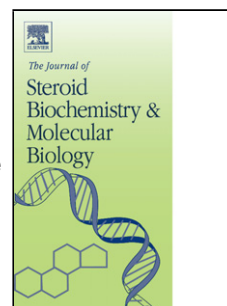


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Comparing metabolic profiles between endurance athlete and non-athlete females reveals differences in androgenic and corticosteroids levels

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Abstract

Endurance training is associated with physiological changes in elite athletes, but little is known about female-specific effects of endurance training. Despite the significant rise in female sports participation, findings from studies performed on male athletes are largely extrapolated to females without taking into consideration sex-specific differences in metabolism. Subsequently, this study aimed to investigate the steroid hormone profiles of elite female endurance athletes in comparison with their non-athletic counterparts. Untargeted metabolomics-based mass spectrometry combined with ultra-high-performance liquid chromatography was performed on serum samples from 51 elite female endurance athletes and 197 non-athletic females. The

results showed that, compared to non-athletic females, certain androgenic, pregnenolone, and progestin steroids were reduced in elite female endurance athletes, while corticosteroids were elevated. The most significantly altered steroid hormones were 5 α -androstane-3 α ,17 α -diol monosulfate (FDR = 1.90×10^{-05}), androstenediol (3 α , 17 α) monosulfate (FDR = 2.93×10^{-04}), and cortisol (FDR = 2.93×10^{-04}). Conclusively, the present study suggests that elite female endurance athletes have a unique steroid hormone profile with implications on their general health and performance.

Keywords: steroids; endurance training; metabolomics; female; athletes; sports.

1. Introduction

Endurance training is a type of exercise that seeks to progressively improve the anaerobic threshold, i.e., the point when anaerobic metabolism is initiated, and results in complex alterations to the muscle metabolism [1]. Skeletal muscle cells respond to regular bouts of endurance exercise via mitochondrial adaptation, including increase in mitochondrial numbers, energetic demand, and fatty acid oxidation [2]. In fact, fatty acid oxidation is improved by endurance training, which also contributes to a decreased utilization of carbohydrates in elite endurance athletes [3]. Endurance training also increases the capacity for oxygen transport, resulting in an enhanced performance and greater resistance to fatigue [4]. Like other types of exercise, endurance training is generally beneficial to one's health, but, at the elite level, it can cause damage to the airways as well as an increased susceptibility to asthma development [1,5].

Despite the dramatic increase in female sports participation in the past decades, studies that are specific to elite female athletes lag behind those on males, and the physiological findings from male-specific research are simply extrapolated to their female counterparts [3,6]. The lack of female-specific data is especially problematic considering that elite female endurance athletes are commonly affected with impaired bone health, low energy availability, and menstrual dysfunction, a trio of conditions that is referred to as the female athlete triad [7]. Disordered eating is also prevalent among elite female endurance athletes, a condition that further exacerbates the female athlete triad and contributes to a deficiency of steroid hormones [8–10].

Steroid hormones can be grouped into two major categories: the corticosteroids and the sex steroids. Corticosteroids, which are further divided into glucocorticoids and mineralocorticoids, mediate stress responses, inflammation, and carbohydrate metabolism, while the sex steroids, which are classified into androgens, estrogens, and progestogens, are involved in the development of sexual characteristics [11]. In a sports context, different concentrations of

circulating steroid hormones, namely testosterone and estrogen, can significantly impact the athletic performance of female athletes.

Circulating levels of testosterone, the primary androgen in males, are generally accepted as the hormonal basis for sex-based differences in athletic performance, with men having circulating testosterone levels that are 15- to 20-fold higher than those in women [12]. Female athletes with circulating testosterone levels above the normal range, either due to doping or hyperandrogenism, exhibited a significantly enhanced performance compared to their normal counterparts [13]. In contrast, estrogen contributes to reduced athletic performance and a higher risk of catastrophic ligament injury in female athletes, and circulating levels of estradiol, the primary estrogen in females, fluctuate through the month due to the menstrual cycle [13,14].

Pre-menopausal females are thought to be better equipped than males to handle endurance sports due to their superior utilization of fatty acids [15–17]. During low- to moderate-intensity exercise, women typically derived a larger share of their exercise energy expenditure from fatty acids compared to their male counterparts [18]. This sex disparity in substrate utilization is thought to be related to the higher levels of circulating estrogen in women as well as differences in body fat composition and distribution [19–22]. It was suggested that the beta oxidation cycle, which degrades fatty acids (acyl-CoA) into two-carbon (acetyl-CoA) units, may be regulated by estrogen due to the fact that several beta-oxidation enzymes, such as medium-chain acyl-CoA dehydrogenase (MCAD) and short-chain hydroxyacyl-CoA dehydrogenase (SCHAD), have upstream estrogen response elements [20].

In a sports context, analysis of steroid metabolites is largely limited to anti-doping surveillance, and metabolomics profiling is not yet a routine analytical activity of the World Anti-Doping Agency (WADA) [23]. However, emerging lines of scientific inquiry suggests that epigenome-induced changes in metabolism influence elite athletic performance [24]. Only a few studies have investigated the differences in steroid metabolite profiles between elite endurance athletes and non-athletes, especially for females. To help fill this gap, this study aims to explore whether elite female endurance athletes exhibit a distinct steroid signature that distinguishes them from non-athletic females.

2. Material and methods

2.1 Study design

Serum samples from 51 consenting elite female endurance athletes from different endurance sports backgrounds were collected from the anti-doping laboratory in Italy. Briefly, blood samples were collected by doping officers in serum separator tubes, then delivered to the anti-doping laboratory within 36 hours under cooling conditions. Once received, samples were immediately centrifuged to separate the serum and then stored at -20°C until analysis. **Figure 1** classifies the sports background of each participant based on the intensity level of the dynamic component, i.e., the estimated percent of maximal oxygen uptake, and the static component, i.e., the estimated percent of maximal voluntary contraction [25]. Elite female endurance athletes were included only if they had competed at the national and/or international levels and had tested negative for prohibited substances. To act as non-elite athlete controls, metabolomics data from 197 healthy females from a Qatar Biobank cohort was extracted [26]. All sample collection was carried out in accordance with the Declaration of Helsinki, and all protocols carried out in this study were approved by the Institutional Review Boards of Qatar University (QU-IRB 1277-E/20).

		Endurance		
		Low <40% Max O ₂	Moderate 40-70% Max O ₂	High >70% Max O ₂
Power	High >50% MVC			Boxing (7) Rowing (5) Cycling (4) Kayaking (1)
	Moderate 20-50% MVC		Athletics (17)	Swimming (15) Hockey (1)
	Low <20% MVC			Tennis (1)

Figure 1. Classification of elite female athletes' sports based on peak dynamic (endurance) and static (power) components achieved while competing.

2.2 Untargeted metabolomics

Serum samples were shipped on dry ice to Metabolon, Inc. (Durham, North Carolina) for untargeted metabolomic profiling in line with established protocol [27]. Briefly, metabolite measurement was carried out using Waters ACQUITY ultra-high-performance liquid chromatography as well as a Thermo Scientific Q-Exactive high-resolution/accurate mass spectrometer interfaced with a heated electrospray ionization (HESI-II) source and Orbitrap mass analyzer operated at 35,000 mass resolution. Detected chromatographic peaks were identified by comparing their fragmentation spectra (MS/MS), mass-to-charge (m/z) ratios, and retention time indices to those of authenticated standards in Metabolon's known chemical reference library [28].

