Undergraduate Student, Sciences and Engineering



MONITORING THE PRESENCE AND INVESTIGATION OF TOXIGENIC FUNGI AND MYCOTOXINS IN POULTRY FEED AND ITS PRODUCTS

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Abstract

Contaminating poultry feed and their products with mycotoxins produced by fungi may cause many health effects on animals and human if they were at high concentrations. Therefore, it is imperative to regularly monitor the concentration of mycotoxins specially aflatoxin and ochratoxin A in the poultry feed and their products. In the present study, we demonstrated that Aspergillus flavus was the major contaminant. Ochratoxin A did not exceed the detection limit 50 ng/kg but in one sample has exceeded the European Union maximum limit for aflatoxins of 20 µg/kg. Aflatoxin b1 was detected in chicken liver samples. Almost all samples were contaminated with fungi but only 4 feed samples showed aflatoxin concentration within the detection limit.

Literature Review

Mycotoxins are mainly produced by Aspergillus, Fusarium, Rhizopus, Alternaria and Penicillium (Joshaghani et al., 2013). Some species of fungi produce these mycotoxins which are toxic metabolites that are secreted and found in feed, food and food products. Approaching 300 to 400 mycotoxins are indicated (Pereira et al., 2019). There most leading mycotoxins are aflatoxins (AFs), zearalenone (ZEA), ochratoxin A (OTA), trichothecenes (TRCs) and fumonisins (FMs) (Rai et al., 2015). Some of the mycotoxins are identified as strong carcinogenic agents like AFs B1. The intake of food having mycotoxins may lead to mycotoxicoses which are diseases caused by such intake (Rai et al., 2015).

Objectives

- 1- Explore the presence of mycotoxins in poultry feed samples in Qatar.
- 2- Isolate and identify toxigenic fungi from poultry feed samples.
- 3- Explore the genes encoding mycotoxins and production potential of the isolated fungi.

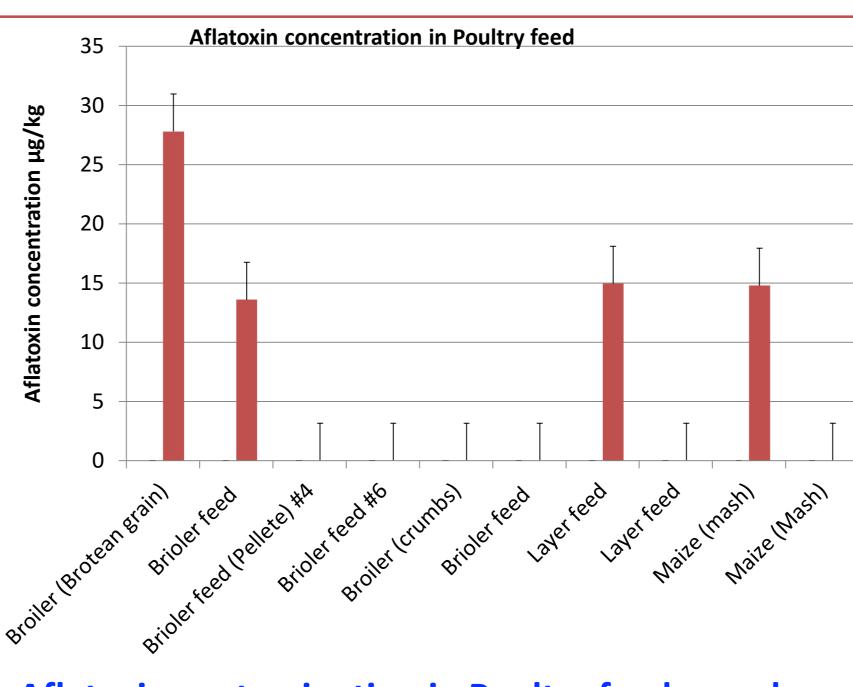
Materials and Methods

Materials: Growth media DRBC, PDB, PDA, YES. Primers TS1, TS4, PAR1, PAR2, FLA1, FLA2, CAR1,CAR2,NIG and Gene-specific primers (OMT-208 – OMT-1232, APA-450 – || PCR amplification of DNA from fungal isolates, using the primers Fla1/Fla2 **APA-1482** and VER-496 — VER-1391, **AoOTA-L** — **AoOTA-R)** (amplicon size 500 bp). PCR, ELISA plate reader, plant DNA extraction kit.

Methods: Fungal species were isolated and identified by inoculating on DRBC, MEA and PDA. DNA was extracted using the plant DNA extraction kit and PCR is used for the molecular | The Publication was made possible by NPRP grant # 8-392identification of isolates primers PAR1/PAR2, FLA1/FLA2, 4-003 from the Qatar National Research Fund (a member CAR1/CAR2, ITS1/NIG. Then, detect aflatoxegenic genes (APA, of Qatar Foundation) and QU grant QUST-2-CAS-2019-34. VER and OMT) Aflatoxin level was detected using total The findings achieved herein are solely the responsibilities aflatoxin ELISA kit and Ochratoxin A was detected by of the authors. Ochratoxin A ELISA kit. Finally, ELISA assay was used for Special thanks for Dr. Zahoor Ul Hassan and Mrs. Reem Almycotoxins quantification of pure isolates.

