

# Glucose and transferrin liganded PLGA nanoparticles internalization in Non-small cell lung cancer cells

Category: Undergraduate students  
Discipline: Medical, Biomedical and Health sciences

Sarra Benammar, Fatima Mraiche, Jensa Mariam Joseph, Katerina Goracinova  
QU Health, College of Pharmacy, Qatar University

## Background

- Multi functional core shell (CS) nanoparticles (NPs) are among the most efficient drug delivery systems (DDS) for active targeting; their hydrophilic corona improves tumor homing and targeting ligands result in more efficient internalization (Figure (1)).
- These NPs represent promising new DDS for cancer targeting of overly-expressed glucose (Glu) and Transferrin (Tf) receptors at the surface of NSCLC cells.
- Having this background, we hypothesized that the internalization rate (IR) will be higher for Glu or Tf decorated NPs compared to hydrophilic CS PLGA NPs.
- Therefore, the aim of our study was to design hydrophilic CS PLGA NPs decorated with Glu or Tf and to compare their efficacy of internalization to the non-liganded hydrophilic CS PLGA NPs.

## Study Objectives

- To design NPs with improved IR by decorating the PLGA NPs with Glu and Tf ligands.
- To characterize the prepared nanoparticles in terms of particle size, zeta potential, particle size distribution, surface morphology, physicochemical interactions and cytotoxicity.
- To compare the cellular uptake of the prepared Glu and Tf PLGA-NP to that of non-liganded PLGA-NP in A549 NSCLC cells.

## Materials

- Poly(D,L-lactide-co-glycolide), lactide:glycolide 50:50, Mw 38 kDa, PLGA-Glu, Mw: 38 kDa, (PolySciTech (Akina Inc., West Lafayette, IN, USA))
- 5DTAF and transferrin human (Sigma Aldrich)
- PEO-PPO-PEO, Lutrol®F127 (BASF, Germany)
- A549 cell culture line (ATCC, USA)
- MTT (BoosterBio, USA)

## Methods

- PLGA and Glu PLGA NPs preparation by ultrasound assisted nanoprecipitation

Dissolution of the polymer in tetrahydrofuran (THF)

Dropwise addition of polymer THF solution to the Lutrol®F127 water solution and sonication (Sonica Q; amplitude 50%, power 40-43 W and energy 10.000 – 10.800 J)

Removal of THF at 25 C under gentle agitation

Washing of the excess Lutrol®F127 in 3 washing cycles with µQ H<sub>2</sub>O using ultrafiltration

Dispersion in µQ water and Lyophilization

- Transferrin was conjugated to PLGA NPs using EDC/NHS coupling.
- Zeta potential, Z-average hydrodynamic diameter and polydispersity index were then analyzed using Zetasizer Nano ZS (Malvern Instruments, UK).
- SEM was done to characterize the morphology of the prepared NPs using benchtop SEM JCM 6000-JEOL.
- FTIR (PerkinElmer spectrum 400) studies were recorded from 4000 to 400cm<sup>-1</sup> in order to evaluate the interactions in nano-systems.
- Internalization studies were done using A549 cells, fluorescently labeled NPs and a plate reader Victor X4 (Perkin Elmer, MA, USA).
- Growth inhibition studies (MTT test) were performed during 24 and 48h using A549 NSCLC cells.

## Results and Discussion

### Scanning electron microscopy (SEM)

- The micrographs show that the NPs have a spherical shape and low polydispersity.

