



# Shrinking the battlefield in cancer therapy: Nanotechnology against cancer stem cells

Queenie Fernandes<sup>a,b,1</sup>, Lubna Therachiyil<sup>c,d,1</sup>, Abdul Q. Khan<sup>c</sup>, Takwa Bedhafi<sup>d</sup>, Hesham M Korashy<sup>d</sup>, Ajaz A. Bhat<sup>e</sup>, Shahab Uddin<sup>a,f,g,h,\*</sup>

<sup>a</sup> College of Medicine, Qatar University, Doha, Qatar

<sup>b</sup> Translational Cancer Research Facility, Hamad Medical Corporation, National Center for Cancer Care and Research, PO. Box 3050, Doha, Qatar

<sup>c</sup> Academic Health System, Hamad Medical Corporation, Translational Research Institute, Doha 3050, Qatar

<sup>d</sup> Department of Pharmaceutical Sciences, College of Pharmacy, QU Health, Qatar University, Doha 2713, Qatar

<sup>e</sup> Department of Human Genetics-Precision Medicine in Diabetes, Obesity and Cancer Program, Sidra Medicine, Doha, Qatar

<sup>f</sup> Academic Health System, Hamad Medical Corporation, Dermatology Institute, Doha 3050, Qatar

<sup>g</sup> Laboratory of Animal Research Center, Qatar University, Doha 2713, Qatar

<sup>h</sup> Department of Biosciences, Integral University, Lucknow, Uttar Pradesh 22602, India

## ARTICLE INFO

### Keywords:

Nanotherapy  
Cancer stem cells  
Signaling pathways  
Pharmacokinetics  
Drug resistance

## ABSTRACT

Cancer remains one of the leading causes of mortality worldwide, presenting a significant healthcare challenge owing to the limited efficacy of current treatments. The application of nanotechnology in cancer treatment leverages the unique optical, magnetic, and electrical attributes of nanomaterials to engineer innovative, targeted therapies. Specifically, manipulating nanomaterials allows for enhanced drug loading efficiency, improved bioavailability, and targeted delivery systems, reducing the non-specific cytotoxic effects characteristic of conventional chemotherapies. Furthermore, recent advances in nanotechnology have demonstrated encouraging results in specifically targeting CSCs, a key development considering the role of these cells in disease recurrence and resistance to treatment. Despite these breakthroughs, the clinical approval rates of nano-drugs have not kept pace with research advances, pointing to existing obstacles that must be addressed. In conclusion, nanotechnology presents a novel, powerful tool in the fight against cancer, particularly in targeting the elusive and treatment-resistant CSCs. This comprehensive review delves into the intricacies of nanotherapy, explicitly targeting cancer stem cells, their markers, and associated signaling pathways.

## 1. Introduction

Cancer is a global health menace that continues to be one of the primary causes of death worldwide. Its incidence is influenced by a myriad of factors, which can broadly be categorized into genetic and environmental. (Laconi et al., 2020; Kaminska et al., 2015; Siegel et al., 2023; Song and Giovannucci, 2016; Vineis and Wild, 2014).

In recent years, an increasing number of studies have stated that cells with stem-cell-like properties may play an indispensable role in oncogenic transformation, and the development of tumors (Wang and Dick, 2005; Koren and Fuchs, 2016; Zhu and Fan, 2018; Ayob and Ramasamy, 2018; Yu et al., 2012) is on the rise. Various other reports also state that voiding the system of stem cells is essential in preventing relapses and

achieving long-term remission (Polyak and Hahn, 2006; Jordan, 2005). Advancements in the knowledge of the specifics of cancer stem cells (CSCs) have unveiled the importance of their targeted eradication in treating cancer. Indeed, the numerous drawbacks and challenges posed by conventional cancer therapeutics (especially in developing resistance to drugs) have made way for newer and more advanced technologies that are much required to improvise.

Nanotechnology is one such area that is aimed at utilizing the characteristics of nanomaterials in the diagnosis, prognosis, and treatment of cancers at both cellular and molecular levels. Interestingly, nanomaterials are very similar to biomolecules in terms of microscopic sizes, and therefore their use may be exploited in the engineering of various potentially useful medical applications.

\* Corresponding author at: Translational Research Institute and Dermatology Institute, Academic Health System, Hamad Medical Corporation, Doha, Qatar.  
E-mail address: [SKhan34@hamad.qa](mailto:SKhan34@hamad.qa) (S. Uddin).

<sup>1</sup> These authors contributed equally to this work.

<https://doi.org/10.1016/j.ejps.2023.106586>

Received 24 June 2023; Received in revised form 7 September 2023; Accepted 18 September 2023

Available online 19 September 2023

0928-0987/© 2023 The Author(s). Published by Elsevier B.V. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

Contrary to atoms and other macroscopic molecules commonly used as drugs in anticancer therapy, nanoparticles possess a high ratio of volume to surface area and can be engineered to have a variety of compositions, shapes, and surface chemical characteristics (Xia et al., 2009; Peer et al., 2007). Customizing these properties molds nanomaterials into an attractive target for developing a new generation of drug delivery vehicles and diagnostic and imaging probes.

The scope of the current work is to bring to the surface the existing burden of cancer while shedding light on the vitality of nanomedicine in cancer. This review will mainly focus on the application of nanomedicine in targeting CSCs, outline the clinical utilization of such technology, and highlight the current challenges and future implications.

## 2. Cancer epidemiology and burden of the disease

Cancer, a pathological condition characterized by the abnormal growth of cells, is known to be the leading cause of death all over the globe (Bray et al., 2021). It was known to be the most dreaded disease of the 20<sup>th</sup> century and is becoming increasingly prevalent in the 21<sup>st</sup> century (Roy and Saikia, 2016) (Fig. 1).

The burden of cancer is rapidly rising in both the aging and growing populations, while the major risk factors are largely linked to socio-economic development (Omran, 2005; Gansler et al., 2010). According to the most recent GLOBOCAN (2020) statistics for cancer, nearly 19.3 million new cancer cases and 10 million cancer-related deaths were observed worldwide in 2020 (Mattiuzzi and Lippi, 2019). Moreover, although cancer is thought to be a gender-neutral disease, men were found to have a 19 times greater incidence rate compared to women (Mattiuzzi and Lippi, 2019). Among the various classes of cancer, breast cancer was identified as the leading cause of cancer incidence across the globe in 2020. Incidence rates are elevated in countries with a higher development index and are, therefore, much more common in such countries than transitioning countries. These elevated statistics among more developed nations may likely arise due to lifestyle-related and/or access to improved cancer diagnostics and mammographic screening programs in these countries (Mattiuzzi and Lippi, 2019) countries.

Similarly, the incidence and mortality rates of lung cancer are much higher in developed countries than developing or transitioning countries. After breast cancer, lung cancer remains the second most commonly diagnosed cancer world wide<sup>16</sup>. In addition, it is also the leading cause of cancer deaths in the year 2020 (Sung et al., 2021). It is not surprising that lung cancer ranks as the leading cause of cancer-related disease and mortality in men across the globe; however, it ranks third in cancer incidence rates in women, after breast and colorectal cancer (Mattiuzzi and Lippi, 2019; Sung et al., 2021). Evidently, due to its relevance as a tobacco smoking disease, lung cancer is less defined among women (Thun et al., 2012). Moreover, apart from tobacco smoking, in certain East Asian countries, the incidence of lung cancer has also been attributed to exposure to air pollutants and particles generated through the household combustion of solid fuels for cooking and other household uses (Mu et al., 2013; Turner et al., 2020). However, the strict enforcement of tobacco control laws and the promotion of anti-smoking campaigns have led to a decline in tobacco usage in both men and women, as observed in China (Lin et al., 2023) and in various other countries (Flor et al., 2021; Wilson et al., 2012).

Finally, colorectal cancer ranks third as the most frequently occurring cancer globally after breast and lung cancer. In addition, colorectal cancer also ranks as the second leading cause of cancer-related death worldwide (Sung et al., 2021). The highest rates of colorectal cancer incidence have been reported in Europe, with Hungary and Norway ranking first in men and women, respectively. However, the incidence rates of rectal cancer are observed to be exceptionally high in East Asian countries. On the other hand, the lowest rates of both colon and rectal cancer are found in the African continent and across south central Asia.

A general trend has been observed in the rise of colorectal cancer among countries with a high development index (Bray and Soerjomataram, 2015; Fidler et al., 2016). In general, factors like lifestyle, diet, obesity, and the consumption of alcohol, red meat, processed foods, etc., have been strongly associated with colorectal cancer (Siegel et al., 2020; Sullivan et al., 2015). Evidently, in many countries, healthier changes to lifestyle and diet and the adequate implementation of screening programs, have deeply reflected in a decline in colorectal cancer cases (Arnold et al., 2020; Edwards et al., 2010).

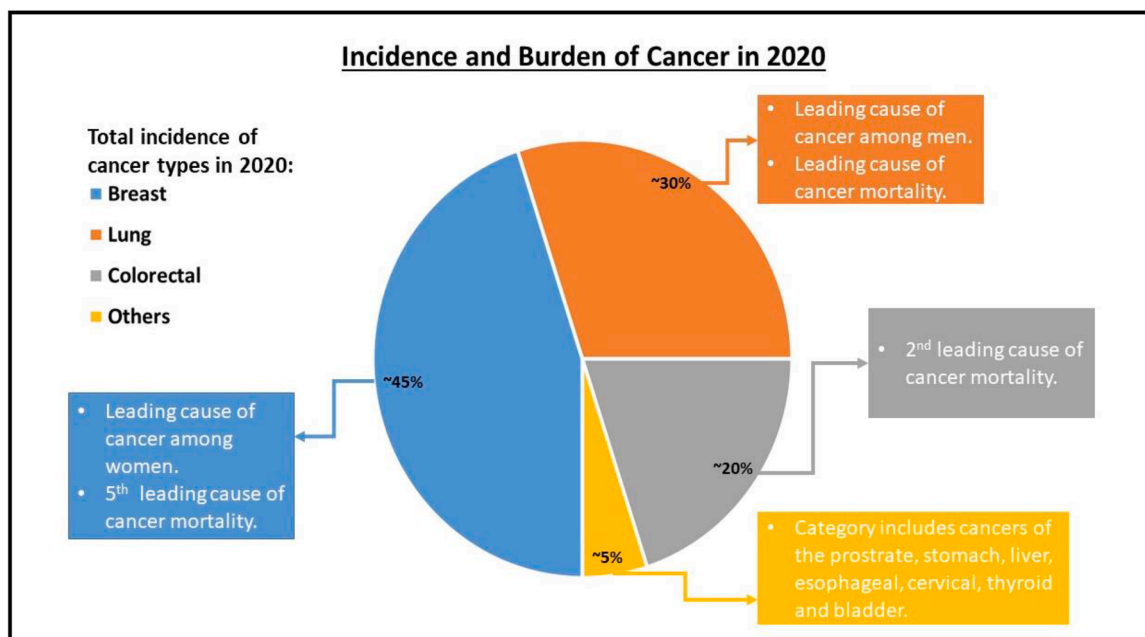


Fig. 1. A depiction of the incidence and burden of cancer (Sung et al., 2021). According to the most recent cancer statistics, breast cancer is the leading type of cancer incidence followed by lung and colorectal cancer. Lung cancer tops as the leading cause of cancer mortality followed by colorectal cancer; while breast cancer ranks 5<sup>th</sup> as a cause of cancer mortality. Breast and lung cancer remain to be the leading causes of gender specific cancers in women and men respectively. (Created with Biorender.com).

