
Streptomyces albus	3	27.79±0.92	5.57±0.99	3.04±0.10
Streptomyces atroolivaceus	4	17.35±0.15	1.21±0.13	2.53±0.22
Streptomyces caesius	5	21.65±0.40	2.56±0.51	2.77±0.09
Streptomyces citreus	6	23.13±0.89	10.08±0.18	6.37±0.21
Streptomyces coelicolor	7	14.36±1.26	4.61±0.85	7.32±0.13
Streptomyces griseinus	8	29.57±0.24	2.03±0.52	2.00±0.07
Streptomyces griseolviridis	9	38.86±0.51	13.26±0.19	2.47±0.08
Streptomyces gougerotii	10	19.67±0.31	3.44±0.12	6.40±0.11
Streptomyces matensis	11	15.75±0.84	2.13±0.39	4.78±0.16
Streptomyces minoensis	12	17.40±0.11	5.81±0.40	6.11±0.11
Streptomyces olivaceoviridis	13	34.19±1.46	5.53±0.16	14.88±0.26
Streptomyces olivaceus	14	15.22±0.25	5.61±0.24	5.68±0.10
Streptomyces phacochromogenes	15	36.57±1.36	6.91±0.46	6.55±0.22
Streptomyces prasinus	16	10.51±0.48	9.56±0.41	3.38±0.06
Streptomyces rimosus	17	39.04±0.61	8.30±0.87	4.70±0.08
Streptomyces rochei	18	32.75±0.23	5.69±0.09	6.73±0.37
Streptomyces scabies	19	28.26±0.67	6.87±0.47	9.16±0.16
Streptomyces sp.	20	30.78±0.79	5.27±0.89	5.55±0.10
Streptomyces sp.	21	22.17±2.29	1.90±0.12	3.94±0.39
Streptomyces tetranusemus	22	24.97±0.49	8.46±1.12	3.95±0.07
Streptomyces viridosporus	23	23.09±1.03	2.92±0.48	8.59±0.15
Streptomyces violaceus	24	27.85±1.31	1.84±0.04	2.74±0.05

Cytokinins

The cytokinin content was measured as µg kinetin equivalent ml⁻¹ of culture filtrate. As can be seen from table 1, only 5 species, namely *S. alboviridis*, *S. coelicolor*, *S. olivaceoviridis*, *S. scabis* and *S. viridosporus* proved to be capable of production of substantial amounts of cytokinin-substances. The remaining other strains seemed, however, to have a slight or negligible cytokinin - producing potential.

Biomass growth

As can be seen from Table 2, all of the employed strains were able to grow in starch - casein medium, though with variable growth magnitudes. Thus, 7 species, namely S. albidoflavus, S. atroolivaceus, S. griseoviridis, S. gougerotii,

biomass weight.

Production of α-amylase

The results presented in table 2 showed that isolates: S. albovirids, S. griseovirids, S. olivaceoviridis and S. violaceus were of comparatively high α-amylase -producing potential. Moderate α-amylase activity were recorded with S. albus, S. albidoflavus, S. rimosus, S. scabies and S. tetanusemus, whereas, S. atroolivaceus, S. griseinus, S. gougerotii, S. olivaceus, S. matensis, S. minoensis, S. phaseochromogenus, S. prasinus, S. rochei and S. sp. were of slight α-amylase production. On the other hand, S. caesius, S. citreus, S. coelicolor and S. viridosporus had very weak amylolytic activity.

Production of proteinase

The results in table 2 indicate that, S. albidoflavus, S. albus, S. atroolivaceus, S. caesius, S. gougerotti, S. olivaceus, S. olivaceoviridis, S. phaeochromogenes, S. prasinus, S. rimosus, S. sp., S. viridosporus and S. violaceus were very active in proteinase production, whereas S. alboviridis, S. coelicolor, S. griseinus and S. rochei resulted in moderate activities towards proteinase production. On the other hand, the remaining four species were of negligible activities.