2.3 Statistical Analysis

2.3.1 Multivariate Analysis

Analysis was performed using R version 4.0.3 (www.r-project.org/). Raw data from both cohorts were pooled together, quantile normalized to correct for batch effects, and log transformed. Principal component analysis (PCA) was conducted using SIMCA® version 16.0.2 (Sartorius AG, Göttingen, Germany) with the metabolite missingness cut-off set at 50%. Next, orthogonal partial least squares discriminant analysis (OPLS-DA) was performed using SIMCA® version 16.0.2 to examine the discriminatory effect of the metabolites on the phenotype of interest. For this study, our focus was on 29 metabolites related to steroid biosynthesis.

2.3.2 Univariate Regression

A general linear model was run by regressing the dependent variable on independent confounders and phenotype of interest, i.e., athletes and non-athletic females incorporated as categorical variables. The linear model consisted of PC1 as a confounder and was used to assess the difference in steroid synthesis in athletes versus non-athletic females. PC2 was not included in the model as it showed association with the phenotype in the study. The false discovery rate (FDR) was used to correct for multiple testing, and the significance of association was determined to be $FDR < 0.05$.

3. Results

3.1 Principal component analysis (PCA)

Non-targeted metabolomics was applied to compare the metabolic signatures of 51 female elite athletes and age-matched non-athletic female controls. PCA was performed based on 29 steroid metabolites to capture the global view of the data. Whereas PC1 showed no clear separation between the two groups, PC2 explained 6.53% of the total variance in the data (**Figure 2**). We removed PC2 from the linear regression analysis due to its role as a possible confounder. No other batch effects were seen in our data.

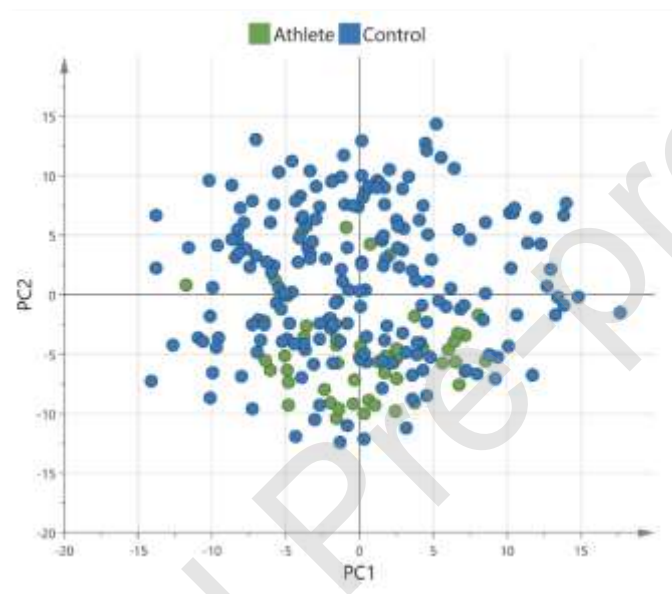


Figure 2. Principal component analysis (PCA) of steroid metabolite levels in elite female endurance athletes and non-athletic females.

3.2 Orthogonal partial least squares discriminant analysis (OPLS-DA)

Based on 29 steroid metabolites, OPLS-DA was performed to identify components that best differentiate between the study groups (elite female endurance athletes and non-athletic females), while dissecting orthogonal components which do not differentiate between these groups. The OPLS-DA score plot showed a remarkable separation of the two phenotypes, suggesting that steroid metabolites are significantly different between the two groups (**Figure 3**). The discriminatory component accounted for 96% of the variation between elite female endurance athletes and non-athletes ($R^2Y = 0.96$, $Q^2Y=0.90$).

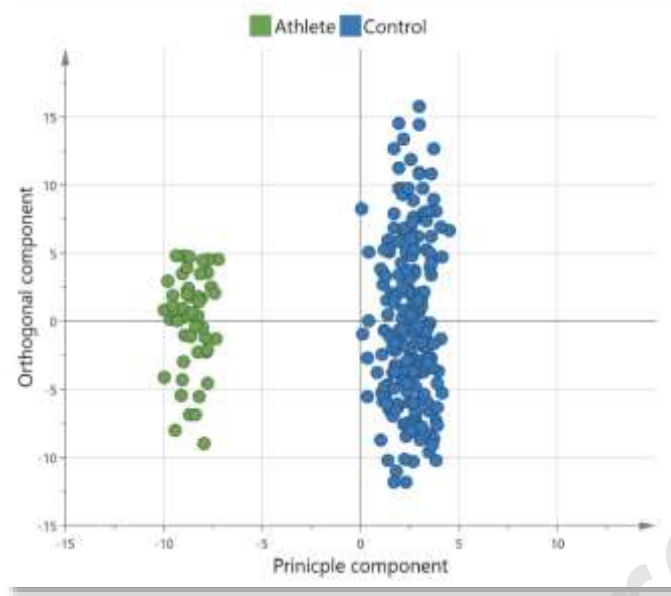


Figure 3. Orthogonal partial least squares discriminant analysis (OPLS-DA) model comparing steroid metabolite levels between elite female endurance athletes and non-athletes.

The corresponding loading scores revealed that androgenic, progestin, and pregnenolone steroids were increased in non-athletic females, while corticosteroids were increased in elite female endurance athletes (**Figure 4**).

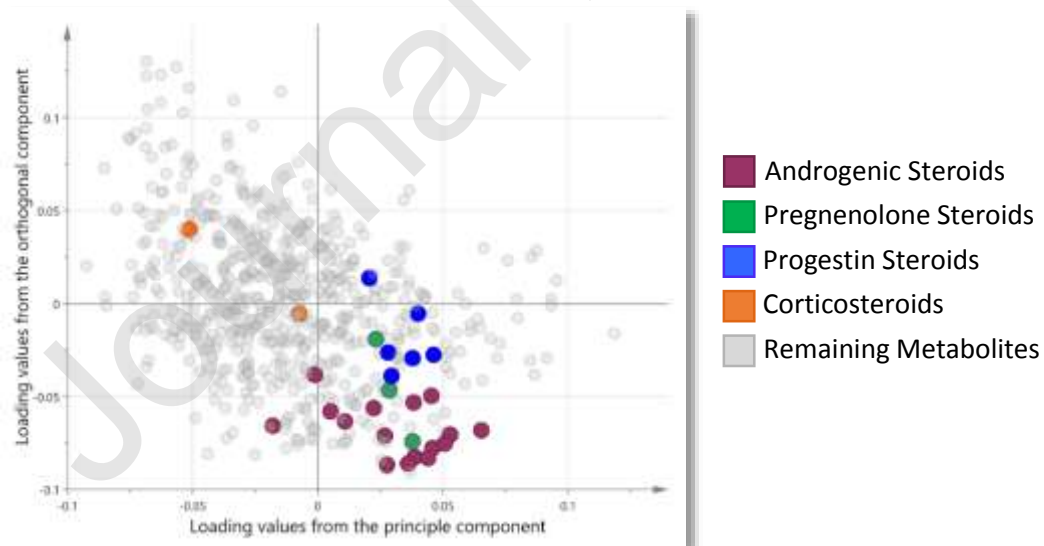


Figure 4. Loading plot of OPLS-DA showing the principal and orthogonal components after quantile normalization.