Apart from breast, lung, and colorectal cancer ranking as the top three most frequently occurring cancers, other cancers of the prostate, stomach, liver, esophagus, cervix, thyroid, and bladder are also often on the rise across all nations worldwide (Sung et al., 2021). Overall, an estimated 28.4 million new cancer cases are projected to occur in 2040 (Sung et al., 2021). These figures reflect a 47 % increase in the number of cases in 2020, thus depicts the magnitude of the burden of cancer in the near future. Cancer diagnosis and management have come a long way in the last five decades (Roy and Saikia, 2016). In terms of cancer therapy, chemotherapy, radiotherapy, and surgery remain the mainstay of cancer therapeutics.

### 3. Cancer stem cells (CSCs)

Despite effective treatment regimen including chemotherapy and radiotherapy followed by surgical removal of the tumor mass, chemoresistance is a prevailing challenge that contributes to relapse and recurrence in cancer. More specifically, the resistance to chemotherapeutic drugs and recurrence or relapse of cancer may be correlated to the presence of cancer cells with attained stemness properties, commonly known as Cancer Stem Cells (CSCs). In recent years, the relevance of CSCs is becoming significantly important. It is speculated that CSCs are known to originate through the oncogenic transformation of normal stem cells in an organism (Soltysova et al., 2005). Stem cells are defined by their sole characteristics of self-renewal and pluripotency. Interestingly, the identification and targeting of CSCs led to newer implications regarding cancer therapeutics.

CSCs are known to possess excessive capabilities of aberrant differentiation, renewal, tumor initiation, and progression (Ayob and Ramasamy, 2018; Zhou et al., 2023; Yilmaz et al., 2023); therefore, the

sustenance of CSCs has been a major factor leading to poor prognosis and survival in cancer (Palomeras et al., 2018). In addition, CSCs are also shown to be a potent inducer of metastasis (Sahai, 2005). Reports suggest that most drugs that target cancer cells mostly fail to eradicate CSCs, thereby aiding tumor recurrence and permitting the perpetuation of cancer (Brasseur et al., 2017; Cho and Clarke, 2008). A particular study reported that a poly(propylene imine) drug delivery system coupled with paclitaxel showed high therapeutic potential in targeting and eliminating ovarian CSCs, leading to more efficient remission (Tefas et al., 2021). The major characteristics of CSCs are depicted in Fig. 2. Furthermore, various other reports suggest that CSCs could potentially use numerous signaling pathways for their differentiation and self-renewal. The major pathways involved in CSC growth and maintenance are the Wnt/ $\beta$ -catenin, Notch, and Sonic hedgehog pathways (Essex et al., 2019; Borah et al., 2015; Farnie and Clarke, 2007; Clement et al., 2007). In addition, other cell growth signaling pathways, such as PI3K/AKT, JAK/STAT, mTOR, and NF- $\kappa$ B, are found to be deregulated in CSCs, thus further promoting their characteristics of self-renewal (Yang et al., 2020; Ajani et al., 2015; Kroon et al., 2013).

Though the origin of CSCs still remain unclear, several theories have been proposed. Hypothetically, CSCs are either normal stem cells that are mutationally changed to be oncogenic or they are cancer cells acquiring stemness within individual tumors of the same origin (Plaks et al., 2015).

CSCs express specific cell surface markers, such as CD9, CD24, CD44, and CD133 (Abbaszadegan et al., 2017). A higher expression of drug efflux mediating transport proteins such as ATP-binding cassette superfamily G member 2 (ABCG2) is also observed in these specialized cells that facilitate drug efflux in cancer cells thereby evading drug effects. (Kukul et al., 2021). Moreover, most CSCs are reported to have a

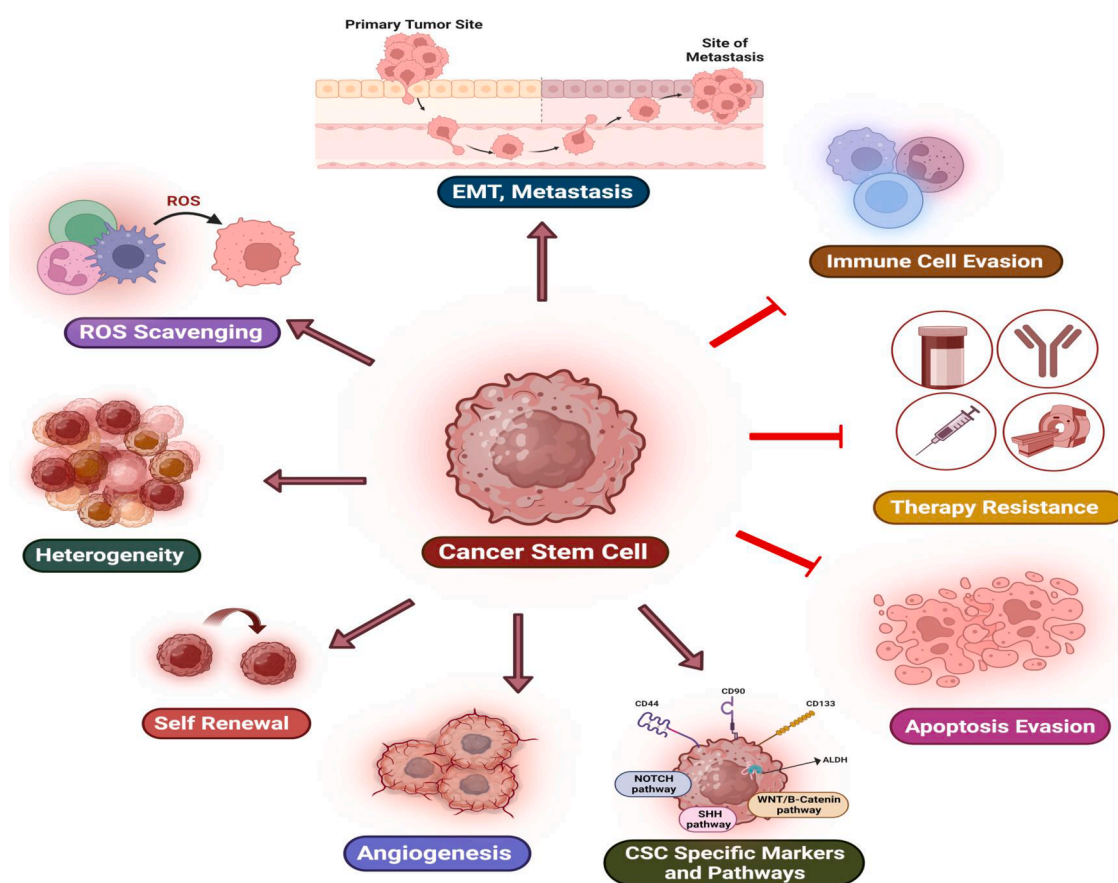


Fig. 2. Schematic representation of characteristic features of Cancer Stem Cells. Abbreviations: EMT- Epithelial to mesenchymal transition, ROS: Reactive Oxygen Species. (Created with Biorender.com).

higher expression of a specific marker named aldehyde dehydrogenase (ALDH) (Abbaszadegan et al., 2017). Enhanced ALDH activity also appears to confer resistance to specific chemotherapeutics wherein reports suggest that ALDH-positive breast cancer cells were capable of forming xenograft tumors with as little as 500 CSCs (Abdullah and Chow, 2013). Moreover, a major mechanism by which CSCs exhibit chemoresistance is through an enhanced DNA damage response through hypoxia inducible factor transcription factors (Olcina et al., 2010). Reportedly, the characteristic features of CSCs, such as the expression of CSC-specific markers, vary across different types of cancers (Cho and Clarke, 2008; Quintana et al., 2008). For instance, in breast cancer, CSCs were found to exhibit the expression of CD44+/ESA+/CD24-/ALDH1 markers, however in leukemia, CSC markers like CD34+/CD38-/CD96 were prominently expressed (Wang and Dick, 2005; Hosen et al., 2007). In gastric cancer, CD15/Lewis X, Lgr5/GPR49 are the majorly observed CSC markers. CD27, CD38, CD20/MS4A1, CD19, and CD138/Syndecan-1 are identified in CSCs of myeloma (Zhao et al., 2017). A comprehensive understanding of the tumor's CSC-mediated plasticity and aggressiveness is essential for enhancing therapeutic approaches that target and eradicate CSCs (Eun et al., 2017).

#### 4. The rise of nanotechnology in the management of cancer

Insufficient drug delivery systems and rising anti-neoplastic therapy resistance are the main challenges to developing effective cancer medicines. The complexity of tumor pathophysiology, highlighted by their intricate blood and lymphatic vasculature, exacerbates these difficulties even more, drastically reducing the potential advantages of conventional cancer therapy. Additionally, the labyrinthine circulatory network and pathophysiological characteristics of malignant neoplasms significantly reduce the efficacy of conventional oncological therapies. Thus, addressing these impediments is paramount to improving the therapeutic outcome in cancer management (Hejmady et al., 2020).