Effect of different incubation periods on growth regulating substances produced by three isolates of *Streptomyces* species.

Total auxins

Streptomyces olivaceoviridis and Streptomyces rimosus exhibited a progressive increase in auxins production till 84 hrs incubation, followed by a marked reduction at 96 hrs. On the other hand, Streptomyces rochei resulted in high amounts of auxins during the first two periods (24-48 hrs) where upon, auxins production decreased gradually during the subsequent incubation period. Streptomyces olivaceoviridis appeared to be the most active producer for auxins (Table 3).

Gibberellins

The data presented in Table 3 indicates that *Streptomyces olivaceoviridis* isolate No. 13 showed a gradual increase in the gibberellin production till 60 hrs, followed by a slight decrease at 72 hrs. Again gibberellin production increased at 24 hrs, followed by a marked decrease at 96 hrs. Both *S. rimosus* and *S. rochei* showed a progressive increase in gibberellin production during all the incubation periods.

Cytokinins

S. olivaceoviridis and S. rochei showed a progressive increase in cytokinins production till 84 hrs, where upon a marked decrease in case of former strain was observed at 96 hrs. On the other hand, S. rimosus produced a comparatively higher amount of cytokinins at 24 hrs, but a gradual decrease in cytokinin production was manifested during the subsequent incubation period. Nevertheless, S. olivaceoviridis proved, by far, to be more active for production of cytokinins than S. rimosus and S. rochei.

Effect of different incubation periods on biomass weight, α -amylase and proteinase production by three isolates of *Streptomyces* species.

The data given in table 4 show that biomass weight of S. olivaceoviridis increased gradually till 60 hrs followed by a gradual decrease up to 96 hrs. α -amylase production was very high at the first incubation period, where the production decreased gradually till 60 hrs. The production of α -amylase was stable at 60 and 72 hrs. Again a slight increase or decrease in α -amylase production was observed at 84 and 96 hrs respectively. Proteinase production increased gradually till 60 hrs, remained unchanged up to 84 hrs and decreased drastically at 96 hrs.

Biomass weight of *S. rimosus* was increased progressively until 60 hrs, followed by a gradual decrease till 96 hrs. α -

amylase production was high at 24 hrs, then decreased gradually during the subsequent incubation period. On the other hand, proteinase production increased gradually during the first two incubation periods followed by a slight decrease at 60 hrs, where upon a gradual decline was detected during the subsequent incubation period.

Concerning S. rochei, the biomass weight was increased gradually till 72 hrs, then leveled-off during the remaining incubation period. α-Amylase production was very high after 24 hrs then leveled-off till 72 hrs where upon a gradual decline was detected during the subsequent growth period. Proteinase production increased progressively till 84 hrs incubation period followed by a drastic decrease at 96 hrs.

DISCUSSION

This study conducted to determine the growth regulators producing potential of 24 Streptomyces species clearly shows that these organisms exhibit a great deal of variations in their ability to elaborate growth promoters in their culture media. Thus, Streptomyces olivaceoviridis, S. rimosus and S. rochei were superior to the remaining other species in their capacity produce growth-promoting substances to particularly gibberellin-like ones. This may explain the comparatively higher proteinase and α-amylase activities of the former strains. Gibberellic acid has been suggested to be acting at the gene level as a depressor of those genes which code for hydrolytic enzymes [27, 28]. The relatively high protease activity may have an indirect role in auxin synthesis via the production of amino acids especially tryptophan which incorporates in the biosynthesis of IAA and related substances.

El-Sayed et al., [29] found that the metabolite of Streptomyces mutabilis contained tryptophan and anthranilic acid, and that of S. atroolivaceus contained tryptamine and indole acetaldehyde. Also, IAA synthesis follows different pathways in the two species: via indole acetamide in S. mutabilis, and via tryptamine, tryptophan and indole acetaldehyde in S. atroolivaceus.