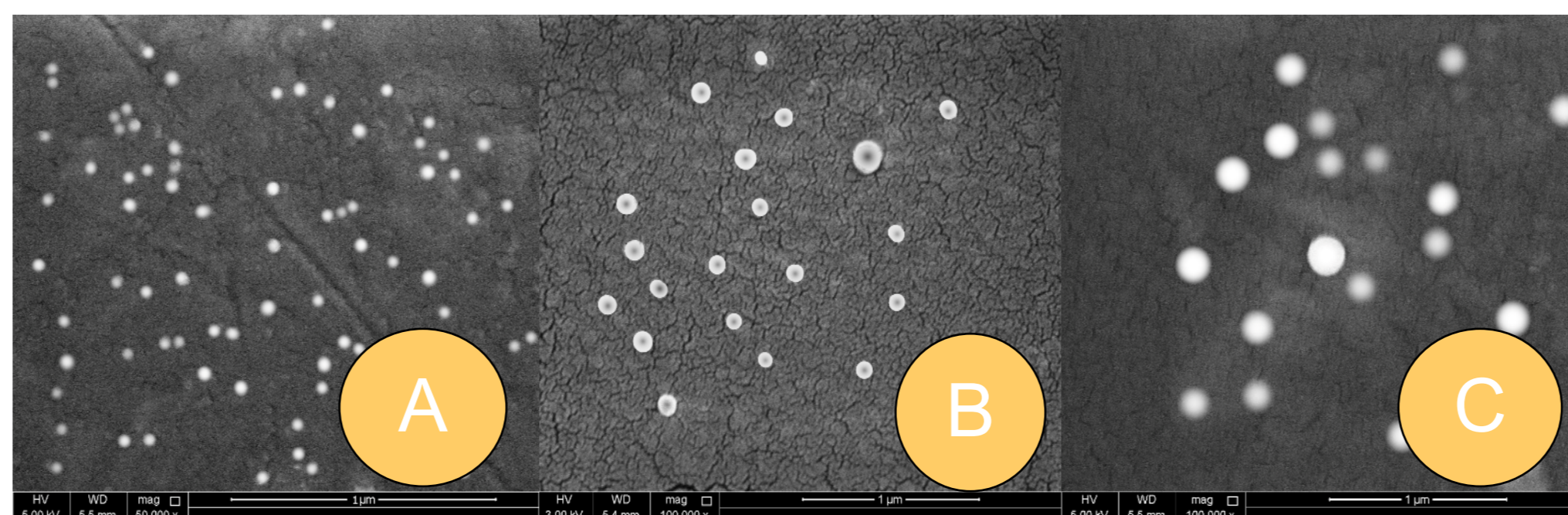


Figure 1: Scanning electron microscopy (SEM) Images of (A) PLGA NPs, (B) Glu-PLGA NPs and (C) Tf-PLGA NPs

### Particle Size and Zeta Potential Analysis

- Particle size, zeta potential and polydispersity index of the PLGA-NPs, Glu-PLGA NPs and Tf-PLGA NPs are presented in Table 1.
- NPs size was from 120 to 130nm, they were uniform with PDI less than 0.025, their Z potential was negative.
- Uniformity of dosage units is important for the consistency of nano-bio interactions, internalization, subcellular localization, efficacy and safety of nanomedicines.
- No significant differences in Z-average Dh and Zeta Potential were noticed with time, which confirms the stability against aggregation in biological media.

Table (1): Z-average hydrodynamic diameter, polydispersity index and zeta potential of the NPs in a phosphate buffer pH 7.4

Sample	z-average Dh(nm)±SD	PDI±SD	Z potential (mV) ±SD
Glu-PLGA NPs	122.9nm±3.4	0.019±0.000	-24.9mV±2.8
Tf-PLGA NPs	137.2nm±2.4	0.013±0.002	-15.1mV±3.4
PLGA-NPs	132.3nm±3.1	0.023±0.003	-31.9mV±4.8