The tumor microenvironment (TME) effectively interacts with cancer cells at various levels including regulating cell proliferation, evading growth suppression and immune-surveillance, evading apoptosis, regulating angiogenesis and promoting EMT/metastasis (Hanahan and Coussens, 2012). Moreover, the intractions between CSCs and microenvironment facilitate drug evading mechanisms towards anti-cancer therapies thus elevating disease recurrence (Albini et al., 2015). Plasticity of CSCs refers to their ability to dynamically interconvert between stem-like and non-stem-like states. This plasticity is influenced by various factors within the tumor microenvironment, including hypoxia, acidity, and interactions with stromal cells, immune cells, and extracellular matrix components (Thankamony et al., 2020). Understanding this ability of CSCs within the TME is however crucial for the development of effective therapeutic strategies for several reasons including identifying critical targets such as signaling pathways, designing targeted delivery systems, and in developing combination therapies. In fact, it is becoming increasingly clear that the tumor microenvironment greatly influences a wide range of factors that may reduce the effectiveness of treatment. A noteworthy aspect of this challenge is the non-cellular mechanisms of drug resistance that emerge from poorly vascularized regions within the tumor mass. These regions present substantial barriers to successfully transporting chemotherapeutic agents, thus thwarting their optimal delivery. Further complicating the matter is the reduced microvascular pressure within the tumor, which severely impedes drug absorption at the tumor site. This physiological characteristic limits the extent of drug penetration into the tumor, curtailing the therapeutic potential of administered agents. These intricacies of the tumor microenvironment and its vascular dynamics underscore the multifaceted challenges facing effective drug delivery and absorption, highlighting the imperative for innovative strategies in cancer therapeutics (Tredan et al., 2007).

Furthermore, cellular mechanisms of drug resistance pose a significant hurdle in effective cancer treatment. This often materializes as a

consequence of tumor cells evading programmed cell death or apoptosis, which is a critical component of the natural defense mechanism against the proliferation of malignant cells. Concurrently, modifications in cellular transport mechanisms, such as the efflux systems mediated by P-glycoprotein, have been recurrently implicated in multi-drug resistance in various cancer types (Halder et al., 2022; Bruno et al., 1987).

Amidst these challenges, an emerging solution resides in the realm of nanotechnology. Nanotechnology may be traditionally defined as the use of nanoparticles specifically within the size range of 5 to 500 nm as target molecular devices in at least one dimension. In particular, previous research attempts to engineer nanotherapeutic agents have cumulated in the development of potent tools like liposomes, nanotubes, polymer nanoparticles, dendrimers, inorganic nanoparticles and other micellar compounds (Ediriwickrema and Saltzman, 2015). Specifically, the association of chemotherapeutic agents with colloidal nanoparticles holds substantial promise for mitigating cellular and non-cellular barriers contributing to drug resistance in cancer. By exploiting the unique properties of these nanoscale entities, we can potentially navigate around the inherent and acquired resistive mechanisms, thereby enhancing the overall therapeutic index of anticancer treatments (McCarthy et al., 2005; Brigger et al., 2002; Davis et al., 2010). Moreover, nanotechnology-based strategies are capable of specifically targeting the complexities of the tumor microenvironment that are key to understanding tumor progression (Mendes et al., 2021).

Furthermore, the microscopic sizes of nanoparticles coupled with their expansive surface area facilitating high-capacity drug loading, render them superior facilitators for enhanced anti-neoplastic therapies (Bedhiafi et al., 2023; Bedhiafi et al., 2023). Examples of nanoparticles leveraged in nano-drug delivery systems extend to a broad range, encompassing metal nanoparticles, nanocapsules, nanoshells, liposomes, and dendrimers. This is further underscored by the extensive body of research validating their potential in cancer therapeutics (Yilmaz et al., 2023). Given these properties, it is indisputable that nanomedicine embodies robust, transformative solutions that are poised to revolutionize the diagnosis, management, and treatment of cancer (Gindy and Prud'homme, 2009).

In particular, advancements in protein engineering, formulation sciences, and drug delivery systems, all forged through the dexterous application of nanotechnology, instill unprecedented optimism in cancer patients. This wave of innovative strategies, powered by the intricate marriage of biotechnology and nano-engineering, continues to invigorate the landscape of cancer therapeutics, offering renewed hope in the ongoing battle against this formidable disease. (Peer et al., 2007).

Several nanomaterials have received FDA approval for use in humans (Peer et al., 2007). The applications of nanomaterials as drug carriers offer the advantage of a considerably enhanced surface area for the coating of antitumor molecules (Kim et al., 2010). In particular, targeting of tumors can be achieved through nanomaterials introduced into the bloodstream which are more efficiently assimilated into the tumor; owing to their improved permeability into the microscopic pores of the vasculature of the early tumor (Hobbs et al., 1998).

More specifically, applying nanomaterials as drug carrier systems could reduce the related adverse effects by inhibiting non-specific uptake into tissues (Prencipe et al., 2009; Alexis et al., 2008; Xu et al., 2023). In line with this, certain studies have reported that the recombinant human tumor necrosis factor-alpha (TNF- $\alpha$ ) was less taken up by healthy tissue when coated on gold nanoparticles than its direct injection. Thus subjects could tolerate much higher doses of the compound, leading to the unleashing of a comparatively enhanced antitumor effect. (Libutti et al., 2010; Visaria et al., 2006). Similarly, yet another study reported that patients could tolerate much higher doses of drugs through nanoparticles with comparatively no side effects (Kim et al., 2004). This is most likely due to the altered pharmacokinetic behavior of the drugs achieved through coupling them with nanomaterials. In addition, nanosystems have also been utilized to reduce toxicity and drug resistance issues (Sun et al., 2023), particularly of conventionally used

cancer drugs like cisplatin (Han et al., 2022; Ghosh et al., 2021). However, despite the success of nanoparticle-based drug delivery systems, reports have also brought to light their potential cytotoxicity, thus highlighting major safety concerns. The toxic effects are known to be triggered both in the intestines as well as in the bloodstream (Araujo et al., 2015). Moreover, the charge and the surface properties of the nanoparticles are also known to produce detrimental interactions within biological systems (Arora et al., 2012). In addition, the cytotoxic effects of nanoparticles may be further pronounced as the pharmacokinetic properties of the drug are liable to change once incorporated into nanosystems (Baldrick, 2010).

The promise of nanomedicine is to provide sustainable therapeutic options against cancer cells that largely counter the drawbacks of conventional chemotherapy and other theranostic drugs while also offering reliable solutions for the early detection/diagnosis of various classes of cancer (Hejmady et al., 2020). Moreover, nanomedicine can also provide highly potent personalized cancer therapies that could offer synergistic solutions to the existing issues of drug resistance (Bar-Zeev et al., 2017).

Moreover, in light of cancer diagnostics, nanotechnology has also risen as attractive agent for cancer imaging. In particular, nanomaterials could serve as contrast agents for diagnostic imaging (Kim et al., 2010). According to a particular report (Harisinghani et al., 2003), nanomaterials were used in prostate cancer to detect lymph node metastases. Here, owing to their smaller sizes, the nanoparticles could traverse through the lymphatic vessels and enter the lymph nodes, thus ensuring uptake by the resident macrophages. Moreover, according to another report (Enochs et al., 1999), using nanoparticles allowed for identifying tumor margins for extended periods on an MRI, as compared to conventional approaches. This is because nanoparticles can easily enter the tumor (owing to their smaller sizes) and take much longer to diffuse out of the tumor (Perrault et al., 2009). Superparamagnetic iron oxide nanoparticles (SPIONs) have been widely used as MRI contrast agents to enhance the visualization of tumors (Li et al., 2013). Similarly, gold nanoparticles can be used as contrast agents in CT scans due to their high X-ray attenuation coefficient (Cormode et al., 2014).

On the whole, the extension of nanotechnology into the various strata of cancer management, like therapeutics, diagnostics, and prognostics, serves as sufficient evidence to highlight the technology's robustness in alleviating the disease's burdens. Table 2 summarizes the common applications of nanotechnology in cancer diagnostics and therapeutics and their associated limitations.

## 5. Nanomedicine in targeting cancer stem cells

Nanotechnology has evolved to play a critical role in modern medicine by developing improved therapeutic options compared to conventional treatment methods. Owing to its effectiveness in modulating distribution, as well as target site accumulation of the drug (that allows for the increase of specificity and decrease of toxicity), nanomedicine has been increasingly used for the enhancement of existing therapeutic options, thus eventually leading to an increase in the therapeutic index of the drugs (Garbayo et al., 2020).

Recently, there is considerable interest in eliminating CSCs using nanoformulation-based anticancer therapies wherein nanoparticles are specifically formulated to accurately identify and target CSCs based on their molecular markers (Lyakhovich and Leonart, 2016). Below are the applications of nanoformulations aimed at the specific targeting of CSC markers and CSC-associated signaling pathways, as evidence of their roles as antitumor mediators (Table 1). Also, Fig. 3 depicts the types, applications, and molecular targets of nanoformulations used to eliminate CSCs.

### 5.1. Nanotherapy targeting CSC markers

#### 5.1.1. CD44-targeted nanotherapy

Cluster of differentiation (CD) refers to certain glycoproteins observed on the tumor cells' surface. As previously mentioned, several antigens belonging to certain CDs have been exclusively identified as molecular markers of CSCs. Recent advancements in nanomedicine have been highly implicated in targeting these markers, especially CD44, CD90, and CD133. CD44 is a cell surface glycoprotein that plays a crucial role in cell-cell and cell-matrix interactions, cell adhesion, and migration. It is highly expressed on the surface of many CSCs and is associated with tumor progression, metastasis, and therapy resistance. Therefore, CD44 is considered a promising target for the selective targeting of CSCs. It is one of the most studied CSC markers that regulate metastasis (Senbanjo and Chellaiah, 2017). The expression of CD44 has been identified in several cancers of the breast (Brown et al., 2011), colon (Su et al., 2011), head and neck (Dubey et al., 2022), ovaries (Martincuks et al., 2020) and pancreas (Al-Hajj et al., 2003). Application of CD44 antibodies via nanoformulations of biodegradable poly [(D, L-lactide co-glycolide)-co-PEG] (PDLGA-co-PEG) polymeric micelles loaded with paclitaxel were reported to elevate the intracellular concentration of the drug and provide effective delivery to the CSCs in breast cancer (Gener et al., 2015). CD44-targeted nanotherapy involves the use of nanoparticles that are functionalized with ligands or antibodies that specifically bind to CD44 expressed on the surface of CSCs. These nanoparticles can be loaded with therapeutic agents, such as chemotherapeutic drugs, siRNAs, or miRNAs, that can inhibit the growth and survival of CSCs. When administered, these nanoparticles selectively bind to CD44-positive CSCs, facilitating the targeted delivery of the therapeutic agents specifically to the CSCs while minimizing off-target effects on normal cells (Xu et al., 2020).

Moreover, magnetic nanoparticles coupled with anti-CD44 antibodies were shown to have an enhanced effect on targeting CSCs in pancreatic and breast cancer (Aires et al., 2016). Yet another study showed that liposomal doxorubicin functionalized with anti-CD44 antibodies effectively increased the circulation time, biodistribution, and efficiency of the drug compared to conventional drug delivery systems (Arabi et al., 2015). In addition, CD44-targeted glutathione-sensitive hyaluronic acid-mercaptopurine prodrug (HA-GS-MP) linked via carbonyl vinyl sulfide was also used in the treatment of acute myeloid leukemia (AML) (Qiu et al., 2017), where the nanoformulation proved to be highly stable and soluble, and thereby attained rapid drug release. Similarly, in colorectal cancer, a recent study reported that SN38 (a chemotherapeutic agent) loaded into human serum albumin (HSA)-hyaluronic acid (HA) nanoparticles (SH/HA NPs) combined with radiotherapy might be a promising therapeutic artifact for colorectal cancer in the near future (Yang et al., 2023).