Tryptophan -----> Indole acetamide -----> Indole acetic acid

$$H$$
 O $R-CH_2$ - $C-COOH$ $R-CH_2$ - $C-NH_2$ $R-CH_2$ COOH NH_2 Tryptophan $T-C-C-C-C$ Tryptophan $T-C-C-C-C$

Fig. 1: Pathway of IAA synthesis according to El-Sayed et al., [29].

Gordon et al., [30] found that free cytokinin is released by Frankia isolate HFPArI3 in defined culture medium. El-Shanshoury [2] reported that S. atroolivaceus produced activity, exogenous IAA, kinetin and gibberellic acid along with other growth regulating substances. Furthermore, plant

Table 2
Preliminary screening for α-amylase and proteinase-producing potential of some isolates of thallobacteria grown in starch-casein shaken cultures for 4 days at 28°C. Each value is the mean of duplicate determinations ± standard errors

Organisms Names	Strain Nos.	Biomass Weight g.l ⁻¹	α-Amylase Unit ml ⁻¹	Proteinase Unit ml ⁻¹
Streptomyces albidoflavus	1	3.32±0.20	13.40±0.13	37.50±0.00
Streptomyces alboviridis	2	2.54±0.08	19.04±7.57	27.50±1.06
Streptomyces albus	3	2.62±0.11	11.25±0.37	39.50±0.35
Streptomyces atroolivaceus	4	3.74±0.21	5.17±0.13	34.00±0.71
Streptomyces caesius	5 .	2.44±0.05	2.14±2.53	30.00±0.35
Streptomyces citreus	6	1.88±0.11	1.07±0.63	27.75±0.18
Streptomyces coelicolor	7	2.74±0.10	2.04±0.00	28.50+0.00
Streptomyces griseinus	8	1.74±0.03	6.64±0.50	24.00±0.00
Streptomyces griseolviridis	9	3.08±0.00	13.75±0.13	19.25±0.18
Streptomyces gougerotii	_,10	3.48±0.14	7.32±0.38	34.75±0.53
Streptomyces matensis	11	2.74±0.06	7.94±0.63	10.00±0.00
Streptomyces minoensis	12	1.66±0.30	6.00±1.01	25.00±0.00
Streptomyces olivaceoviridis	13	3.1±0.19	16.78±0.25	45.00±0.00
Streptomyces olivaceus	14	2.06±0.04	8.57±0.25	40.00±0.53
Streptomyces phacochromogenes	15	2.36±0.12	8.74±0.13	38.50±0.00
Streptomyces prasinus	16	2.00±0.72	7.07±0.50	36.20±0.53
Streptomyces rimosus	17	2.72±0.51	11.43±0.25	39.75±0.18
Streptomyces rochei	18	2.11±0.02	8.39±0.37	38.00±0.71
Streptomyces scabies	19	1.60±0.29	11.78±0.51	16.50±0.71
Streptomyces sp.	20	2.24±0.17	5.78±0.50	15.00±0.71
Streptomyces sp.	21	2.68±0.15	6.25±0.13	33.50±1.06
Streptomyces tetranusemus	22	1.70±0.22	10.15±0.12	11.54±0.17
Streptomyces viridosporus	23	3.34±1.20	2.48±0.37	41.50±0.35
Streptomyces violaceus	24	3.54±1.01	16.07±0.25	37.25±0.53

Table 3
Effect of different incubation period on growth-regulating substance produced by three isolates of *Streptomyces* species. Value listed are expressed as the mean of duplicate determination \pm s.r.