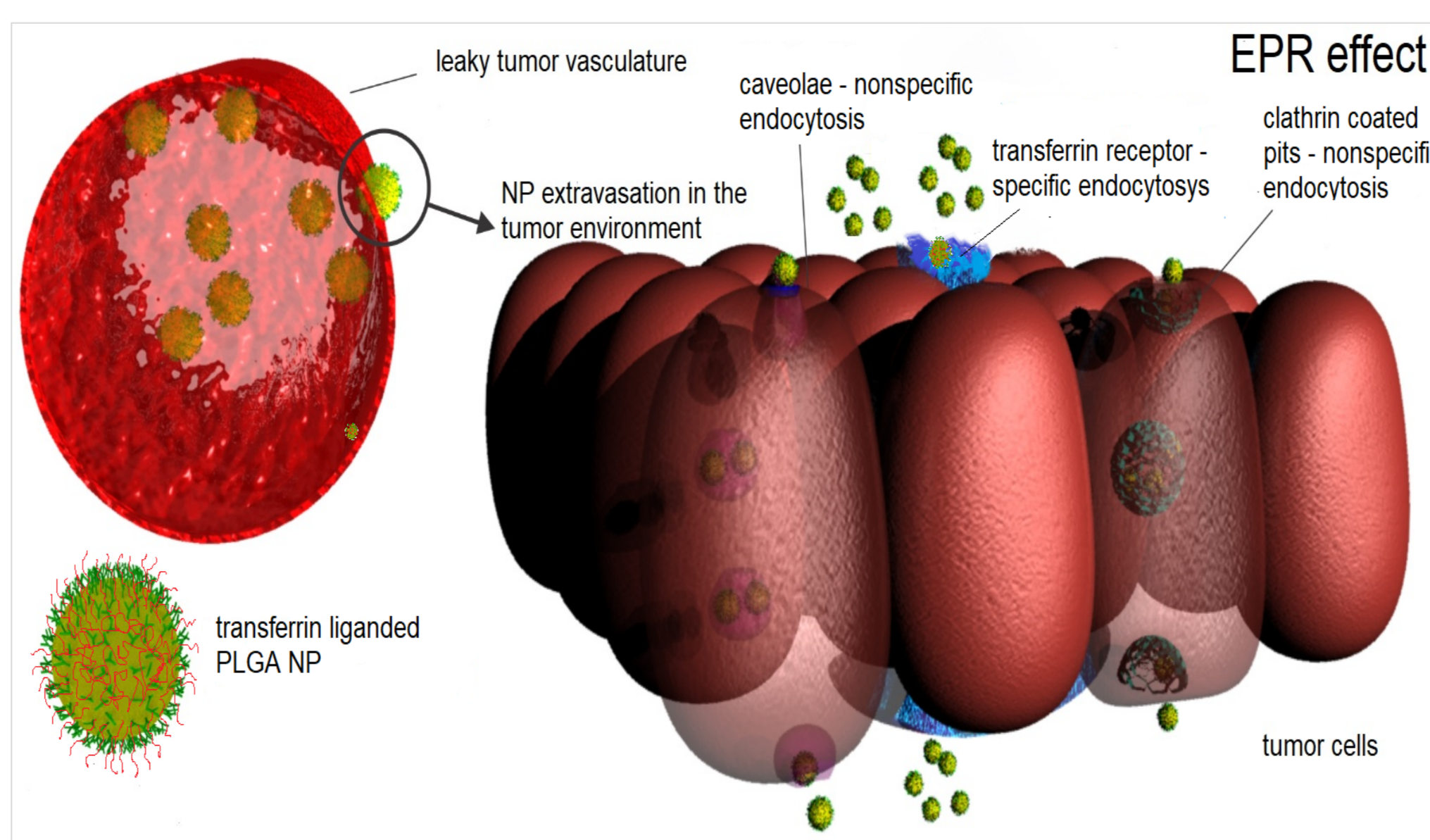


Figure (1): Ligand-receptor interaction and internalization of Tf liganded PLGA NPs after their tumor localization

### FTIR Studies

- Glu-PLGA spectrum did not show any characteristic peaks related to Glu. Slight differences were noticed in the C=O ester band (shifted to 1743 nm); C-H stretches region was richer and there was a new CH<sub>2</sub> vibration at 613 cm<sup>-1</sup>.
- Tf-PLGA NPs spectrum showed peaks of amide I and amide II (at 1648 cm<sup>-1</sup> and 1543 cm<sup>-1</sup>, for amide I and amide II vibrations, respectively), coming from the protein adsorbed at the surface of the NPs.