The impact of CD44-targeted nanotherapy on CSCs includes:

- (1) Inhibition of CSC Growth and Survival: The targeted delivery of therapeutic agents to CD44-positive CSCs can lead to the inhibition of key signaling pathways (e.g., Wnt/ $\beta$ -catenin, Notch, Hedgehog) that are essential for the self-renewal and survival of CSCs (Chen et al., 2018).
- (2) Induction of Apoptosis: The targeted delivery of chemotherapeutic drugs or siRNAs that induce apoptosis can lead to the selective elimination of CD44-positive CSCs (Vinogradov and Wei, 2012).
- (3) Inhibition of CSC Migration and Invasion: CD44 plays a crucial role in the migration and invasion of CSCs. Targeting CD44 with nanoparticles can inhibit these processes, thereby preventing the metastasis of CSCs (Hassn Mesrati et al., 2021).
- (4) Sensitization to Other Therapies: Targeting CD44-positive CSCs with nanoparticles can lead to the downregulation of multidrug resistance genes and the sensitization of CSCs to other therapies, such as radiation or chemotherapy (Vinogradov and Wei, 2012).

**Table 1**

List of nanoformulations targeting CSC specific markers and signaling pathways. Abbreviations: DOX: Doxorubicin, DS-PLGA: poly lactic-co-glycolic acid -encapsulated DiSulfuran, GSI DAPT:  $\gamma$ -secretase inhibitors (N-[N-(3,5-difluorophenacetyl)-L-alanyl]-S-phenylglycine t-butyl ester), HA-CS NP: hyaluronic acid -chitosan nanoparticles, HA-SS-PLGA: hyaluronic acid-cystamine-poly(lactic-co-glycolic acid), IONP: Iron Oxide Nano Particle, MSN: Mesoporous Silica Particles, NanoHHI: Nanoparticle Encapsulated Hedgehog Pathway Inhibitor HPI-1, PaxSLN: Paclitaxel-encapsulated core-shell nanoparticle, PDLGA-co-PEG: Poly(ethylene glycol)-block-Poly(lactic-co-glycolic acid), PEI-PGMA: Polyethyleneimine-PolyGlycidal Methacrylate polyethyleneimine.

Nanoformulations	Target	Type of Cancer	Samples /cell lines ( <i>In vitro</i> / <i>In vivo</i> )	Significance	Reference
<b>CSC Marker-based Nanotherapy</b>					
Anti-CD44 antibodies + nano-formulations of PDLGA-co-PEG polymeric micelles loaded with paclitaxel.	CD44	Breast	<ul style="list-style-type: none"> <li>• MCF7 cell line</li> <li>• MCF7-ALDH1A1/tdTomato</li> <li>• MDA-MB-231</li> <li>• HCT8,</li> <li>• HCT116</li> <li>• HCT116-ALDH1A1/tdTomato</li> <li>• MCF7-ALDH1A1/tdTomato cells</li> </ul>	Improved intracellular concentration of the drug and enhanced delivery to CSCs.	<a href="#">Gener et al. (2015)</a>
Magnetic nanoparticles + anti-CD44 antibodies		Breast and Pancreas	<ul style="list-style-type: none"> <li>• Panc-1</li> <li>• MDA-MB231 cancer cell lines</li> <li>• MCF-10A cell line</li> </ul>	Enhanced targeting of CSCs	<a href="#">Aires et al. (2016)</a>
PPI drug delivery system + paclitaxel		Ovarian	<ul style="list-style-type: none"> <li>• Ascitic fluid with cancer cells</li> <li>• Athymic nu/nu mice</li> </ul>	Improved therapeutic elimination of CSCs.	<a href="#">Shah et al. (2013)</a>
Nano-formulation of liposomal doxorubicin + anti-CD44 antibodies		Colon	<ul style="list-style-type: none"> <li>• C-26 cancer cell line</li> </ul>	Increased the circulation time, biodistribution and efficiency of the drug.	<a href="#">Arabi et al. (2015)</a>
SN38 loaded with SH/HA nanoparticles + radiotherapy		Colorectal	<ul style="list-style-type: none"> <li>• SW620, HT29 cancer cell lines</li> </ul>	Promising CSC-targeted therapy.	<a href="#">Yang et al. (2023)</a>
Salinomycin + nanoparticles loaded with CD133 aptamers	CD133	Osteosarcoma	<ul style="list-style-type: none"> <li>• Saos-2 CD133(+) cancer cell line</li> </ul>	Offered high specificity and improved drug efficiency to CD133+ CSCs, resulting in tumor cell death.	<a href="#">Ni et al. (2015)</a>
MSNs + anticancer drugs / siRNA		laryngeal carcinoma	<ul style="list-style-type: none"> <li>• Hep-2 cells</li> <li>• BALB/c-nu/nu mice</li> </ul>	Improvement in therapeutic outcome.	<a href="#">Qi et al. (2016)</a>
Nanoencapsulation of curcumin		Brain	<ul style="list-style-type: none"> <li>• Embryonal tumor derived lines: DAOY and D283Med</li> <li>• Glioblastoma neurosphere lines: HSR-GBM1 and JHH-GBM14</li> </ul>	Enhanced its anti-tumor effects	<a href="#">Lim et al. (2011)</a>
CdSe core nanocrystals (Trifluoperazine + anti-CD90 antibody)	CD90	Leukemia	<ul style="list-style-type: none"> <li>• CD90<sup>+</sup> leukemia CSCs</li> </ul>	Enhanced cancer cell death.	<a href="#">Asghari et al. (2019)</a>
PaxSLN core-shell nanoparticle	ALDH	Colorectal	<ul style="list-style-type: none"> <li>• CT26 colorectal carcinoma animal model</li> </ul>	Enhanced targeting of ALDH expressing cells.	<a href="#">Dastidar et al. (2019)</a>
DS-PLGA Nanoparticles		Liver	<ul style="list-style-type: none"> <li>• Huh7</li> <li>• PLC/PRF/5 cancer cell lines</li> </ul>	Improve half-life of the drug.	<a href="#">Wang et al. (2017)</a>
<b>CSC-Associated Signalling Pathway-based Nanotherapy</b>					
Cationic lipid-protamine nanoparticles	Wnt/ $\beta$ -Catenin Pathway	Melanoma	<ul style="list-style-type: none"> <li>• Murine BRAF-mutant melanoma cell lines BPD6 and D4M (BRAF<sup>V600E</sup>, PTEN<sup>-/-</sup>, syngeneic with C57BL/6)</li> <li>• C57BL/6 mice</li> </ul>	Repressed expression of Wnt5a and inhibited tumor growth.	<a href="#">Liu et al. (2018)</a>
Nano-formulation of Cromolyn		Colorectal	<ul style="list-style-type: none"> <li>• Male Wistar albino rats</li> </ul>	Enhanced anticancer effects	<a href="#">Motawi et al. (2017)</a>
PEI-PGMA nanoparticles + shRNA		Human Embryonic Kidney, breast cancer, colon cancer, leukemia	<ul style="list-style-type: none"> <li>• HEK293</li> <li>• MCF7, and MDA-MB231</li> <li>• COLO205</li> <li>• Jurkat</li> <li>• Brca2/p53 knockout mice,</li> <li>• Apc knockout model (AhCre-ErT Apc<sup>fl/fl</sup>) mice</li> </ul>	Effective targeting of c-Myc	<a href="#">Tangudu et al. (2015)</a>
Gold nanoshells + anti-FZD7 antibodies		TNBC	<ul style="list-style-type: none"> <li>• MDA-MB-231 and MCF-10A</li> </ul>	Effective inhibition of cell growth and proliferation.	<a href="#">Riley and Day (2017)</a>
IONPs		Breast	<ul style="list-style-type: none"> <li>• MDA-MB-231 cancer cell line</li> </ul>	Enhanced delivery and inhibit the growth and proliferation of cancer cells.	<a href="#">Miller-Kleinhenz et al. (2018)</a>
MSN + GSI DAPT	Notch Signalling Pathway	Breast	<ul style="list-style-type: none"> <li>• MDA-MB-231 and MCF7 cancer cell lines</li> </ul>	Achieved a significant reduction of CSCs.	<a href="#">Mamaeva et al. (2016)</a>
Fe3O4@SiO2(FITC)/PEI-FA/Notch-1 shRNA nanoparticles			<ul style="list-style-type: none"> <li>• MDA-MB-231 cancer cell line</li> </ul>	Reduced Notch-1 expression and cell growth, and induced cancer cell death.	<a href="#">Yang et al. (2014)</a>

(continued on next page)

Table 1 (continued)

Nanoformulations	Target	Type of Cancer	Samples /cell lines ( <i>In vitro</i> / <i>In vivo</i> )	Significance	Reference
DOX-miR-34a co-loaded HA-CS nanoparticles		TNBC	<ul style="list-style-type: none"> <li>MDA-MB-231 cancer cell line</li> </ul>	targeted modulation of the notch signaling pathway resulting in tumor cell death.	Deng et al. (2014)
Anthothecol-encapsulated PLGA nanoparticles	Hedgehog Signalling	Pancreas	<ul style="list-style-type: none"> <li>AsPC-1, Mia-Paca-2 and PANC-1 cancer cell lines</li> </ul>	Inhibition of pancreatic CSC.	Verma et al. (2015)
$\alpha$ -Mangostin-loaded PLGA nanoparticles	Pathway	Pancreas	<ul style="list-style-type: none"> <li>Transgenic (Kras(G12D), and Kras(G12D)/tp53R270H) mice</li> </ul>		Verma et al. (2016)
Long-circulating GlA-B-loaded oil-cored polymeric nanocapsules		Ovarian cancer, Pancreatic cancer and glioma	<ul style="list-style-type: none"> <li>SKOV3</li> <li>PANC1</li> <li>GL261</li> </ul>	Offered superior stability, cytotoxicity and bio-distribution of the drug.	Ingallina et al. (2017)
HA-SS-PLGA nanoparticles + doxorubicin and cyclopamine		Breast	<ul style="list-style-type: none"> <li>cancer cell lines</li> <li>MDA-MB-231</li> <li>MCF-7</li> </ul>	Enhanced drug efficacy	Hu et al. (2015)
Cyclopamine-paclitaxel polymer nanoparticles		Pancreas	<ul style="list-style-type: none"> <li>Cancer cell lines</li> <li>Orthotopic MiaPaca-2-luc pancreatic xenograft model</li> <li>Orthotopic PDX models</li> <li>KPC-Luc transgenic mouse model</li> </ul>	Inhibition of hedgehog signalling and enhanced therapeutic activity.	Zhao et al. (2018)
NanoHHI		Liver	<ul style="list-style-type: none"> <li>Tumor specimens from HCC patients</li> </ul>	Decreased growth and metastasis	Xu et al. (2012)
NanoHHI + gemcitabine		Pancreas	<ul style="list-style-type: none"> <li>Pa03C</li> <li>HEK293 cancer cell lines</li> </ul>	Inhibition of tumor growth and proliferation.	Chenna et al. (2012)
Polyethylenimine-Spherical nucleic acid nanoparticles		Brain	<ul style="list-style-type: none"> <li>U87-MG cancer cell line</li> </ul>	Reduced chemoresistance and stemness properties.	Melamed et al. (2018)