OPLS-DA also yielded a variable importance projection (VIP) list that reflected the overall importance of the steroid metabolites differentiating elite female endurance athletes and non-athletic females (**Table S1**).

3.3 Linear regression

In order to identify steroid metabolites that significantly change between the two studied groups, the steroid metabolites were regressed against elite female endurance athletes versus non-athletic females after correcting for PC1. Out of 29, 15 steroid metabolites were found to be significantly changed between the two groups (FDR<0.05) (**Table 1**). Androgenic and progestin steroids were lower in elite female endurance athletes, while the corticosteroid cortisol was higher compared to controls. The results of the univariate linear regression model shown in **Table 1** corresponded with the top metabolites shown in the VIP list (**Table S1**) from the multivariate OPLS-DA model.

Table 1. Steroid metabolites that most significantly differentiate between elite female endurance athletes and non-athletic females.

Metabolites	Sub-pathway	Count	Estimate	Std Error	p-value	FDR
5alpha-androstan-3alpha,17alpha-diol monosulfate	Androgenic Steroids	197	-0.45	0.09	6.78×10^{-07}	1.90×10^{-05}
Androstenediol (3alpha, 17alpha) monosulfate (2)	Androgenic Steroids	244	-0.40	0.09	3.13×10^{-05}	2.93×10^{-04}
Cortisol	Corticosteroids	246	0.41	0.09	2.65×10^{-05}	2.93×10^{-04}
Androstenediol (3beta,17beta) monosulfate (1)	Androgenic Steroids	247	-0.38	0.10	1.74×10^{-04}	1.22×10^{-03}
Andro steroid monosulfate C19H28O6S (1)*	Androgenic Steroids	245	-0.47	0.13	2.86×10^{-04}	1.60×10^{-03}
5alpha-androstan-3alpha,17beta-diol monosulfate (2)	Androgenic Steroids	223	-0.43	0.12	6.38×10^{-04}	2.98×10^{-03}
Androsterone sulfate	Androgenic Steroids	248	-0.36	0.11	1.86×10^{-03}	6.52×10^{-03}
5alpha-pregnan-3beta,20beta-diol monosulfate (1)	Progestin Steroids	247	-0.62	0.19	1.75×10^{-03}	6.52×10^{-03}
16a-hydroxy DHEA 3-sulfate	Androgenic Steroids	248	-0.39	0.13	2.67×10^{-03}	8.30×10^{-03}
Androstenediol (3alpha, 17alpha) monosulfate (3)	Androgenic Steroids	248	-0.30	0.10	4.83×10^{-03}	1.13×10^{-02}
21-hydroxypregnenolone disulfate	Pregnenolone Steroids	248	-0.26	0.09	4.66×10^{-03}	1.13×10^{-02}
5alpha-pregnan-3beta-ol,20-one sulfate	Progestin Steroids	185	-0.47	0.16	4.45×10^{-03}	1.13×10^{-02}
5alpha-pregnan-3beta,20alpha-diol monosulfate (2)	Progestin Steroids	247	-0.62	0.23	7.73×10^{-03}	1.66×10^{-02}
Epiandrosterone sulfate	Androgenic Steroids	248	-0.28	0.11	1.27×10^{-02}	2.53×10^{-02}
5alpha-pregnan-3beta,20alpha-diol disulfate	Progestin Steroids	248	-0.41	0.19	2.96×10^{-02}	5.53×10^{-02}

¹ *Count*: number of included samples; *Estimate*: estimated beta value (effect size); *Std error*: standard error; *p-value*: nominal p-value; *FDR*: false discovery rate.

The most significantly associated steroid metabolites in elite female endurance athletes were revealed to be 5 α -androstan-3 α ,17 α -diol monosulfate (FDR = 1.90×10^{-05}), androstenediol (3 α ,17 α) monosulfate (2) (FDR = 2.93×10^{-04}), cortisol (FDR = 2.93×10^{-04}), androstenediol (3 β ,17 β) monosulfate (1) (FDR = 1.22×10^{-03}), 16 α -hydroxy DHEA 3-sulfate (FDR = 8.30×10^{-03}), androsterone sulfate (FDR = 6.52×10^{-03}), and 5 α -pregnan-3 β -ol,20-one sulfate (FDR = 1.13×10^{-02}) (**Figure 5**).

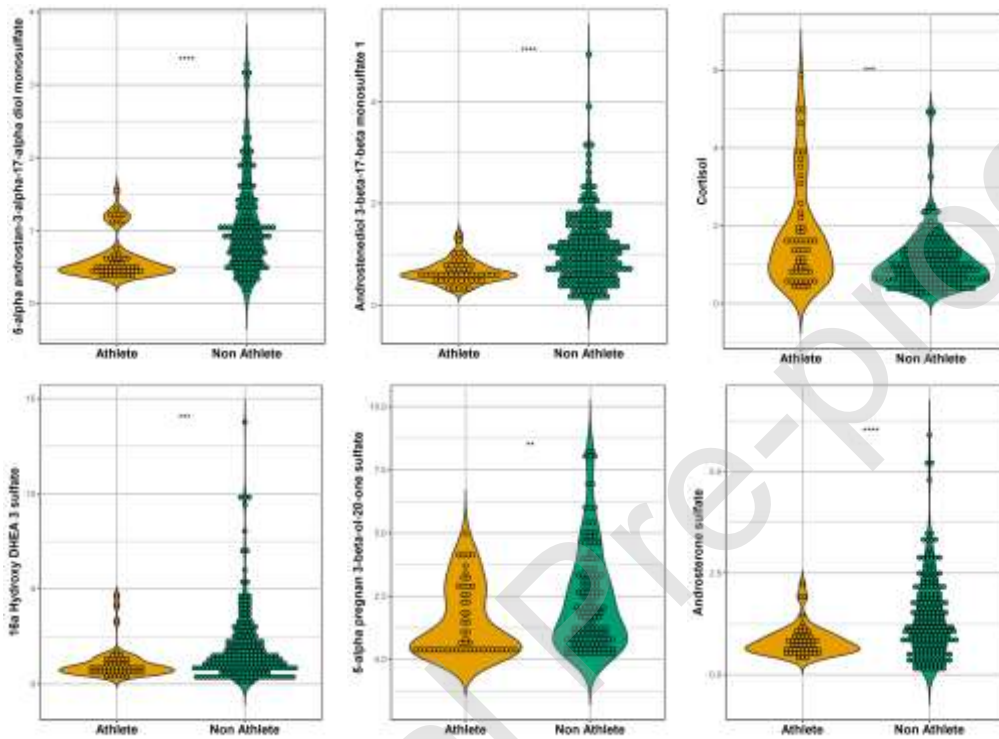


Figure 5. Boxplots of the steroid metabolites that are significantly associated with elite female endurance athletes as revealed by linear regression (FDR < 0.05). **/***/**** indicate a nominal p-value < 0.001/0.0001/0.00001.

The biochemical relationships of the 15 significantly associated steroid metabolites (FDR<0.05) were ascertained from the Kyoto Encyclopedia of Genes and Genomes (KEGG) PATHWAY database (<https://www.genome.jp/pathway/>) (steroid biosynthesis reference pathway map00140) (**Figure 6**).