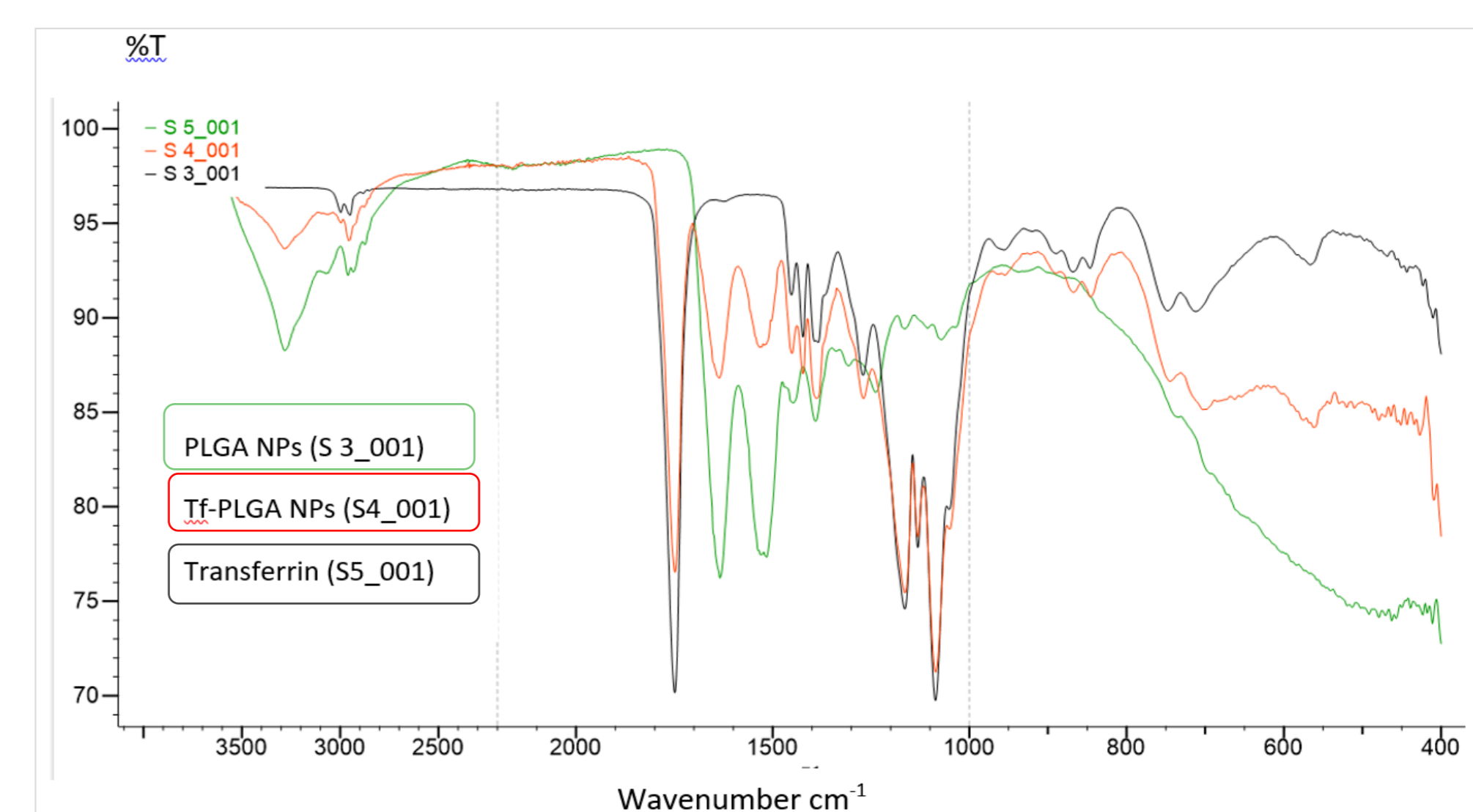