### 5.1.2. CD133-targeted nanotherapy

CD133 is a transmembrane glycoprotein that is a universal marker of CSCs across various types of cancers (Glumac and LeBeau, 2018; Taib et al., 2023). CD133-targeted nanoparticles are designed to selectively target and deliver therapeutic agents to cancer cells expressing the CD133 antigen on their surface. This is typically achieved by functionalizing the surface of nanoparticles with antibodies, peptides, or small molecules that have a high affinity for CD133. The specificity of these targeting ligands for CD133 is crucial for achieving selective targeting of CD133-expressing CSCs (Tian et al., 2022). Targeting CD133 for effective drug delivery has been successful in osteosarcoma, wherein a study reported that Poly (lactic-co-glycolic acid) nanoparticles loaded with CD133 aptamers successfully delivered salinomycin with high specificity to CD133 expressing CSCs, resulting in tumor cell death (Ni et al., 2015). Further, according to another study, mesoporous silica nanoparticles (MSNs) were utilized to deliver both anticancer drugs, specifically cisplatin, and short interfering RNA (siRNA) targeting the mRNA of CD133, into CD133+ cancer cells, resulting in a significant improvement in the therapeutic outcome of laryngeal carcinoma (Qi et al., 2016). Specifically, MSNs were used to deliver siRNAs against ABCG2 transport protein. Moreover, curcumin, a natural compound that possesses potent anticancer activities as evidenced in several groups of cancers (Si et al., 2023; Mishra et al., 2023; Dytrych et al., 2023), was also highly efficient in inhibiting the growth of CD133-expressing cells (Zhou et al., 2023). Therefore, studies have attempted the nanoencapsulation of curcumin to enhance its anti-tumor effects further. Such formulations are made by combining N-isopropyl acrylamide, vinylpyrrolidone, and acrylic acid in a molar ratio of 60:20:20 to encapsulate curcumin; thereby leading to the generation of "nano curcumin", which is found to be effective in targeting CSCs in the brain (Li and Zhang, 2014; Lim et al., 2011).

In a recent study, a 15-nucleotide base-pair CD133 aptamer targeting glycoprotein antigen CD133/AC133 in Glioblastoma stem cells was conjugated to gold-PEG-CB-839 to target CD133+ stem cells. It has been reported that the high affinity of gold nanoparticles to thiol groups helped in effective penetration of the blood brain barrier and biocompatibility. Moreover, the study also showed that this approach ensured

precise drug delivery to CSCs (Poonaki et al., 2022). In an earlier study, Swaminathan et al reported that polymeric nanoparticles loaded with paclitaxel and surface functionalized with anti-CD133 antibody demonstrated efficient inhibition of tumor-regrowth in vivo (Swaminathan et al., 2013).

However, there are several challenges and potential off-target effects associated with CD133-targeted nanotherapy:

- (1) Heterogeneity of CD133 Expression: CD133 is not uniformly expressed on all CSCs and its expression can vary between different types of cancer and even within the same tumor. This heterogeneity can result in incomplete targeting of CSCs and may leave some CSCs unaffected by the therapy.
- (2) Non-Specific Uptake: Nanoparticles can be taken up by cells via non-specific endocytosis, leading to the uptake of nanoparticles by non-targeted cells. This non-specific uptake can result in off-target effects and toxicity to normal cells.
- (3) Expression of CD133 on Normal Cells: CD133 is also expressed on the surface of some normal stem cells. Targeting CD133 may therefore affect normal stem cells and lead to toxicity in non-tumor tissues.
- (4) Development of Resistance: CSCs may develop resistance to CD133-targeted nanotherapy by downregulating the expression of CD133 or by activating alternative signaling pathways that promote their survival.

To address these challenges and minimize off-target effects, several strategies can be employed:

- (1) Combination Targeting: Combining CD133-targeting with other CSC markers or signaling pathways can help achieve more comprehensive targeting of CSCs and minimize the risk of resistance development.
- (2) Optimization of Nanoparticle Properties: The size, shape, and surface properties of nanoparticles can be optimized to minimize non-specific uptake and maximize selective targeting.

**Table 2**

Application of nanoparticles in cancer therapy and diagnostics and their associated limitations.

Nanoparticle Type	Applications	Drawbacks
Cancer Diagnostics		
Nanoshells	Nanoshells facilitate the conversion of electrical energy to light energy thus enabling diagnostic imaging that is devoid of the heavy-metal toxicity issues <a href="#">Nunes et al. (2019)</a> .	The application of nanoshells are limited by their large sizes <a href="#">Nunes et al. (2019)</a> .
Magnetic nanoparticles (MNPs)	Nebulization of MNPs along with Epidermal growth factor receptor (EGFR) have been used to detect the epidermal growth protein, that is commonly associated with non-small cell lung cancer (NSCLC) <a href="#">Murase et al. (2015)</a> .	High costs and potential safety issues <a href="#">Ulbrich et al. (2016)</a> .
Colloidal Gold Nanoparticles	Colloidal gold nanoparticles act as efficient contrast agents, providing enhanced diagnostic imaging owing to the permeability tension effect in tumor tissues <a href="#">Fu et al. (2018)</a> .	-
Near Infrared (NIR) Quantum Dots	NIR Quantum dots are capable of emitting fluorescence in the near-infrared spectrum in order to enhance the tracking of diagnostic devices used in solid cancers <a href="#">Parungo et al. (2004)</a> , <a href="#">Gao et al. (2004)</a> , <a href="#">Dubertret et al. (2002)</a> .	Low tissue penetration and lacks effective quantification <a href="#">Aswathy et al. (2010)</a> .
Cancer Therapeutics		
Polymeric Micelles	Polymeric micelles act as superior cancer drug carriers due to their smaller volumes and strong thermodynamic stability that allows for enhanced endothelial permeability, while also minimizing kidney rejection <a href="#">Savic et al. (2003)</a> .	Low stability in the bloodstream <a href="#">Ghezzi et al. (2021)</a> .
Liposomes	Liposomes are known to enhance the delivery of hydrophobic drugs into the intracellular spaces of tumors <a href="#">Bozzuto and Molinari (2015)</a> .	Costly methods of formulation and assembly <a href="#">Ghezzi et al. (2021)</a> .
Carbon Nanotubes	Carbon nanotubes as cancer drug carrier are particularly advantageous due to their light absorbing properties that enable them to heat up, thus allowing better penetration into tumors <a href="#">Burlaka et al. (2010)</a> .	Cytotoxicity and safety issues <a href="#">Batra et al. (2022)</a> .

- (3) Use of Alternative Targeting Ligands: Developing alternative targeting ligands with higher specificity and affinity for CD133 can help improve the selectivity of CD133-targeted nanoparticles.

CD133-targeted nanotherapy has shown promising results in various preclinical studies. For example, nanoparticles functionalized with anti-CD133 antibodies and loaded with chemotherapeutic drugs have been shown to selectively target CD133-positive cancer stem cells (CSCs) and inhibit tumor growth in mouse models of liver cancer and glioblastoma ([Poonaki et al., 2022](#)). Additionally, CD133-targeted liposomes encapsulating siRNA have been shown to effectively silence target genes in CD133-positive CSCs and inhibit tumor growth in mouse models of colorectal and liver cancer ([Wang et al., 2020](#)).

However, the translation of CD133-targeted nanotherapy from

preclinical studies to clinical trials has been challenging due to several factors:

- (1) Heterogeneity of CD133 Expression: The heterogeneous expression of CD133 among different types of cancer and even within the same tumor makes it difficult to predict the efficacy of CD133-targeted nanotherapy in a clinical setting.
- (2) Off-Target Effects: The expression of CD133 on normal stem cells raises concerns about potential off-target effects and toxicity in non-tumor tissues.
- (3) Development of Resistance: The potential for CSCs to develop resistance to CD133-targeted nanotherapy by downregulating CD133 expression or activating alternative signaling pathways is a significant concern for the long-term efficacy of this approach.

To our knowledge, there are currently no clinical trials specifically evaluating the efficacy and safety of CD133-targeted nanotherapy. However, several clinical trials have investigated the efficacy and safety of therapies targeting CD133-positive CSCs using other approaches, such as CAR-T cells or monoclonal antibodies. These studies have shown some promising results, but also highlight the challenges associated with targeting CD133-positive CSCs.

### 5.1.3. CD90-targeted nanotherapy

CD90 is a glycosylphosphatidylinositol-anchored glycoprotein observed in leukocytes and several CSCs, including breast and liver. Cells with CD90 expression are confirmed to possess elevated tumorigenicity and metastatic ability ([Yang et al., 2008](#); [Yang et al., 2008](#)). A particular report has addressed the use of CD90 as a potential therapeutic target in cancer ([Bakalova et al., 2004](#)), advocating that photosensitizer trifluoperazine loaded with anti-CD90 antibody-mediated water-soluble CdSe core nanocrystals showed effective cell death in leukemic cells.

### 5.1.4. ALDH-targeted nanotherapy

ALDH is a molecular marker of CSCs that belongs to a family of detoxifying enzymes ([Li et al., 2015](#); [Chen et al., 2014](#)). Overexpression of ALDH is observed in many cancers, such as ovarian cervical, lung, melanoma, and breast cancer ([Therachiyil et al., 2022](#); [Shao et al., 2014](#); [Liu and Zheng, 2013](#); [Vira et al., 2012](#); [Sher et al., 2022](#)). A particular study ([Dastidar et al., 2019](#)) reported the development of a novel nanoformulation of paclitaxel-encapsulated-core-shell nanoparticle of cetyl alcohol (PaxSLN) that could target aldehyde ALDH overexpressing cancer cells after oral administration. Disulfiram is a specific (DS) inhibitor of ALDH with proven cytotoxic effects on a wide range of CSCs of the colon, breast, and glioblastoma ([Hothi et al., 2012](#); [Liu et al., 2012](#); [Yip et al., 2011](#)). Disulfiram irreversibly inhibits ALDH2 by carbamylation of the catalytic Cys302 residue and thereby prevents acetaldehyde metabolism. This in turn increases circulating acetaldehyde levels in the cells, exerting a toxic effect to CSCs ([Lipsky et al., 2001](#)).