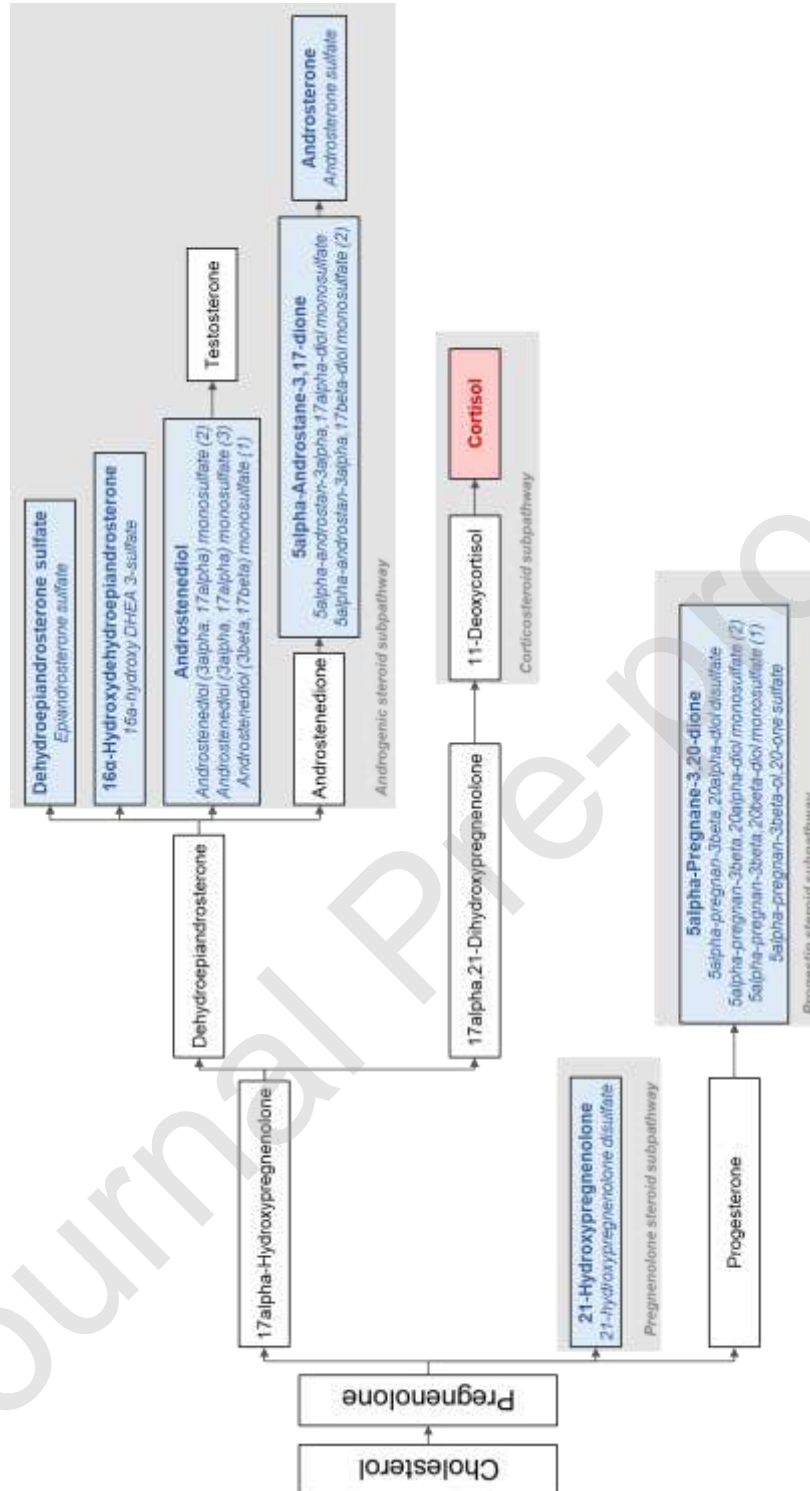


Figure 6. A summary of the biochemical relationships between the steroid metabolites that were significantly associated with elite female endurance athletes. This summary is based on the steroid biosynthesis reference pathway (map00140) from the Kyoto Encyclopedia of Genes and Genomes (KEGG). *Blue shading indicates down-regulated steroid metabolites, while red shading represents up-regulated steroid metabolites. Steroid metabolites are italicized when no exact match is found in KEGG.*

Discussion

Elite athletic performance is dependent on genetics as well physiological adaptation to years of training [29–32]. In particular, endurance-based training remodels the skeletal muscle metabolome by enhancing its oxidative enzyme activity and increasing its mitochondrial content, resulting in a distinct metabolic profile that sets high-endurance athletes apart from their high-power counterparts [27,33]. Previous findings have shown that exercise-induced changes in steroid hormone concentrations are modulated by both the exercise intensity as well as the level of physical fitness [27,34,35]. Taking it further, this study aimed to investigate the differences in steroid metabolism between elite female endurance athletes and non-athletic females.

Our findings revealed that 15 steroid metabolites were significantly altered between elite female endurance athletes and non-athletic females. Except for cortisol, all the significantly altered steroid metabolites were in a sulfated form, causing them to remain inactive until desulfation [36]. Sulfation is an integral part of phase II metabolism, the latter of which encompasses various conjugation reactions that allow metabolites to be excreted from the body [11]. Steroids from four metabolic sub-pathways, i.e., androgens, corticosteroids, progestin, and pregnenolone, were identified as being significantly altered between the two groups.

Androgens can directly enhance sports performance via their effects on behavioral patterns, bone mass, lean body mass, erythropoietin, and visuospatial abilities [13]. In fact, female athletes with high androgen levels are estimated to have a 2-5% greater competitive benefit compared to those with levels within the normal female range [37]. Our findings revealed that among the top changed androgenic metabolites were 5 α -androstan-3 α ,17 α -diol monosulfate (FDR = 1.90×10^{-05}), androstenediol (3 α , 17 α) monosulfate (FDR = 2.93×10^{-04}), androsterone sulfate (FDR = 6.52×10^{-03}), 16 α -hydroxy DHEA 3-sulfate (FDR = 8.30×10^{-03}), and epiandrosterone sulfate (FDR = 2.53×10^{-02}). The precursor to the aforementioned metabolites, dehydroepiandrosterone (DHEA), is classified as a prohibited substance by WADA, but the extent of its impact on athletic performance is still subject to debate, especially in female athletes [38,39].

5 α -androstan-3 α ,17 α -diol monosulfate was the most significantly impacted steroid metabolite in elite female endurance athletes (FDR = 1.90×10^{-05}). Interestingly, a previous study found that, in healthy male subjects, changes in 5 α -androstan-3 α ,17 α -diol monosulfate levels were associated with resistance exercise and not endurance exercise [40]. 5 α -androstan-3 α ,17 α -diol monosulfate levels were significantly decreased in women who used hormone contraceptives and suffered from provoked vestibulodynia, a chronic pain disorder that affected 7 to 16% of the female population [41]. In males, this metabolite was correlated with survival outcome and fatigue levels in prostate cancer patients, and it was identified as a commonly enriched metabolite in young and old Japanese men [42–44]. Metabolome analysis has shown that 5 α -androstan-3 α ,17 α -diol monosulfate is

not one of the steroid metabolites impacted by acetaminophen (paracetamol) use [45]. It is worthwhile to note that the vast majority of studies investigating the impact of acetaminophen on endurance performance were conducted in males [46]. Likewise, androstenediol (3 α , 17 α) monosulfate has been previously linked to the *SULT2A1* gene during quantitative trait locus analysis [47]. *SULT2A1* encodes a sulfotransferase that catalyzes steroid sulfation in the adrenal glands and liver. Certain *SULT2A1* variants modulate levels of dehydroepiandrosterone sulfate (DHEA-S), an adrenal androgen metabolite, in women with polycystic ovary syndrome [48,49]. DHEA-S is the most abundant steroid hormone in humans, and it is notable in that it, along with its precursor DHEA, is responsible for 75% of estrogens in premenopausal women [50]. Via the activity of the 3- β -hydroxysteroid dehydrogenase and 5 α -reductase enzymes, DHEA can be synthesized into epiandrosterone, a testosterone precursor with weak androgenic activity [51].