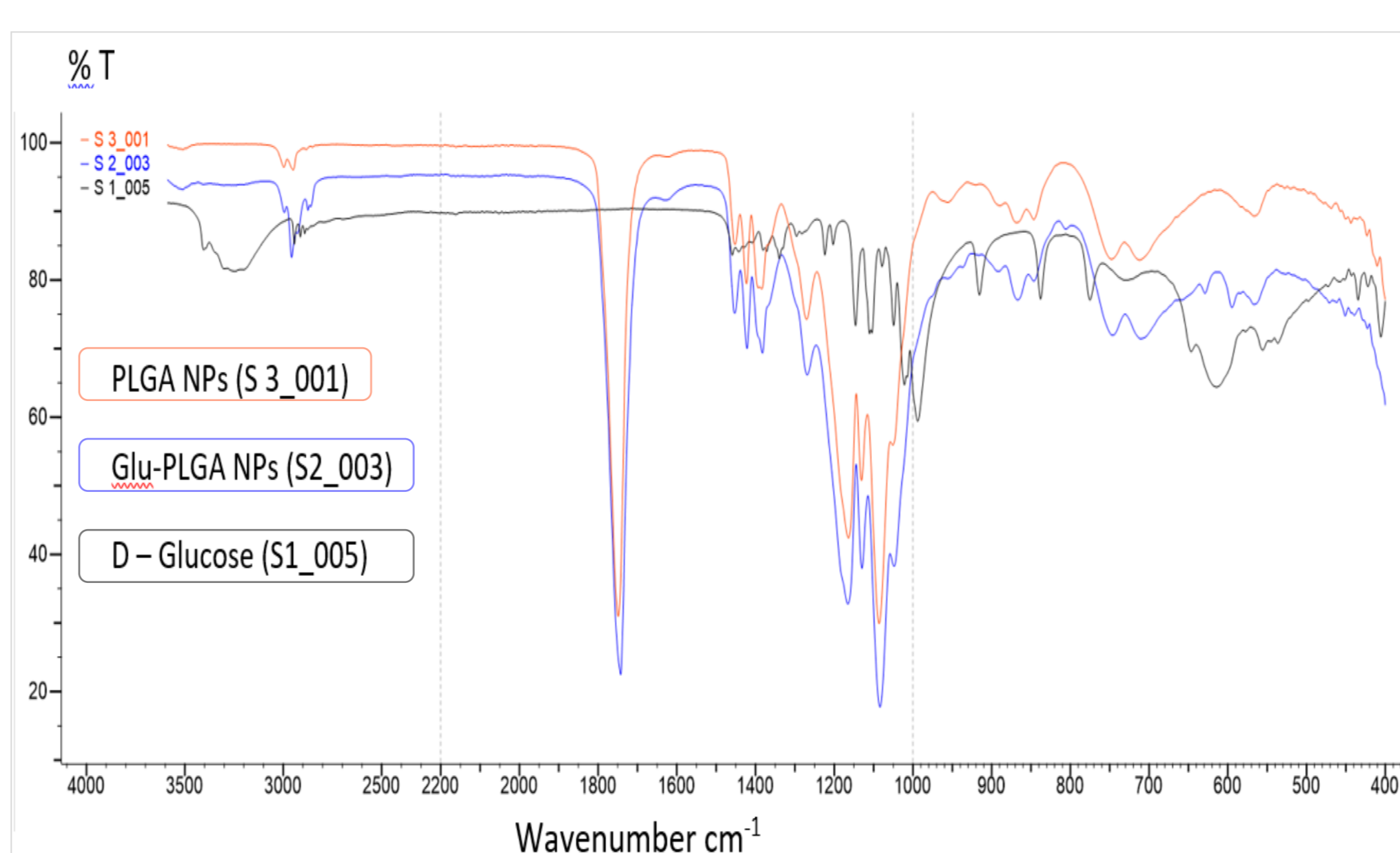