Results and Discussion

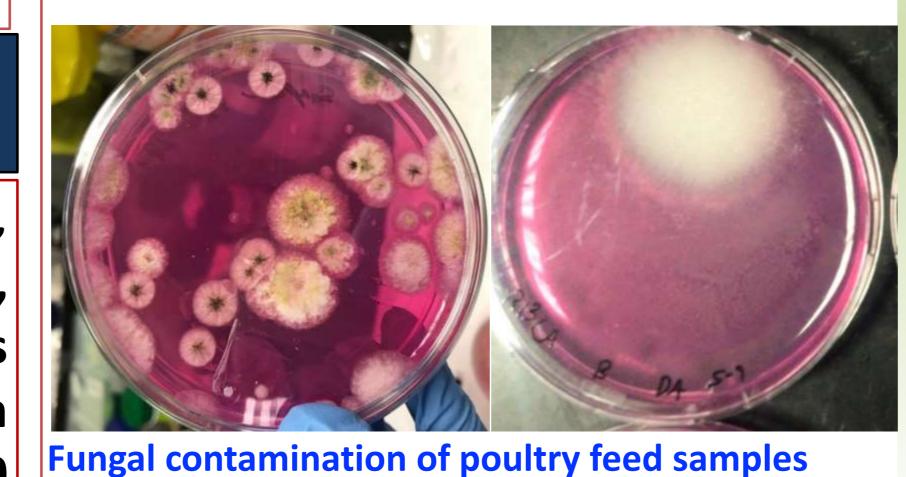
1. Poultry feed samples are contaminated with **Aflatoxins**



- **Aflatoxin was** detected in 2 Maize, 1 layer feed and 1 broiler feed samples out of 10 samples
- **Maize (Brontean** grain) presented highest aflatoxins level (27.8 μ g/kg) followed by layer and broiler feed.

Aflatoxin contamination in Poultry feed samples

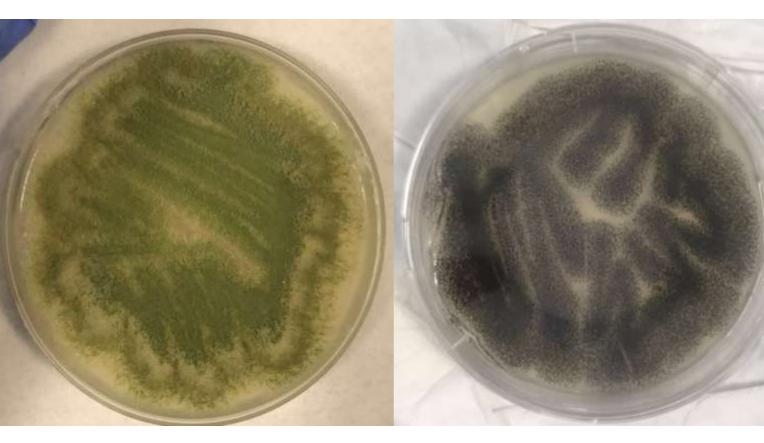
2. Several fungi were isolated from Poultry feed samples Selective and



differential screening of fungal isolates from Poultry feed samples using selective and differential fungal media. **Different colonies**

where isolated to be identified.

3. Morphological identification of aflatoxigenic and Ochratoxigenic fungi, isolated from poultry feed samples

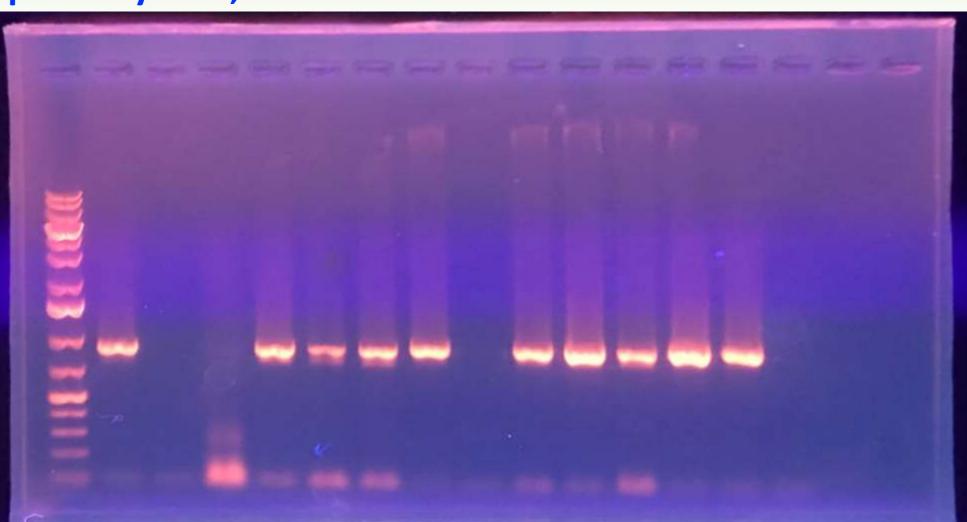


A. Flavus and A. Niger on PDA

Morphological identification through (size, shape, color, sporulation, form, elevation) of aflatoxigenic and Ochratoxigenic fungi. Among total 17 isolates, 3 were A.Niger and 1 A.creceous, 12 were identified either A. flavus or A. parasiticus, since the two species are very morphologically similar.