Epiandrosterone sulfate, the 3-sulfate of epiandrosterone, is associated with the *CYP3A7* variant rs11568825, the latter of which exhibited a significant genome-wide association with DHEA-S [47,52]. Epiandrosterone sulfate has also been identified as a potential biomarker of chronic widespread musculoskeletal pain in adult female twins [53]. Musculoskeletal pain accounts for the majority of young athletes' visits to sports medicine clinics, with female athletes being more likely to suffer from sports-related injuries compared to their male counterparts [54,55]. A growing number of studies have reported that hormonal changes related to the menstrual cycle negatively impact sports performance in elite female athletes [56,57].

Similarly, androsterone sulfate, the 3-sulfate of androsterone, is formed as a result of *SULT2A1* activity, and it is the most abundant 5- α -reduced androgen metabolite in serum [58]. In men, plasma androsterone sulfate levels can be used as a marker of 5 α -reductase activity, the latter of which participates in androgen and estrogen metabolism [59]. Interestingly, screening for 5 α -reductase deficiency can identify disorders of sex development that were previously undiagnosed in young elite female athletes [60].

16 α -hydroxy DHEA 3-sulfate is an estriol precursor that naturally originates from pregnancy but can be pathological in other contexts, as it is strongly associated with increased breast cancer risk in postmenopausal women [61–63]. This metabolite is also significantly downregulated in cases of rheumatoid arthritis, a disease which affects women 4-5 times more frequently than men [64,65]. In male soldiers undergoing military training, 16 α -hydroxy DHEA 3-sulfate is significantly correlated with changes in body mass index and energy balance [66].

Unlike the androgenic steroids, the impact of corticosteroids on sports performance is much less agreed upon, with no current evidence that short-term use could enhance performance [67]. In the present study, cortisol, a glucocorticosteroid, was among the top three most significantly altered steroid metabolites between elite female endurance athletes and non-athletic females (FDR = 2.93×10^{-04}). Cortisol is the most abundant endogenous

glucocorticosteroid in humans, mediating the inflammatory and stress responses as well as immune function and carbohydrate metabolism [68]. Prolonged bouts of exercise result in elevated cortisol levels, which, in turn, maintain blood glucose by stimulating gluconeogenesis [69]. Moreover, extreme long-term endurance training might result in telomere shortening, the latter of which may be mediated by cortisol responsiveness [70,71]. Other than exercise-induced stress, one possible reason that cortisol was upregulated in the current study could be due to an altered carbohydrate utilization pattern in the elite female endurance athletes.

Although plasma, serum, and salivary cortisol levels are correlated with one another, attempts to compare salivary cortisol levels between athletes and non-athletes have yielded mixed results. One systematic review found that only a few studies reported elevated salivary cortisol levels in female athletes but no such elevation during resting conditions, which indicated a low discriminative capacity between groups [72]. A recent study showed that salivary cortisol levels in elite female athletes were significantly higher in the aftermath of a competition compared to rest or training days [73]. Similarly, another systematic review demonstrated that female athletes did not exhibit a significant anticipatory cortisol response prior to a sport competition [74].

Regarding the remaining subpathways, there was little literature available about the steroid metabolites in the pregnenolone and progestin subpathways. **Table 2** compares the steroid metabolites that were significantly changed in this study with the results of previous serum metabolic profiling studies.

Table 2. Previously reported associations of the 15 significantly changed steroid metabolites between elite female endurance athletes and non-athletic females. A tick mark indicates that our findings were confirmed in a previous study.

Cause	Population	Ref.	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15
Acetaminophen use	455 active adults	[45]	✓	✓			✓				✓	✓		✓			
Acute kidney injury	435 cirrhosis patients	[75]									✓						
Depression	99 ART-treated HIV-infected adults	[76]		✓								✓	✓		✓		✓
Early pregnancy	61 young women	[77]	✓				✓	✓	✓						✓	✓	✓
Malaria	199 Gouin children	[78]	✓	✓	✓		✓	✓				✓			✓	✓	
Moderate- and high-endurance sports	191 elite athletes	[27]		✓												✓	
Postmenopausal breast cancer	1,564 female cases and matched controls	[62]	✓			✓		✓			✓			✓			
Provoked vestibulodynia	109 female cases and matched controls	[41]	✓	✓			✓	✓	✓	✓	✓						
Rheumatoid arthritis	236 Arab cases and controls	[64]	✓	✓									✓	✓	✓	✓	✓
Strenuous military training	25 male Norwegian army soldiers	[66]	✓	✓		✓	✓							✓			
T2, T3, and T4 prostate cancers	137 Caucasian male smokers	[79]					✓										
World Trade Center lung injury	223 male firefighters	[80]	✓	✓			✓	✓	✓		✓				✓	✓	✓

1. 16 α -hydroxy DHEA 3-sulfate; 2. 21-hydroxypregnenolone disulfate; 3. 5 α -androstan-3 α ,17 α -diol monosulfate; 4. 5 α -androstan-3 α ,17 β -diol monosulfate (2); 5. 5 α -pregnan-3 β ,20 α -diol disulfate; 6. 5 α -pregnan-3 β ,20 α -diol monosulfate (2); 7. 5 α -pregnan-3 β ,20 β -diol monosulfate (1); 8. 5 α -pregnan-3 β -ol,20-one sulfate; 9. Andro steroid monosulfate C19H28O6S (1)*; 10. Androstenediol (3 α , 17 α) monosulfate (2); 11. Androstenediol (3 α , 17 α) monosulfate (3); 12. Androstenediol (3 β ,17 β) monosulfate (1); 13. Androsterone sulfate; 14. Cortisol; 15. Epiandrosterone sulfate.

A major limitation of this study is the lack of detailed background information about the study participants. Usage of hormonal contraception and different stages of the menstrual cycle introduce hormonal variations between participants and may act as confounding factors that have not been accounted for in this study [13]. However, the effects of these potential confounders were diluted out when considering the mean differences among the elite female athletes who participated in this study. Other limitations to the present study include the relatively small sample size and the different sports backgrounds of the participants, which is an issue because athletes from different sports disciplines exhibit distinct oxidative, inflammatory, and xenobiotic profiles, all of which may have an effect on steroid metabolism [81–84].

Conclusions

Endurance training on a consistent basis results in physiological and metabolic changes. The findings of the present study suggest that elite female endurance athletes have a distinct steroid hormone profile that sets them apart from non-athletic females. Despite limited information about the participants and possible confounding factors influencing their metabolic profiling, the emerging data revealed significant differences in the levels of various steroid metabolites between the two studied groups. It is worthwhile to note that there is a major dearth of studies in the sports sciences focusing on female athletes. The present study should be viewed as a preliminary exploratory study that will need to be followed by controlled replication studies to confirm our findings and investigate if steroid hormone profiles are linked to demographic factors, health, and/or sports performance in elite female athletes.