However, a major drawback of DS is its short half-life in the bloodstream. A nanoformulation created through the encapsulation of DS in poly(lactic-co-glycolic acid) (PLGA) nanoparticles (DS-PLGA) was proven to improve its half-life in the bloodstream. This formulation also showed synergistic cytotoxicity with 5-fluorouracil and sorafenib in liver cancer stem-like cells ([Wang et al., 2017](#)).

### 5.1.5. Low-density lipoprotein receptor (LDL-R) -targeted nanotherapy

LDL receptor regulates the endocytosis of cholesterol-rich LDL and helps maintain the plasma level of LDL. In the context of cancer, LDL-R is observed to have a differential expression in different cancers. Report from Yang et al states that oxidised LDL could promote stemness in bladder cancer with hypercholesterolemia ([Yang et al., 2021](#)). In this regard, a recent report showed targeting LDL-R via nanoparticle conjugated with ligands to facilitate effective drug delivery to CSCs in colon, liver and kidney cancer cells. The group successfully conjugated PCSK9



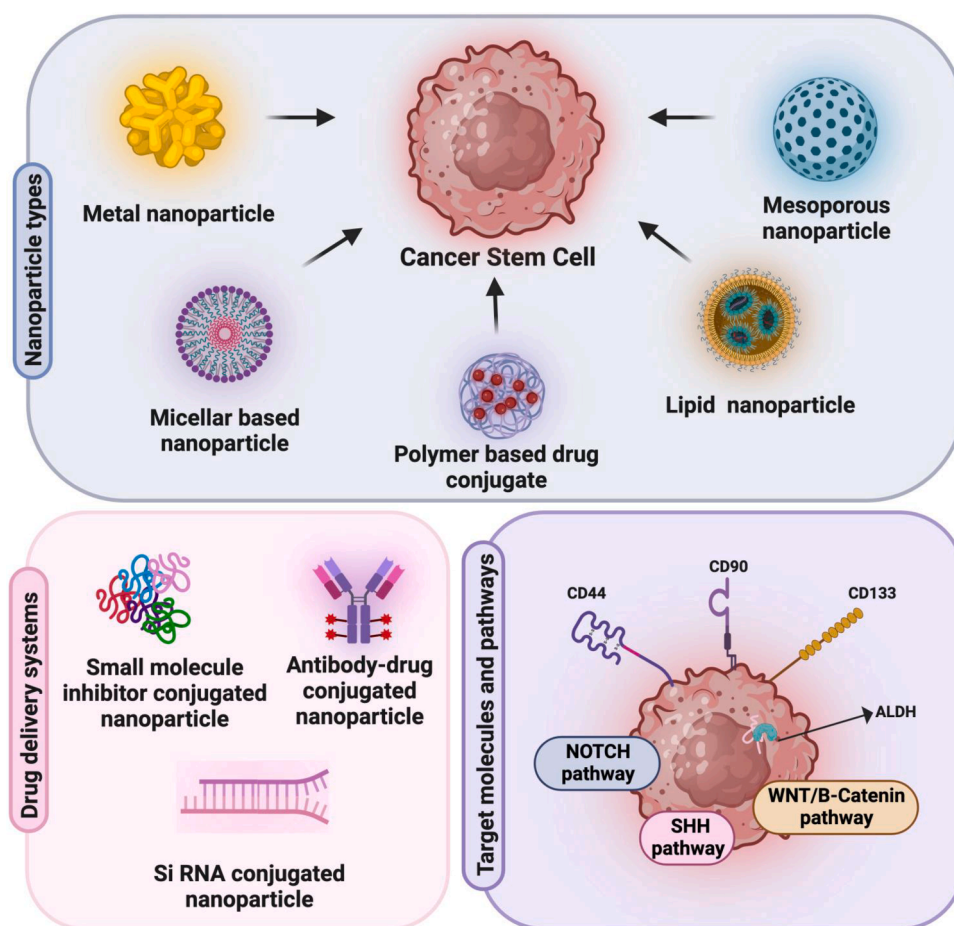


Fig. 3. Types and applications of nanomedicine in targeting cancer stem cells Created with Biorender.com).

(an LDL-R ligand) with liposomes to deliver chemotherapeutic drug paclitaxel (PTX) to cancer cells. They found that conjugated liposomes had a significantly higher growth inhibition than the unconjugated (control) liposomes in colorectal cancer cell line HCT116 (Charbe et al., 2022). Moreover, the study confirms that this novel PCSK9 conjugated liposomes has a potent and precise *in vitro* anticancer activity could be a promising targeted delivery system for cancer treatment.

## 5.2. Nanotherapy targeting CSC-associated signaling pathways

### 5.2.1. Wnt/ $\beta$ -catenin pathway

Wnt signaling is a key pathway involved in cellular development and stemness and is aberrantly upregulated in several cancers like pancreatic ductal carcinoma, breast, colon, and others (Zhan et al., 2017; Nasrolahi et al., 2023; Ram Makena et al., 2019). As a major pathway involved in conferring cellular properties of stemness, chemoresistance and metastasis, resulting in poor patient outcomes, targeting the Wnt pathway remains a promising strategy in combating tumorigenesis (Valcourt et al., 2020). The molecular inhibitors of the Wnt pathway may be naturally occurring compounds (Liu et al., 2018) or synthetic agents (Bhattacharyya et al., 2017; Motawi et al., 2017). Several studies have shown that the conjugation of Wnt inhibitors with nanoparticles successfully inhibits the pathway, thereby unleashing an anti-proliferation effect on the cancer cells. A particular study demonstrated cationic lipid-protamine nanoparticles that delivered plasmid DNA encoding a trap protein capable of binding Wnt5a, thereby repressing its expression and inhibiting tumor growth in melanoma. This study confirms that Wnt5a trapping resulted in remodeling the fibrotic immunosuppressive TME, and facilitated T cell infiltration, thereby offering a promising platform for the treatment of desmoplastic BRAF-mutant melanoma.

This is especially true when combined with a commonly accepted chemotherapy that can further lead to preparation of a tumor specific vaccine (Liu et al., 2018).

Moreover, Niclosamide (NIC) and cromolyn are synthetic Wnt inhibitors that have been successfully incorporated into nanoparticle carriers, and they act by promoting Frizzled internalization, and thereby block wnt pathway (Chen et al., 2009). In a study, the group successfully developed and preclinically tested NIC-loaded polypeptide nanoparticles that were synthesized by conjugating NIC to recombinant chimeric polypeptides (CPs) to generate CP-NIC, that behaved as a pro-drug to considerably increased the plasma exposure of NIC, thereby increasing the effect of the inhibitor. Furthermore, nanoformulating NIC with polypeptides to overcome its low solubility and bioavailability has effectively inhibited  $\beta$ -catenin levels in the cytosol. Moreover, a Cromolyn nanoparticle based-formulation has also been shown to have an anticancer effect in rats with induced colorectal cancer (Motawi et al., 2017). The study showed that cromolyn chitosan nanoparticles (CCSNPs) sustained drug release pattern over 48 h significantly reduced tumor-signaling molecules. Conclusively, CCSNOs ameliorated the whole tumor pathophysiology, enhanced the drug effect and could provide a better and novel approach for colon cancer treatment (Motawi et al., 2017).

Using nanoparticles in conjugation with siRNA or microRNA (miRNA) are also promising approach against targeting the Wnt pathway, as  $\beta$ -catenin, the major protein involved in the pathway, is practically undruggable. According to a particular report,  $\beta$ -catenin inhibition can be achieved using lipid nanoparticles (LNPs) containing dicer substrate siRNA (DsiRNA) targeting CTNBN1, the gene that encodes  $\beta$ -catenin (Ganesh et al., 2016).

Gold nanoshells are reported to be effective in delivering Frizzled7

(FZD7) antibodies and inhibiting cell growth and proliferation in triple-negative breast cancer (TNBC) (Riley and Day, 2017). The use of peptide nanoparticle compounds has also been demonstrated against Wnt signaling using ultra-small magnetic iron oxide nanoparticles (IONPs) coated with peptides that target both Wnt/LRP5/6 and uPA receptors. These were shown to successfully deliver, inhibiting the growth and proliferation of breast cancer cells (Miller-Kleinhenz et al., 2018).

### 5.2.2. Notch signaling pathway

The Notch signaling pathway is associated with pro-survival genes that regulate self-renewal and proliferation of cancer cells and determine stem cell fate (Mumm and Kopan, 2000). As a frequently deregulated pathway in cancer, Notch hyperactivation is almost always associated with carcinogenesis, thus highlighting it as a useful therapeutic target. Notch signaling is crucial for stem and progenitor cell functions in several tissues (Matsui, 2016). Moreover, Notch reduces the expression of PTEN, a tumor suppressor gene, and enhances the expression of c-Myc oncogene, thereby increasing the occurrence of cancer (Hill and Wu, 2009). Out of the several inhibitors of notch that have been extensively studied, gamma-secretase inhibitors (GSIs) that block the cleavage of the intracellular domain of notch (thus preventing its translocation to the nucleus) have been broadly investigated (Espinoza and Miele, 2013). However, as the oral administration of GSI was found to be linked to several severe gastrointestinal problems, its coating on nanoparticles considerably alleviated such issues. Interestingly, a particular study (Mamaeva et al., 2016) attempted the development of MSN to deliver a GSI, namely, N-[N-(3,5-Difluorophenacetyl)-L-alanyl]-S-phenyl glycine t-butyl Ester (DAPT) into breast cancer cell lines MDA-MB-231.

Consequently, this study thereby reported a significant reduction in the number of CSCs. In addition, polylactic acid-based nanoparticles have also been used to deliver DAPT, in combination with the EGFR-inhibitor erlotinib, to TNBC cells (Wan et al., 2019). A nanoformulation of shRNAs targeting Notch-1 has also been developed using iron oxide-silica nanoparticles for use in TNBC. This study reported that the nanoparticles successfully reduced Notch-1 expression and cell growth while also triggering cell death (Yang et al., 2014). In addition, using chitosan-based nanoparticles coupled with miRNA (miR-34) has also been demonstrated to influence the Notch signaling pathway, resulting in cell death in TNBC (Deng et al., 2014). These studies indicate that notch signaling could potentially serve as a huge target to inhibit tumor growth and chemoresistance in cancer cells.

