

Novel Analysis of Glycan Structures: Nanoscale Approach

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Extended Abstract

Glycans are complex saccharide moieties covering all cell surfaces - presented on different biomolecules. Almost 75% of all proteins are glycosylated, and these glycans can form thousands of different structures.[1] Moreover, these structures may slightly change during a specific disease progress – depending on the biomarker observed, there is a possibility to distinguish between healthy individuals and people suffering from a specific disease, mostly cancer (prostate cancer using PSA as a biomarker)[2] or autoimmune diseases (rheumatoid arthritis, system sclerosis observing IgG N-glycosylation).[3] Viral adhesion on cell surface and subsequent penetration is also dependent on the glycan epitopes present on a cell surface.[4, 5] Using nanoscale manipulation of biorecognition elements (antibodies, lectins as glycan-binding proteins or glycans) using self-assembled monolayers (SAMs) allowed to prepare highly sensitive, reproducible and robust biosensors for detection of various analytes – from glycoproteins and whole viral particles to intact cells. Moreover, using SAMs allows to control a biorecognition element's density, orientation and anti-fouling properties of our surfaces.[6] Using nanomaterials like gold nanoparticles of different size or graphene oxide flakes leads to improved characteristics of prepared devices – electrochemical and impedimetric biosensors in this case. Electrochemical devices, mainly in combination with different nanostructures, provide cheap, highly reliable and sensitive platform for glycomic analyses.[7]

We present here a novel approach for a glycoprofiling of various analytes (antibodies, PSA, viral hemagglutinins and viruses and eukaryotic cell lines) using small, low cost, highly sensitive electrochemical devices based on different platforms compared to standardly used LC, CE or MS methods for the glycan analysis. Beside electrochemical impedance spectroscopy and voltammetry, other methods for the surface characterization were used (quartz crystal microbalance, surface plasmon resonance, atomic force and scanning electron microscopy and x-ray photoelectron spectroscopy) and our results were compared to outputs from other analytical methods (protein microarray, enzyme-linked lectin assay and MALDI-TOF MS).

References

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