Declarations

Supplementary Materials: Table S1 – Variable importance projection (VIP) list of steroid metabolites between elite female endurance athletes and non-athletic females.

Author Contributions: Conceptualization, MAE, FD and FB; methodology, MAE; formal analysis, MAE and NRA; resources, FD and FB; writing—original draft preparation, AHT and NRA; writing—review and editing, MAE, NRA and AHT; visualization, NRA and AHT; supervision, MAE; project administration, MAE; funding acquisition, MAE. All authors have read and agreed to the published version of the manuscript.

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References

1. Morici G, Gruttad'Auria CI, Baiamonte P, Mazzuca E, Castrogiovanni A, Bonsignore MR. Endurance training: is it bad for you? *Breathe (Sheff)*. 2016;12:140–7.
2. Booth FW, Ruegsegger GN, Toedebusch RG, Yan Z. Chapter Six - Endurance Exercise and the Regulation of Skeletal Muscle Metabolism. In: Bouchard C, editor. *Progress in Molecular Biology and Translational Science* [Internet]. Academic Press; 2015 [cited 2021 Oct 30]. p. 129–51. Available from: <https://www.sciencedirect.com/science/article/pii/S1877117315001489>
3. Hargreaves M, Spriet LL. Skeletal muscle energy metabolism during exercise. *Nat Metab*. 2020;2:817–28.
4. Amann M, Calbet JAL. Convective oxygen transport and fatigue. *J Appl Physiol (1985)*. 2008;104:861–70.
5. Irewall T, Söderström L, Lindberg A, Stenfors N. High incidence rate of asthma among elite endurance athletes: a prospective 4-year survey. *Journal of Asthma*. Taylor & Francis; 2021;58:735–41.
6. Castanier C, Bougault V, Teulier C, Jaffré C, Schiano-Lomoriello S, Vibarel-Rebot N, et al. The Specificities of Elite Female Athletes: A Multidisciplinary Approach. *Life*. Multidisciplinary Digital Publishing Institute; 2021;11:622.
7. Melin A, Tornberg ÅB, Skouby S, Møller SS, Sundgot-Borgen J, Faber J, et al. Energy availability and the female athlete triad in elite endurance athletes. *Scandinavian Journal of Medicine & Science in Sports*. 2015;25:610–22.
8. Nazem TG, Ackerman KE. The Female Athlete Triad. *Sports Health*. 2012;4:302–11.
9. Schorr M, Miller KK. The endocrine manifestations of anorexia nervosa: mechanisms and management. *Nat Rev Endocrinol*. 2017;13:174–86.
10. Warren MP. Endocrine Manifestations of Eating Disorders. *The Journal of Clinical Endocrinology & Metabolism*. 2011;96:333–43.
11. Olesti E, Boccard J, Visconti G, González-Ruiz V, Rudaz S. From a single steroid to the steroidome: Trends and analytical challenges. *The Journal of Steroid Biochemistry and Molecular Biology*. 2021;206:105797.
12. Handelsman DJ, Hirschberg AL, Bermon S. Circulating Testosterone as the Hormonal Basis of Sex Differences in Athletic Performance. *Endocr Rev*. 2018;39:803–29.
13. Hirschberg AL. Female hyperandrogenism and elite sport. *Endocr Connect*. 2020;9:R81–92.
14. Chidi-Ogbolu N, Baar K. Effect of Estrogen on Musculoskeletal Performance and Injury Risk. *Frontiers in Physiology* [Internet]. 2019 [cited 2022 Feb 5];9. Available from: <https://www.frontiersin.org/article/10.3389/fphys.2018.01834>
15. Tarnopolsky LJ, MacDougall JD, Atkinson SA, Tarnopolsky MA, Sutton JR. Gender differences in substrate for endurance exercise. *Journal of Applied Physiology*. American Physiological Society; 1990;68:302–8.

16. Tiller NB, Elliott-Sale KJ, Knechtle B, Wilson PB, Roberts JD, Millet GY. Do Sex Differences in Physiology Confer a Female Advantage in Ultra-Endurance Sport? *Sports Med.* 2021;51:895–915.
17. Deaner RO, Carter RE, Joyner MJ, Hunter SK. Men are more likely than women to slow in the marathon. *Med Sci Sports Exerc.* 2015;47:607–16.
18. Tate CA, Holtz RW. Gender and fat metabolism during exercise: a review. *Can J Appl Physiol.* 1998;23:570–82.
19. Isacco L, Duché P, Boisseau N. Influence of hormonal status on substrate utilization at rest and during exercise in the female population. *Sports Med.* 2012;42:327–42.
20. Maher AC, Akhtar M, Vockley J, Tarnopolsky MA. Women Have Higher Protein Content of β -Oxidation Enzymes in Skeletal Muscle than Men. *PLOS ONE. Public Library of Science;* 2010;5:e12025.
21. Maher AC, Akhtar M, Tarnopolsky MA. Men supplemented with 17β -estradiol have increased β -oxidation capacity in skeletal muscle. *Physiological Genomics. American Physiological Society;* 2010;42:342–7.
22. Karastergiou K, Smith SR, Greenberg AS, Fried SK. Sex differences in human adipose tissues – the biology of pear shape. *Biol Sex Differ.* 2012;3:13.
23. Botrè F, Georgakopoulos C, Elrayess MA. Metabolomics and doping analysis: promises and pitfalls. *Bioanalysis. Future Science;* 2020;12:719–22.
24. Pitsiladis YP, Tanaka M, Eynon N, Bouchard C, North KN, Williams AG, et al. Athlome Project Consortium: a concerted effort to discover genomic and other “omic” markers of athletic performance. *Physiol Genomics.* 2016;48:183–90.
25. Mitchell JH, Haskell W, Snell P, Van Camp SP. Task Force 8: Classification of sports. *Journal of the American College of Cardiology.* 2005;45:1364–7.
26. Diboun I, Al-Mansoori L, Al-Jaber H, Albagha O, Elrayess MA. Metabolomics of Lean/Overweight Insulin-Resistant Females Reveals Alterations in Steroids and Fatty Acids. *J Clin Endocrinol Metab.* 2021;106:e638–49.
27. Al-Khelaifi F, Diboun I, Donati F, Botrè F, Alsayrafi M, Georgakopoulos C, et al. A pilot study comparing the metabolic profiles of elite-level athletes from different sporting disciplines. *Sports Medicine - Open.* 2018;4:2.
28. Evans A, Bridgewater B, Liu Q, Mitchell M, Robinson R, Dai H, et al. High Resolution Mass Spectrometry Improves Data Quantity and Quality as Compared to Unit Mass Resolution Mass Spectrometry in High-Throughput Profiling Metabolomics. *Metabolomics [Internet].* 2014 [cited 2021 Oct 26];4. Available from: <https://scite.ai/reports/high-resolution-mass-spectrometry-improves-YeEdV3>
29. Schraner D, Schönfelder M, Römisch-Margl W, Scherr J, Schlegel J, Zelger O, et al. Physiological extremes of the human blood metabolome: A metabolomics analysis of highly glycolytic, oxidative, and anabolic athletes. *Physiol Rep.* 2021;9:e14885.
30. Sarzynski MA, Bouchard C. World-class athletic performance and genetic endowment. *Nat Metab.* 2020;2:796–8.