Targeting Notch signaling pathway has a widely accepted remarkable effect in cancer therapy owing to its potential in inhibiting cell proliferation and inducing apoptotic cell death. Much of the CSC inhibiting effects of inhibiting notch pathway could be attributed to the extensive crosstalk between Notch and other cellular signaling pathways crucial for growth and development of cancer cells including Ras, Akt and NF- $\kappa$ B. Combination of Notch inhibitors with chemotherapeutic drugs have found to have a surprising anticancer effect in many cancers. Though effective, some of the Notch inhibitors when tested at clinical trials have found to have gastrointestinal toxicity. Moreover, low specificity of gamma-secretase inhibitors are also observed, though they are already used in clinical trials. Apart from the side effects and limited existing options for Notch inhibitors, there is growing optimism that Notch inhibition could be a better approach in the future therapy options in cancer (Purow, 2012).

The Notch signaling pathway interacts with various other signaling pathways, and its inhibition can have a broad range of effects on cellular processes. Some of the key pathways that interact with Notch signaling include:

- (1) Wnt/ $\beta$ -Catenin Pathway: The Notch and Wnt pathways are interconnected and can have both synergistic and antagonistic interactions. For example, Notch signaling can inhibit the Wnt

pathway by promoting the degradation of  $\beta$ -catenin, while Wnt signaling can inhibit the expression of Notch target genes.

- (2) Hedgehog Pathway: The Notch and Hedgehog pathways also have complex interactions. In some contexts, Notch signaling can inhibit Hedgehog signaling, while in others, it can promote Hedgehog pathway activation.
- (3) TGF- $\beta$ /Smad Pathway: Notch signaling can interact with the TGF- $\beta$  pathway by regulating the expression of TGF- $\beta$  ligands and receptors, as well as by modulating the activity of Smad proteins.
- (4) MAPK/ERK Pathway: Notch signaling can modulate the activity of the MAPK/ERK pathway by regulating the expression of MAPK pathway components and by interacting with Ras and Raf proteins.
- (5) PI3K/Akt/mTOR Pathway: Notch signaling can activate the PI3K/Akt/mTOR pathway by upregulating the expression of PI3K and Akt, and by promoting the phosphorylation of mTOR.

Inhibition of the Notch signaling pathway can have several potential negative impacts on these interacting pathways:

- (1) Wnt/ $\beta$ -Catenin Pathway: Inhibition of Notch signaling can lead to the activation of the Wnt pathway, which may promote tumorigenesis and metastasis in certain contexts.
- (2) Hedgehog Pathway: Inhibition of Notch signaling can lead to the activation of the Hedgehog pathway, which may promote cell proliferation and survival.
- (3) TGF- $\beta$ /Smad Pathway: Inhibition of Notch signaling can lead to the dysregulation of TGF- $\beta$  signaling, which may result in impaired cell differentiation and tissue homeostasis.
- (4) MAPK/ERK Pathway: Inhibition of Notch signaling can lead to the downregulation of the MAPK/ERK pathway, which may impair cell proliferation, survival, and migration.
- (5) PI3K/Akt/mTOR Pathway: Inhibition of Notch signaling can lead to the downregulation of the PI3K/Akt/mTOR pathway, which may impair cell metabolism and survival.

In summary, the Notch signaling pathway interacts with various other signaling pathways, and its inhibition can have a broad range of effects on cellular processes. It is important to consider these interactions and potential negative impacts when targeting the Notch pathway for cancer therapy.

### 5.2.3. Hedgehog (Hh) signaling pathway

The Hh pathway is involved mainly in cell growth and morphogenesis. Moreover, this signal transduction pathway regulates embryonic development and plays a role in the maintenance of stem cells and tissue repair (Ruiz i Altaba et al., 2002). Hh signaling has been widely associated in CSC function and maintenance (Cochrane et al., 2015). Activation of this pathway is initiated upon ligand binding to Patched 1, which drives an intracellular signaling cascade, ultimately activating three Gli transcription factors. These factors regulate several genes involved in cell proliferation, inhibition of apoptosis, metastasis, invasion, angiogenesis, and cancer stemness (Melamed et al., 2018; Rubin and de Sauvage, 2006; Pasca di Magliano and Hebrok, 2003). Aberrant activation of Hh signaling is observed in several cancers, including those of the breast, ovary, blood, prostate, and others (Kiesslich et al., 2012; Gonnissen et al., 2013; Bhateja et al., 2019).

The inhibition of Hh pathway members via antibodies, small molecules, and RNA interference has paved its way into clinical trials and are also approved as cancer treatment (Kiesslich et al., 2012). However, the poor bioavailability and resistance to these therapies have invoked the development of novel nanoparticle-based delivery systems. In this regard, antohtocol-loaded PLGA nanoparticles were observed to reduce cell proliferation and induce apoptosis in pancreatic CSCs by inhibiting Gli-DNA binding (Verma et al., 2015). Moreover, according to another report,  $\alpha$ -Mangostin-loaded PLGA nanoparticles were able to target

pancreatic CSC by disrupting Gli-DNA binding (Verma et al., 2016). Similarly, another study (Ingallina et al., 2017) also reported that glabrescione B conjugated with PLGA nanoparticles was effective in eradicating CSCs in solid tumors. In addition, the stability, *in-vitro* cytotoxicity and *in-vivo* bio-distribution of the drug were found to be comparatively much superior in the nanoformulation (Ingallina et al., 2017).

Cycloamine, a natural inhibitor of the Hh signaling pathway, is reported to have poor solubility and stability and is often linked to the occurrence of several side effects. However, the development of a nanoformulation using polymeric nanoparticles in combination with doxorubicin was found to be effective in a breast cancer model (Hu et al., 2015). Similarly, another study brought about the beneficial effect of combination therapy by using cycloamine-paclitaxel polymer nanoparticles in mice models of pancreatic and prostate cancer (Zhao et al., 2018; Yang et al., 2017). Moreover, PLGA nanoparticles formulated with the Gli1/2 antagonist, hedgehog pathway inhibitor-1 (HPI-1), are reported to impede the growth and metastasis of hepatocellular carcinoma in mice. In addition, this nanoformulation was also effective against CD133 expressing CSCs in liver cancers (Xu et al., 2012). Moreover, when combined with the anticancer drug gemcitabine, the nanoformulation successfully inhibited the growth of pancreatic cancer xenografts (Chenna et al., 2012). CSC death was also achieved by utilizing the Gli1 inhibitor GANT61 on PLGA nanoparticles, as reported by a particular study (Borah et al., 2017) demonstrating the destruction of cancer cells through the modulation of the Hh Pathway.

RNA interference using siRNAs is also extensively studied in Hh signaling inhibition, wherein most reports focus on inhibiting GLI proteins. In a particular report, Melamed et al. produced spherical nucleic acids wrapped in polyethyleneimine by conjugating GLI1 siRNA to gold nanoparticles. This nanoformulation successfully inhibited GLI1 and its downstream effectors (Melamed et al., 2018). Moreover, the use of a non-viral gene delivery vector has also been demonstrated in conjugation with a nanoparticle platform that further enhanced its efficacy. Here, the nano-drug delivery system was developed through the conjugation of siGLI1 with 1,2-dioleoyl-3-triethylammonium-propane (DOTAP)-conjugated methoxy-PEG-poly(lactide) copolymer that was able to induce cell death and inhibit cell growth of subcutaneous glioblastoma xenografts *in-vivo* (Zhou et al., 2018).

#### Challenges associated with targeting CSCs in cancer treatment

Targeting Cancer Stem Cells (CSCs) in cancer treatment is a promising strategy to enhance treatment efficacy and sensitize cells to therapy. However, this approach presents several challenges that need to be addressed to maximize its therapeutic potential.

- (1) Heterogeneity of CSCs: CSCs are highly heterogeneous, both within and between tumors. This heterogeneity is reflected in the expression of surface markers, genetic mutations, and signaling pathways, which can vary significantly among CSCs from different tumors (inter-tumoral heterogeneity) or even within the same tumor (intra-tumoral heterogeneity) (Botchkina et al., 2009). This heterogeneity can interfere with the selective targeting of specific CSC sub-populations and may also affect signaling pathways or the tumor microenvironment that might not be the real targets for cancer treatment. Additionally, this heterogeneity can lead to the development of resistance to targeted therapies, as sub-populations of CSCs that are not effectively targeted may survive and contribute to tumor recurrence.
- (2) Complexity of the Tumor Microenvironment: The tumor microenvironment is a complex network of various cell types, extracellular matrix components, and signaling molecules that can influence the behavior of CSCs. For example, interactions with stromal cells, immune cells, or extracellular matrix components can promote the survival, self-renewal, and invasion capabilities of CSCs. Additionally, the tumor microenvironment can also

influence the accessibility of therapeutic agents to CSCs, thereby affecting the efficacy of targeted therapies.

- (3) Molecular Drivers of CSCs: CSCs are regulated by multiple molecular drivers that contribute to their self-renewal, differentiation, and survival. These molecular drivers can include signaling pathways (e.g., Wnt/ $\beta$ -catenin, Notch, Sonic hedgehog), transcription factors (e.g., Sox2, Oct4, Nanog), and epigenetic regulators (e.g., histone deacetylases, DNA methyltransferases). Targeting these molecular drivers is crucial for the effective inhibition of CSCs. However, given the complexity and redundancy of these regulatory networks, it is often challenging to identify and target the key drivers that are essential for CSC maintenance and function.
- (4) Combination Therapies: Given the challenges associated with targeting CSCs, a more effective approach may be to combine therapies that target the molecular drivers of CSCs with standard chemotherapeutic treatments. This combination approach can help overcome the limitations of single-agent therapies, enhance the efficacy of treatment, and minimize the risk of resistance development (Turdo et al., 2019).

In summary, targeting CSCs in cancer treatment presents several challenges related to the heterogeneity of CSCs, the complexity of the tumor microenvironment, and the identification and targeting of key molecular drivers. Addressing these challenges will be crucial for the development of more effective therapeutic approaches that specifically target CSCs and yield better therapy outcomes.

## 6. Conclusion and future perspective

In conclusion, our understanding of cancer and its mechanisms has come a long way, yet the disease remains a formidable adversary, partly due to the limitations of existing treatment modalities. Conventional chemotherapy, the standard tool in our armamentarium, has been mired with issues such as cytotoxicity, lack of selectivity, and induction of multi-drug resistance. These limitations inevitably give rise to an urgent need for alternative therapeutic strategies. The application of nanotechnology in cancer treatment has begun to fulfill this need, offering promising breakthroughs and bringing a new level of sophistication to the treatment landscape.