5. Identification of Aflatoxigenic genes

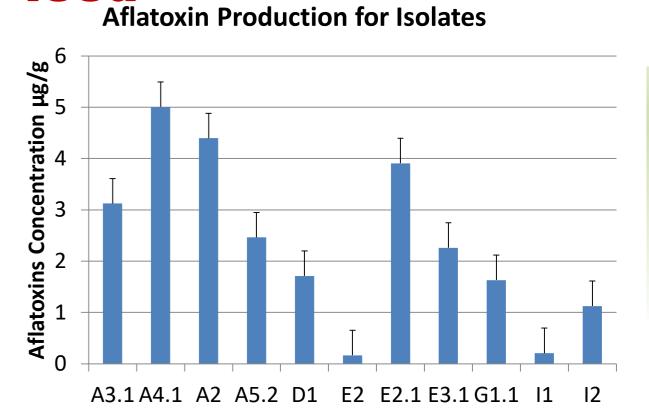
OMT, APA and VER primers were used to confirm the presence of the genes of the aflatoxin biosynthesis pathway: afIP, afIR and afIM.



PCR amplification of DNA from fungal isolates, using the primers VER1/VER2 (amplicon size 895 bp).

1: 100 bp ladder, 2-PCR products of positive control, 3 negative control, others: fungal samples

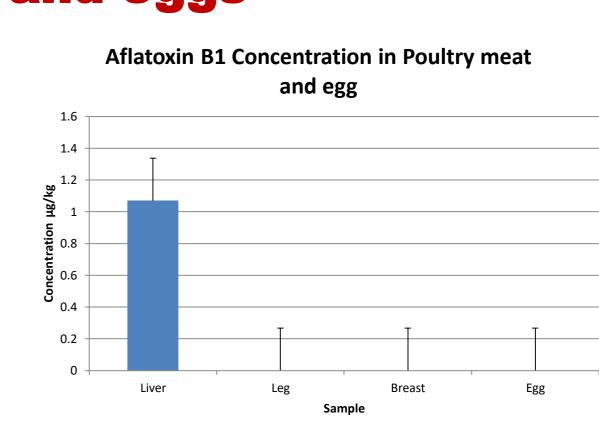
6. Aflatoxin production potential of fungal strains isolated from poultry feed



Aflatoxins production varied between the different samples

Aflatoxins Concentrations in isolated fungi from poultry feed.

7. Detection of Aflatoxin B1 in Meat and eggs



Liver samples **showed Alatoxin B1** concentration average of 1.07 μg/kg

Leg, breast and egg showed negative results for aflatoxin B1

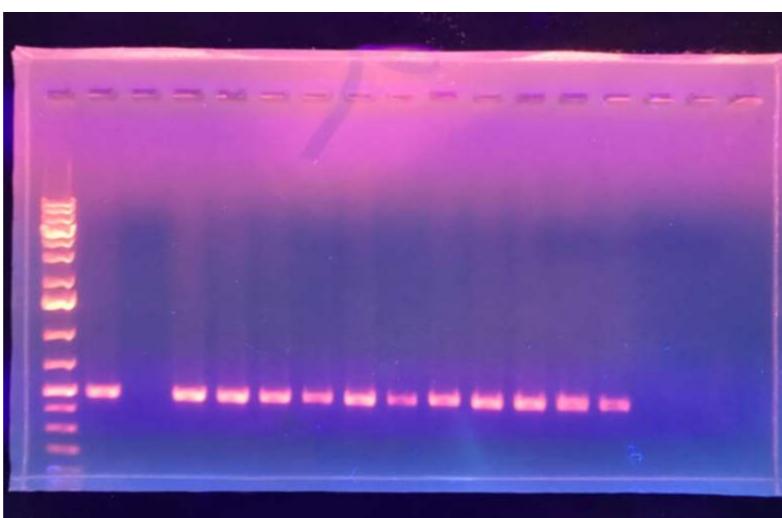
Concentrations of aflatoxin B1 in Poultry liver, leg, breast and egg.

Conclusion

On the basis of our findings, it can be concluded that:

- 1- The explored Poultry feed samples were contaminated with mycotoxins.
- 2- Aspergillus flavus is the most abundant fungus in the Poultry feed samples.
- 3- The synthesis of mycotoxins was confirmed by the presence of the corresponding coding genes in the fungal isolates.
- 4- Chicken liver is contaminated with aflatoxin **B1.**

4. Molecular identification of fungal isolates



were used to confirm that the analysed clones are of A. Flavus.

Primer Fla1/Fla2

Acknowledgments

Asmar.

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