Incubation Period		24			48		60				
Organisms Names	Organisms No.	Total Auxins Eqvuivalent To IAA µg ml ⁻¹	Gibberellins Equivalent To GA ₃ µg ml ⁻¹	Cytokinin Equivalent To Kinetin µg ml ⁻¹	Total Auxins Equivalent To IAA µg ml ^{-l}	Gibberellins Equivalent To GA ₃ µg ml ⁻¹	Cytokinin Equivalent To Kinetin µg ml ⁻¹	Total Auxins Eqvuivalent To IAA µg ml ⁻¹	Gibberellins Equivalent To GA ₃ µg ml ⁻¹	Cytokinin Equivalent To Kinetin µg ml ⁻¹	
Streptomyces olivaceoviridis	13	14.25±4.53	11.87±3.001	12.01±0.21	85.80±1.42	13.62±0.45	21.04±0.68	22.34±0.28	14.14±2.99	14.70±0.50	
Streptomyces rimosus	17	12.73±0.0	4.22±0.52	17.86±0.66	37.96±0.79	14.30±0.73	7.48±0.13	26.46±11.99	6.59±1.50	12.37±0.21	
Streptomyces rochei	18	37.65±0.08	3.58±0.71	2.08±0.71	31.23±0.25	6.94±0.24	5.89±1.00	39.33±5.25	5.84±0.13	4.21±1.01	

Incubation Peri	od (hr)		72			84		96			
Organisms Names	Organisms No.	Total Auxins Eqvuivalent To IAA µg ml ⁻¹	Gibberellins Equivalent To GA ₃ µg ml ⁻¹	Cytokinin Equivalent To Kinetin µg ml ⁻¹	Total Auxins Equivalent To IAA µg ml ⁻¹	Gibberellins Equivalent To GA ₃ µg ml ⁻¹	Cytokinin Equivalent To Kinetin µg ml ⁻¹	Total Auxins Eqvuivalent To IAA µg ml ⁻¹	Gibberellins Equivalent To GA ₃ µg ml ⁻¹	Cytokinin Equivalent To Kinetin µg ml ⁻¹	
Streptomyces olivaceoviridis	13	26.21±0.15	14.22±0.24	16.61±0.40	44.46±9.77	15.41±0.296	28.71±2.03	. 33.92±0.35	13.63±2.5	19.02±0.72	
Streptomyces rimosus	17	30.46±0.64	13.92±0.30	6.67±0.21	39.8±0.69	16.30±0.02	6.73±0.17	33.75±0.01	19.87±2.27	6,31±0.21	
Streptomyces rochei	18	38.84±1.13	6.21±0.71	4.82±0.41	29.68±0.22	8.58±0.22	8.58±0.0	7.19±0.22	17.64±0.0	8.11±0.29	

Table 4

Effect of different incubation periods on biomass weight, α-amylase and proteinase production by three isolates of Sta ptomyces species

	Incubation	-	24	The Control of the State of the Control		48	·		60			72			84	-	-	96	
	Periods Hr.														•				
Organisms Names	Parameters Organisms No.	Biomass Weight g/L	α- Amylase Unit ml		Biomass Weight g/L	α-Amylase Unit ml ⁻¹	Proteinase Unit ml ⁻¹	Biomass Weight g/L	α-Amylase Unit ml ⁻¹	Proteinase Unit ml ⁻¹	Biomass Weight g/L	α-Amylase Unit ml ⁻¹	Proteinase Unit ml ⁻¹	Biomass Weight g/L	α-Amylase Unit ml ^{'1}	Proteinase Unit ml ⁻¹	Biomass Weight g/L	α-Amylase Unit ml ⁻¹	Proteinase Unit ml ⁻¹
Streptomyces	13	2.76±0.07	42.86±0.0	50,75±0.18	3.87±07	18.93±1.01	51.40±0.0	4.39±0.11	16.43±0.50	56.25±0.17	3.61±0.14	16.43±1.01	56,25±0,18	2.94±0.11	16.78±0.25	65.0±0.70	2.78±0.0	15.71±0.50	28.0±0.0
olivaceoviridis																			
Streptomyces	17	2.61±0.0	38,87±0.0	55.00±1.77	3.91±0.01	12.67±0.12	65.50±0.35	3.95±0.03	9.46±0.38	64.50±0.72	2.90±0.1	9.83±0.12	58.50±0.35	2.43±0.08	9,10±0,13	33.00±1.06	2.34±0.05	8.02±0.12	35.0±0.35
rimosus														ger.				₹1	
Streptomyces	18	2.69±0.15	25.00±0.0	25.50±0.35	3.98±1.20	12.42±0.51	25.25±0.18	4.03±2.01	12.30±0.00	27.75±0.18	4.40±0.02	12.32±1.27	29.25±0.18	3.41±0.50	11.74±0.90	38.00±0.00	3.92±0.00	10.00±0.00	17.00±0.01
rochei																			