Figure 2: FTIR spectra of (A) PLGA NPs, Glu-PLGA NPs and D-Glucose and (B) PLGA NPs, Tf-PLGA NPs and Transferrin

### Growth Inhibition Studies (MTT test)

- Cells mitochondrial activity and NPs biocompatibility were confirmed using the MTT test. Results show that, in the evaluated concentration range, PLGA NPs, Glu modified and Tf modified NPs show negligible growth inhibition effect on A549 cells.

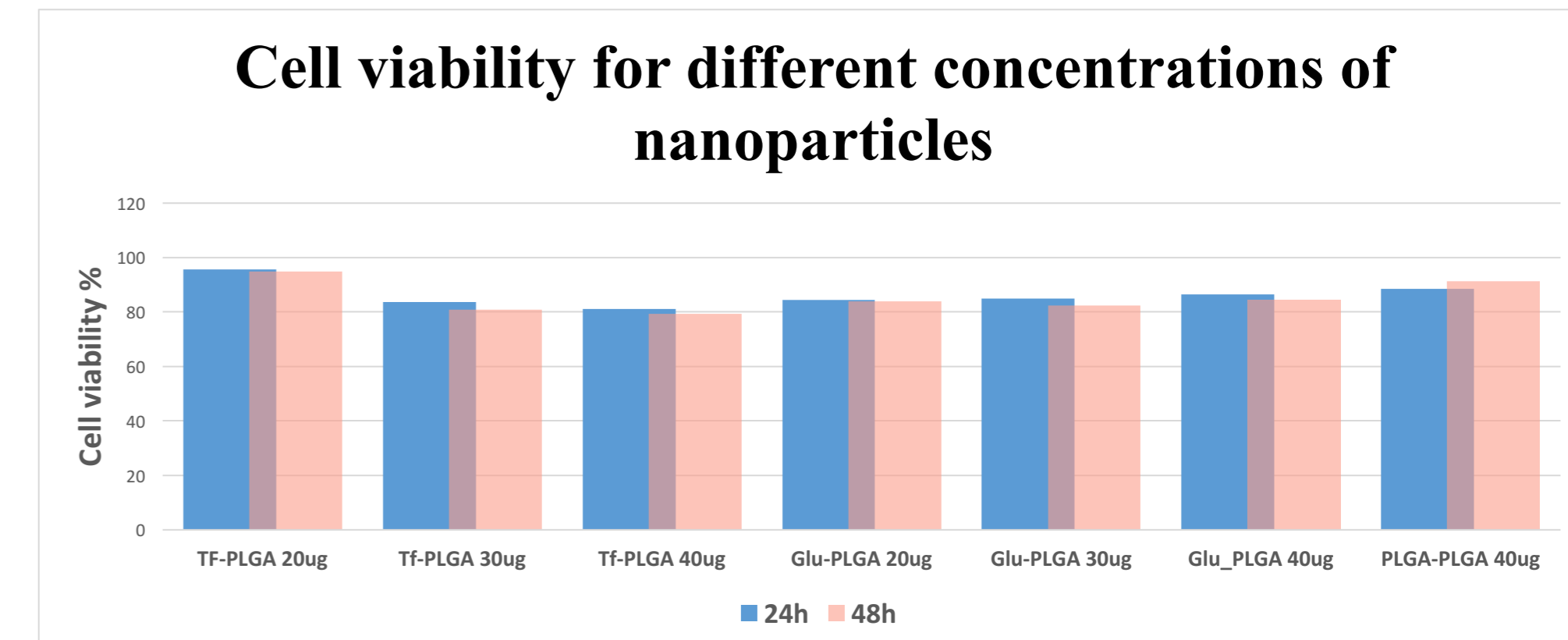


Figure 3: MTT test results for Glu-PLGA NPs, Tf-PLGA NPs and PLGA NPs

### Internalization Studies

- The highest % of internalized NPs was noticed for Glu PLGA NPs.
- IR was affected by the addition of FBS due to protein binding to the NPs and the presence of Tf.