31. Semenova EA, Miyamoto-Mikami E, Akimov EB, Al-Khelaifi F, Murakami H, Zempo H, et al. The association of HFE gene H63D polymorphism with endurance athlete status and aerobic capacity: novel findings and a meta-analysis. *Eur J Appl Physiol.* 2020;120:665–73.
32. Al-Khelaifi F, Yousri NA, Diboun I, Semenova EA, Kostyukova ES, Kulemin NA, et al. Genome-Wide Association Study Reveals a Novel Association Between MYBPC3 Gene Polymorphism, Endurance Athlete Status, Aerobic Capacity and Steroid Metabolism. *Front Genet.* 2020;11:595.
33. Egan B, Zierath JR. Exercise metabolism and the molecular regulation of skeletal muscle adaptation. *Cell Metab.* 2013;17:162–84.
34. Sato K, Iemitsu M, Katayama K, Ishida K, Kanao Y, Saito M. Responses of sex steroid hormones to different intensities of exercise in endurance athletes. *Exp Physiol.* 2016;101:168–75.
35. Al-Khelaifi F, Donati F, Botrè F, Latiff A, Abraham D, Hingorani A, et al. Metabolic profiling of elite athletes with different cardiovascular demand. *Scandinavian Journal of Medicine & Science in Sports.* 2019;29:933–43.
36. Mueller JW, Gilligan LC, Idkowiak J, Arlt W, Foster PA. The Regulation of Steroid Action by Sulfation and Desulfation. *Endocr Rev.* 2015;36:526–63.
37. Berman S. Androgens and athletic performance of elite female athletes. *Curr Opin Endocrinol Diabetes Obes.* 2017;24:246–51.
38. Hahner S, Allolio B. Dehydroepiandrosterone to enhance physical performance: myth and reality. *Endocrinol Metab Clin North Am.* 2010;39:127–39, x.
39. Gravisse N, Vibarel-Rebot N, Labsy Z, Do M-C, Gagey O, Dubourg C, et al. Short-term Dehydroepiandrosterone Intake and Supramaximal Exercise in Young Recreationally-trained Women. *Int J Sports Med.* 2018;39:712–9.
40. Morville T, Sahl RE, Moritz T, Helge JW, Clemmensen C. Plasma Metabolome Profiling of Resistance Exercise and Endurance Exercise in Humans. *Cell Rep.* 2020;33:108554.
41. Labus JS, Mayer EA, Aagaard K, Stains J, Broniowska K, Rapkin A. Reduced concentrations of vaginal metabolites involved in steroid hormone biosynthesis are associated with increased vulvar vestibular pain and vaginal muscle tenderness in provoked vestibulodynia: An exploratory metabolomics study. *Mol Pain.* SAGE Publications Inc; 2021;17:17448069211041852.
42. Huang J, Weinstein SJ, Moore SC, Derkach A, Hua X, Mondul AM, et al. Pre-diagnostic Serum Metabolomic Profiling of Prostate Cancer Survival. *The Journals of Gerontology: Series A.* 2019;74:853–9.
43. Feng LR, Barb JJ, Allen H, Regan J, Saligan L. Steroid Hormone Biosynthesis Metabolism Is Associated With Fatigue Related to Androgen Deprivation Therapy for Prostate Cancer. *Front Cell Dev Biol.* 2021;9:642307.
44. Saito K, Maekawa K, Kinchen JM, Tanaka R, Kumagai Y, Saito Y. Gender- and Age-Associated Differences in Serum Metabolite Profiles among Japanese Populations. *Biol Pharm Bull.* 2016;39:1179–86.

45. Cohen IV, Cirulli ET, Mitchell MW, Jonsson TJ, Yu J, Shah N, et al. Acetaminophen (Paracetamol) Use Modifies the Sulfation of Sex Hormones. *EBioMedicine*. 2018;28:316–23.
46. Grgic J, Mikulic P. Effects of Paracetamol (Acetaminophen) Ingestion on Endurance Performance: A Systematic Review and Meta-Analysis. *Sports*. Multidisciplinary Digital Publishing Institute; 2021;9:126.
47. Al-Khelaifi F, Diboun I, Donati F, Botrè F, Abraham D, Hingorani A, et al. Metabolic GWAS of elite athletes reveals novel genetically-influenced metabolites associated with athletic performance. *Sci Rep*. 2019;9:19889.
48. Goodarzi MO, Antoine HJ, Azziz R. Genes for Enzymes Regulating Dehydroepiandrosterone Sulfonation Are Associated with Levels of Dehydroepiandrosterone Sulfate in Polycystic Ovary Syndrome. *The Journal of Clinical Endocrinology & Metabolism*. 2007;92:2659–64.
49. Louwers YV, de Jong FH, van Herwaarden NAA, Stolk L, Fauser BCJM, Uitterlinden AG, et al. Variants in SULT2A1 affect the DHEA sulphate to DHEA ratio in patients with polycystic ovary syndrome but not the hyperandrogenic phenotype. *J Clin Endocrinol Metab*. 2013;98:3848–55.
50. Maggio M, De Vita F, Fisichella A, Colizzi E, Provenzano S, Lauretani F, et al. DHEA and cognitive function in the elderly. *The Journal of Steroid Biochemistry and Molecular Biology*. 2015;145:281–92.
51. Traish AM, Kang HP, Saad F, Guay AT. Dehydroepiandrosterone (DHEA)—A Precursor Steroid or an Active Hormone in Human Physiology (CME). *The Journal of Sexual Medicine*. 2011;8:2960–82.
52. Ruth KS, Campbell PJ, Chew S, Lim EM, Hadlow N, Stuckey BG, et al. Genome-wide association study with 1000 genomes imputation identifies signals for nine sex hormone-related phenotypes. *Eur J Hum Genet*. 2016;24:284–90.
53. Livshits G, Macgregor AJ, Gieger C, Malkin I, Moayyeri A, Grallert H, et al. An omics investigation into chronic widespread musculoskeletal pain reveals epiandrosterone sulfate as a potential biomarker. *PAIN*. 2015;156:1845–51.
54. Small E. Chronic musculoskeletal pain in young athletes. *Pediatric Clinics of North America*. 2002;49:655–62.
55. Christopher S, Tadlock BA, Veroneau BJ, Harnish C, Perera NKP, Knab AM, et al. Epidemiological profile of pain and non-steroid anti-inflammatory drug use in collegiate athletes in the United States. *BMC Musculoskeletal Disorders*. 2020;21:561.
56. Findlay RJ, Macrae EHR, Whyte IY, Easton C, Whyte LJF (née. How the menstrual cycle and menstruation affect sporting performance: experiences and perceptions of elite female rugby players. *Br J Sports Med*. BMJ Publishing Group Ltd and British Association of Sport and Exercise Medicine; 2020;54:1108–13.
57. Meignié A, Duclos M, Carling C, Orhant E, Provost P, Toussaint J-F, et al. The Effects of Menstrual Cycle Phase on Elite Athlete Performance: A Critical and Systematic Review. *Front Physiol*. 2021;12:654585.
58. Zwicker H, Rittmaster RS. Androsterone sulfate: physiology and clinical significance in hirsute women. *The Journal of Clinical Endocrinology & Metabolism*. 1993;76:112–6.