growth regulators produced by A. brasilense have included auxins, gibberellin-like substances, cytokinins and other unidentified substances [31, 32]. El-Shanshoury [2] carried out a survey of a number of azotobacters from Egyptian soil and demonstrated that some of these micro-organisms contained growth-regulating compounds.

The production of auxins, gibberellins and cytokinins was influenced by the biomass and incubation time [26]. Thus the present investigation clearly showed that the maximum production of enzymes (α-amylase and proteinase) and growth-regulating substances was attained at 84 hrs for Streptomyces olivaceovirids and Streptomyces rimosus, but at 60 hrs for Streptomyces rochei.

REFERENCES

- [1] Waksman, J., 1967. Biosynthesis of α-amylase and protease by *Streptomyces olivaceous* 142. 1. Regulation of α-amylase activity, Acta. Microbiol. 26: 149.
- [2] El-Shanshoury, A.R., 1985. Production of plant growth hormones by certain microorganisms. M.Sc. Thesis, Tanta University, Egypt.
- [3] Prikryl, Z., V. Vancura and M. Wurst, 1985. Auxin formation by rhizosphere bacteria as a factor in root growth, Biol. Plant. 27: 159.
- [4] Katznelson, H. and S.E. Cole, 1965. Production of gibberellins- like substances by bacteria and actinomycetes. Can. J. Microbiol. 11: 733.
- [5] Sobieszczaniki, J., 1966. Studies on the role of microorganisms in the life of cultivated plants. III. Origin of the bacterial substances stimulating the growth of plants. Acta Microbiol. Pol 15: 67.
- [6] Brown, M.E. and S.K. Burlingham, 1968. Production of plant growth substances of *Azotobacter chroococcum*. J. Gen. Microbiol. 53: 135.
- [7] Eklund, E., 1970. Secondary effects of some *Pseudomonas* in the rhizoplane of peat grown cucumber plants, Acta Agric. Second suppl. 17: 7.
- [8] Kampert, M. and E. Strzelczyk, 1984. Effect of pH on production of cytokinin-like substances by bacteria isolated from soil, rhizosphere and mycorrhizosphere of pine (*Pinus silvestris* L.), Acta Microbiologica Polonica. 33: 77.
- [9] Fuji, N., M. Oakubo and H. Mikawarir, 1963. Studies on amylase produced by the actinomycetes: isolation and crystallization of α-amylase, Bull. Fac. Agric. Univ. Miyasaki. 8: 320.
- [10] Hyslop, P. and B.P. Sleeper, 1989. α-amylase of Streptomyces albus, Bact. Proc., 15: 12.
- [11] Shinke, R.K., N.P. Kunimi and H. Nishira, 1975. Isolation and characterization of β-amylase-producing micro-organisms, J. Ferment. Technol. 53: 687.