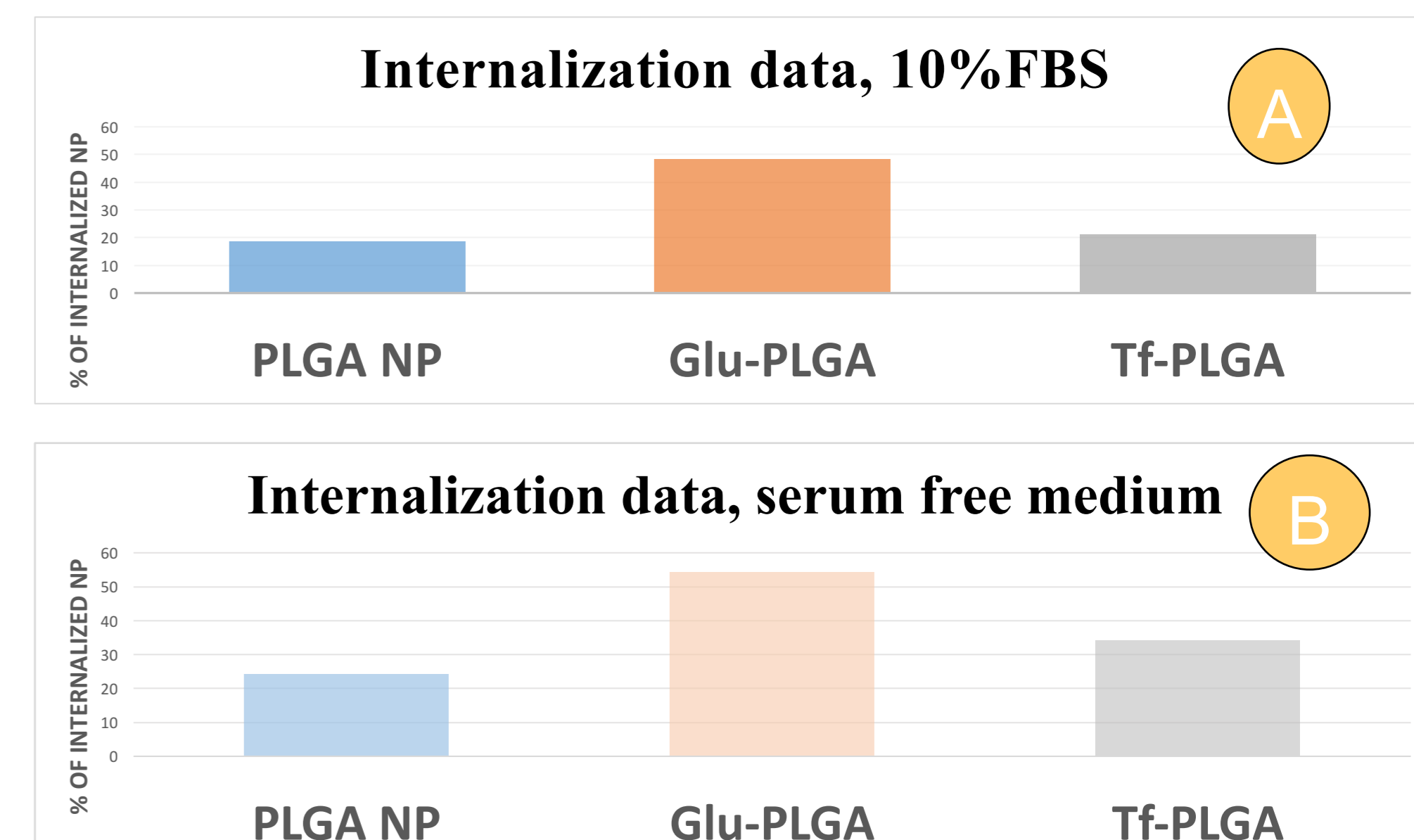


Figure 4: Uptake of particles suspended in (A) DMEM + 10% FBS and (B) in serum free DMEM by A549 cells

## Conclusion

- Receptor specific NPs offer enhanced targetability and increased IR.
- Glu or Tf PLGA NPs were found to have low cytotoxicity and a higher internalization rate compared to non-liganded PLGA NPs. These are important characteristics that can determine the safety of their use as a DDS.
- These findings can be useful in designing future therapeutic alternatives for patients with NSCLC

## Acknowledgment

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## References

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