59. G. Lewis J, George PM, Elder PA. Plasma androsterone/epiandrosterone sulfates as markers of 5 α -reductase activity: Effect of finasteride in normal men. *Steroids*. 1997;62:632–5.
60. Fénichel P, Paris F, Philibert P, Hiéronimus S, Gaspari L, Kurzenne J-Y, et al. Molecular Diagnosis of 5 α -Reductase Deficiency in 4 Elite Young Female Athletes Through Hormonal Screening for Hyperandrogenism. *The Journal of Clinical Endocrinology & Metabolism*. 2013;98:E1055–9.
61. Schweigmann H, Sánchez-Guijo A, Ugele B, Hartmann K, Hartmann MF, Bergmann M, et al. Transport of the placental estriol precursor 16 α -hydroxy-dehydroepiandrosterone sulfate (16 α -OH-DHEAS) by stably transfected OAT4-, SOAT-, and NTCP-HEK293 cells. *The Journal of Steroid Biochemistry and Molecular Biology*. 2014;143:259–65.
62. Moore SC, Mazzilli KM, Sampson JN, Matthews CE, Carter BD, Playdon MC, et al. A Metabolomics Analysis of Postmenopausal Breast Cancer Risk in the Cancer Prevention Study II. *Metabolites*. 2021;11:95.
63. Moore SC, Playdon MC, Sampson JN, Hoover RN, Trabert B, Matthews CE, et al. A Metabolomics Analysis of Body Mass Index and Postmenopausal Breast Cancer Risk. *JNCI: Journal of the National Cancer Institute*. 2018;110:588–97.
64. Yousri NA, Bayoumy K, Elhaq WG, Mohny RP, Emadi SA, Hammoudeh M, et al. Large Scale Metabolic Profiling identifies Novel Steroids linked to Rheumatoid Arthritis. *Sci Rep*. 2017;7:9137.
65. Kvien TK, Uhlig T, Ødegård S, Heiberg MS. Epidemiological aspects of rheumatoid arthritis: the sex ratio. *Ann N Y Acad Sci*. 2006;1069:212–22.
66. Karl JP, Margolis LM, Murphy NE, Carrigan CT, Castellani JW, Madslie EH, et al. Military training elicits marked increases in plasma metabolomic signatures of energy metabolism, lipolysis, fatty acid oxidation, and ketogenesis. *Physiol Rep*. 2017;5:e13407.
67. Verne A, Slack A, Harcourt PR, Budgett R, Duclos M, Kinahan A, et al. Glucocorticoids in elite sport: current status, controversies and innovative management strategies—a narrative review. *Br J Sports Med*. BMJ Publishing Group Ltd and British Association of Sport and Exercise Medicine; 2020;54:8–12.
68. Oakley RH, Cidlowski JA. The Biology of the Glucocorticoid Receptor: New Signaling Mechanisms in Health and Disease. *J Allergy Clin Immunol*. 2013;132:1033–44.
69. HACKNEY AC, WALZ EA. Hormonal adaptation and the stress of exercise training: the role of glucocorticoids. *Trends Sport Sci*. 2013;20:165–71.
70. Steptoe A, Hamer M, Lin J, Blackburn EH, Erusalimsky JD. The Longitudinal Relationship Between Cortisol Responses to Mental Stress and Leukocyte Telomere Attrition. *J Clin Endocrinol Metab*. 2016;102:962–9.
71. Sellami M, Bragazzi N, Prince MS, Denham J, Elrayess M. Regular, Intense Exercise Training as a Healthy Aging Lifestyle Strategy: Preventing DNA Damage, Telomere Shortening and Adverse DNA Methylation Changes Over a Lifetime. *Front Genet*. 2021;12:652497.
72. Cevada T, Vasques PE, Moraes H, Deslandes A. Salivary Cortisol Levels in Athletes and Nonathletes: A Systematic Review. *Horm Metab Res*. © Georg Thieme Verlag KG; 2014;46:905–10.

73. O'Donnell S, Bird S, Jacobson G, Driller M. Sleep and stress hormone responses to training and competition in elite female athletes. *European Journal of Sport Science*. Routledge; 2018;18:611–8.
74. Paridon KN van, Timmis MA, Nevison CM, Bristow M. The anticipatory stress response to sport competition; a systematic review with meta-analysis of cortisol reactivity. *BMJ Open Sport & Exercise Medicine*. *BMJ Specialist Journals*; 2017;3:e000261.
75. Bajaj JS, Garcia-Tsao G, Reddy KR, O'Leary JG, Vargas HE, Lai JC, et al. Admission Urinary and Serum Metabolites Predict Renal Outcomes in Hospitalized Patients With Cirrhosis. *Hepatology*. 2021;74:2699–713.
76. Mukerji SS, Misra V, Lorenz DR, Chettimada S, Keller K, Letendre S, et al. Low Neuroactive Steroids Identifies a Biological Subtype of Depression in Adults with Human Immunodeficiency Virus on Suppressive Antiretroviral Therapy. *J Infect Dis*. 2021;223:1601–11.
77. Handelman SK, Romero R, Tarca AL, Pacora P, Ingram B, Maymon E, et al. The plasma metabolome of women in early pregnancy differs from that of non-pregnant women. *PLoS One*. 2019;14:e0224682.
78. Abdrabou W, Dieng MM, Diawara A, Sermé SS, Almojil D, Sombié S, et al. Metabolome modulation of the host adaptive immunity in human malaria. *Nat Metab*. 2021;3:1001–16.
79. Huang J, Mondul AM, Weinstein SJ, Karoly ED, Sampson JN, Albanes D. Prospective serum metabolomic profile of prostate cancer by size and extent of primary tumor. *Oncotarget*. 2017;8:45190–9.
80. Crowley G, Kwon S, Haider SH, Caraher EJ, Lam R, St-Jules DE, et al. Metabolomics of World Trade Center-Lung Injury: a machine learning approach. *BMJ Open Respiratory Research*. *Archives of Disease in Childhood*; 2018;5:e000274.
81. Al-Khelaifi F, Diboun I, Donati F, Botrè F, Alsayrafi M, Georgakopoulos C, et al. Metabolomics profiling of xenobiotics in elite athletes: relevance to supplement consumption. *J Int Soc Sports Nutr*. 2018;15:48.
82. Sohail MU, Al-Mansoori L, Al-Jaber H, Georgakopoulos C, Donati F, Botrè F, et al. Assessment of Serum Cytokines and Oxidative Stress Markers in Elite Athletes Reveals Unique Profiles Associated With Different Sport Disciplines. *Front Physiol*. 2020;11:600888.
83. Sellami M, Al-muraikhy S, Al-Jaber H, Al-Amri H, Al-Mansoori L, Mazloum NA, et al. Age and Sport Intensity-Dependent Changes in Cytokines and Telomere Length in Elite Athletes. *Antioxidants (Basel)*. 2021;10:1035.
84. Al-Muraikhy S, Ramanjaneya M, Dömling AS, Bettahi I, Donati F, Botre F, et al. High Endurance Elite Athletes Show Age-dependent Lower Levels of Circulating Complements Compared to Low/Moderate Endurance Elite Athletes. *Front Mol Biosci*. 2021;8:715035.