- [12] Allam, A.M., A.M. Hussein and A.M. Ragab, 1977. Studies on the formation of α-amylase of *Thermonospora* vulgaris, Zentral Bl. Bakteriol. 132: 142.
- [13] Naguib, M.I., K.M. Zeinat and F.A. Mansour, 1978. Protease activity of *Streptomyces antibioticus*, Egypt. J. Bot. 21: 9.
- [14] Gabr, M.A. and F.A. Mansour, 1982. Studies on amylase production by some *Streptomyces* isolated from Egyptian soil. The 3rd Egyptian Congress, of Botany. Mansoura, 17-19 February 1: 59.
- [15] Mansour, F.A., 1985. Studies on the proteolytic enzymes of some actinomycetes. 2nd Conf. Agric. (Mansoura), 45.
- [16] Christen, G.L. and W.C. Wang, 1985. Comparison of the heat resistance of lipase and protease produced by psychrotrophic bacteria and the effect on UHT-milk quality, J. Dairy, Science, 68 (Suppl. 1); 260.
- [17] Niskamen, R. and E. Eklund, 1986. Extracellular protease-producing actinomycetes and other bacteria in cultivated soil, J. Agric. Sci. in Finland. 58: 9.
- [18] Gabr, M.A., F.A. Mansour and G.E.L. Mohamed, 1988. Some metabolic changes in *Streptomyces olivaceus* and *S. parvullus* associated with amylase and protease production as affected by nutrient requirements IV. Effect of microelements, Egypt. J. Microbiol. 23, No. 3, pp. 549
- [19] Mansour, F.A., 1977. Biological and physiological studies on some *Streptomyces* species isolated from Egyptian soil. Ph.D. Thesis, Faculty of Science, Mansoura University, Egypt.
- [20] Shindy, W.W. and O.E. Smith, 1975. Identification of plant hormones from cotton ovules, Plant Physiolo., 55: 550.
- [21] Foda, H.A. and S.S. Radwan, 1962. Straight growth test for hormones and inhibitors using coleoptile of some Egyptian gramineous, Ain Shams Sci., Bull. 8: 381.
- [22] Frankland, B. and P.F. Wareing, 1960. Effect of gibberellic acid on hypocotyl growth ot lettuce seedling, Nature 23: 255.
- [23] Esashi, Y. and A.C. Leopold, 1969. Cotyledon expansion as a bioassay for cytokinins, Plant Physiol. 56: 437.
- [24] Bell, D.J., 1955. Mono- and oligosaccharides and acidic monosaccharides derivatives. In Modern Methods of Plant Analysis, Vol. 2, Ed. K. Paech & M.V., Tracey, pp. 1-53 Berlin: Springer-Verlag.
- [25] Anson, M.L., 1938. Estimation of pepsin, trypsin, papain and cathepsin with hemoglobin, J. Gen. Physiol. 22: 79.
- [26] Hamedo, H.A., 1991. Studies on hormonal activities of some bacteria, M. Sc. Thesis, Mansoura University, Egypt.

- [27] Varner, J.E. and G. R. Chandra, 1964. Hormonal copntrol of enzyme synthesis in barley endosperm. Proc. Not. Acad. Sci. M. S. A. 52: 100.
- [28] Galson, A.W. and P.J. Davies, 1970. Gibberellins. In control mechanisms in plant development, Prentice-Hall, Inc., New Jersey, 184 pp.
- [29] El-Sayed, A.M., L.R.G. Valadon and A.R. El-Shanshoury, 1987. Biosynthesis and metabolism of indole-3-acetic acid in *Streptomyces mutabilis* and *S. atroolivaceus*, Microbios Letters. 38: 85.
- [30] Gordon, A., JR. Stevens and M. Berry Allson, 1988.

- Cytokinin secretion by *Frankia* sp. HEP ArI₃ in defined medium, Plant Physiol. 87: 15.
- [31] Tien, T.M., M.H. Gakins and D.H. Hubbell, 1979.

 Plant growth substances produced by Azospirillum brasilense and their effect on the growth of pearl millet (Pennisetum americanum L.). Applied and Environmental Microbiology 37: 1016.
- [32] Barbieri, P., T. Zanelli, E. Galli and G. Zanetti, 1986. Wheat inoculation with Azospirillium brasilense SP6 and some mutants altered in nitrogen fixation and indole-3-acetic aic production, FEMS Microbiological Letters 36: 87.