Nanomaterials, with their unique properties, have demonstrated immense potential for enhancing the delivery of anticancer drugs to the target cells, reducing off-target effects, and specifically targeting CSCs. Clinical studies exploring these nanoformulations have shown promising results, highlighting their potential to become a cornerstone in cancer therapeutics. Yet, despite these strides, the approval of nano-drugs has remained relatively low, reminding us of the challenges that persist. As we advance, researchers and clinicians alike must channel their efforts towards mitigating these challenges. To increase the success rate of nano-drug approval, the focus needs to be on reducing the toxicity of nanomaterials, refining their permeability and retention for optimal drug delivery, and minimizing the protein corona's shielding effect.

Moreover, there is a need to deepen our understanding of cancer stem cell biology and their interactions with nanomaterials, as this can pave the way for developing more specific and effective nano-based therapeutic strategies. In addition, expanding clinical trials of nano-therapeutics, including those targeting CSCs, is crucial for validating their safety and efficacy in a diverse population. The promising future of nanotechnology in cancer treatment is in sight, yet it demands concerted efforts and relentless research. With the necessary commitment and resource allocation, we believe that nanotechnology could significantly change the prognosis for patients battling this deadly disease, truly revolutionizing the field of oncology.

**Ethical approval**

he study does not require ethical approval

**Consent to participate**

Not applicable

**Consent for publication**

Not applicable

**Availability of data and material**

Not applicable

**Funding**

None

**CRedit authorship contribution statement**

**Queenie Fernandes:** Writing – original draft, Project administration. **Lubna Therachiyil:** Conceptualization, Writing – original draft. **Abdul Q. Khan:** Conceptualization, Writing – original draft. **Takwa Bedhafi:** Writing – review & editing. **Hesham M Korashy:** Writing – review & editing. **Ajaz A. Bhat:** Conceptualization, Writing – review & editing. **Shahab Uddin:** Supervision, Writing – review & editing, Writing – original draft.

**Declaration of Competing Interest**

The authors declare no conflict of interest

**Data availability**

No data was used for the research described in the article.

**Acknowledgments**

The authors acknowledge the Qatar National Library to support open-access publication charges.

**References**

- Laconi, E., Marongiu, F., DeGregori, J., 2020. Cancer as a disease of old age: changing mutational and microenvironmental landscapes. *Br. J. Cancer* 122 (7), 943–952.
- Kaminska, M., Ciszewski, T., Lopacka-Szatan, K., Miotla, P., Staroslawska, E., 2015. Breast cancer risk factors. *Prz. Menopauzalny* 14 (3), 196–202.
- Siegel, R.L., Miller, K.D., Wagle, N.S., Jemal, A., 2023. Cancer statistics, 2023. *CA Cancer J. Clin.* 73 (1), 17–48.
- Song, M., Giovannucci, E., 2016. Preventable incidence and mortality of carcinoma associated with lifestyle factors among white adults in the United States. *JAMA Oncol.* 2 (9), 1154–1161.
- Vineis, P., Wild, C.P., 2014. Global cancer patterns: causes and prevention. *Lancet* 383 (9916), 549–557.
- Wang, J.C., Dick, J.E., 2005. Cancer stem cells: lessons from leukemia. *Trends Cell Biol.* 15 (9), 494–501.
- Koren, E., Fuchs, Y., 2016. The bad seed: Cancer stem cells in tumor development and resistance. *Drug Resist. Updat.* 28, 1–12.
- Zhu, P., Fan, Z., 2018. Cancer stem cells and tumorigenesis. *Biophys. Rep.* 4 (4), 178–188.
- Ayob, A.Z., Ramasamy, T.S., 2018. Cancer stem cells as key drivers of tumour progression. *J. Biomed. Sci.* 25 (1), 20.
- Yu, Z., Pestell, T.G., Lisanti, M.P., Pestell, R.G., 2012. Cancer stem cells. *Int. J. Biochem. Cell Biol.* 44 (12), 2144–2151.
- Polyak, K., Hahn, W.C., 2006. Roots and stems: stem cells in cancer. *Nat. Med.* 12 (3), 296–300.
- Jordan, C.T., 2005. Targeting the most critical cells: approaching leukemia therapy as a problem in stem cell biology. *Nat. Clin. Pract. Oncol.* 2 (5), 224–225.

- Xia, Y., Xiong, Y., Lim, B., Skrabalak, S.E., 2009. Shape-controlled synthesis of metal nanocrystals: simple chemistry meets complex physics? *Angew. Chem. Int. Ed. Engl.* 48 (1), 60–103.
- Peer, D., Karp, J.M., Hong, S., Farokhzad, O.C., Margalit, R., Langer, R., 2007. Nanocarriers as an emerging platform for cancer therapy. *Nat. Nanotechnol.* 2 (12), 751–760.
- Bray, F., Laversanne, M., Weiderpass, E., Soerjomataram, I., 2021. The ever-increasing importance of cancer as a leading cause of premature death worldwide. *Cancer* 127 (16), 3029–3030.
- Roy, P.S., Saikia, B.J., 2016. Cancer and cure: a critical analysis. *Indian J. Cancer* 53 (3), 441–442.
- Omran, A.R., 2005. The epidemiologic transition: a theory of the epidemiology of population change. 1971. *Milbank Q.* 83 (4), 731–757.
- Gansler, T., Ganz, P.A., Grant, M., Greene, F.L., Johnstone, P., Mahoney, M., Newman, L. A., Oh, W.K., Thomas Jr., C.R., Thun, M.J., Vickers, A.J., Wender, R.C., Brawley, O. W., 2010. Sixty years of CA: a cancer journal for clinicians. *CA Cancer J. Clin.* 60 (6), 345–350.
- Mattiuzzi, C., Lippi, G., 2019. Current cancer epidemiology. *J. Epidemiol. Glob. Health* 9 (4), 217–222.
- Sung, H., Ferlay, J., Siegel, R.L., Laversanne, M., Soerjomataram, I., Jemal, A., Bray, F., 2021. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J. Clin.* 71 (3), 209–249.
- Thun, M., Peto, R., Boreham, J., Lopez, A.D., 2012. Stages of the cigarette epidemic on entering its second century. *Tob. Control.* 21 (2), 96–101.
- Mu, L., Liu, L., Niu, R., Zhao, B., Shi, J., Li, Y., Swanson, M., Scheider, W., Su, J., Chang, S.C., Yu, S., Zhang, Z.F., 2013. Indoor air pollution and risk of lung cancer among Chinese female non-smokers. *Cancer Causes Control.* 24 (3), 439–450.
- Turner, M.C., Andersen, Z.J., Baccarelli, A., Diver, W.R., Gapstur, S.M., Pope 3rd, C.A., Prada, D., Samet, J., Thurston, G., Cohen, A., 2020. Outdoor air pollution and cancer: An overview of the current evidence and public health recommendations. *CA Cancer J. Clin.*
- Lin, B., Liu, X., Lu, W., Wu, X., Li, Y., Zhang, Z., Fu, R., Zhang, L., Xiong, J., 2023. Prevalence and associated factors of smoking among chinese adolescents: a school-based cross-sectional study. *BMC Public Health* 23 (1), 669.
- Flor, L.S., Reitsma, M.B., Gupta, V., Ng, M., Gakidou, E., 2021. The effects of tobacco control policies on global smoking prevalence. *Nat. Med.* 27 (2), 239–243.
- Wilson, L.M., Avila Tang, E., Chander, G., Hutton, H.E., Odelola, O.A., Elf, J.L., Heckman-Stoddard, B.M., Bass, E.B., Little, E.A., Haberl, E.B., Apelberg, B.J., 2012. Impact of tobacco control interventions on smoking initiation, cessation, and prevalence: a systematic review. *J. Environ. Public Health* 2012, 961724.
- Bray F., Soerjomataram I., The Changing Global Burden of Cancer: Transitions in Human Development and Implications for Cancer Prevention and Control. In: Gelband H, Jha P, Sankaranarayanan R, Horton S, editors. *Cancer: Disease Control Priorities, Third Edition (Volume 3)*. Washington (DC): The International Bank for Reconstruction and Development / The World Bank; 2015. Chapter 2.
- Fidler, M.M., Soerjomataram, I., Bray, F., 2016. A global view on cancer incidence and national levels of the human development index. *Int. J. Cancer* 139 (11), 2436–2446.
- Siegel, R.L., Miller, K.D., Goding Sauer, A., Fedewa, S.A., Butterly, L.F., Anderson, J.C., Cercek, A., Smith, R.A., Jemal, A., 2020. Colorectal cancer statistics, 2020. *CA Cancer J. Clin.* 70 (3), 145–164.
- Sullivan T., Sullivan R., Ginsburg O.M., Screening for Cancer: Considerations for Low-and Middle-Income Countries. In: Gelband H, Jha P, Sankaranarayanan R, Horton S, editors. *Cancer: Disease Control Priorities, Third Edition (Volume 3)*. Washington (DC): The International Bank for Reconstruction and Development / The World Bank; 2015. Chapter 12.
- Arnold, M., Abnet, C.C., Neale, R.E., Vignat, J., Giovannucci, E.L., McGlynn, K.A., Bray, F., 2020. Global burden of 5 major types of gastrointestinal cancer. *Gastroenterology* 159 (1), 335–349 e15.
- Edwards, B.K., Ward, E., Kohler, B.A., Ehemann, C., Zauber, A.G., Anderson, R.N., Jemal, A., Schymura, M.J., Lansdorf-Vogelaar, I., Seeff, L.C., van Ballegooijen, M., Goede, S.L., Ries, L.A., 2010. Annual report to the nation on the status of cancer, 1975–2006, featuring colorectal cancer trends and impact of interventions (risk factors, screening, and treatment) to reduce future rates. *Cancer* 116 (3), 544–573.
- Soltysova, A., Altanerova, V., Altaner, C., 2005. Cancer stem cells. *Neoplasma* 52 (6), 435–440.
- Zhou, H., Tan, L., Liu, B., Guan, X.Y., 2023. Cancer stem cells: Recent insights and therapies. *Biochem. Pharmacol.* 209, 115441.
- Yilmaz, M., Kaplan, F., Mender, I., Gryaznov, S.M., Dikmen, Z.G., 2023. Cancer stem cells and anti-tumor immunity. *Curr. Stem. Cell Res. Ther.* 18 (4), 445–459.
- Palomeras, S., Ruiz-Martinez, S., Puig, T., 2018. Targeting breast cancer stem cells to overcome treatment resistance. *Molecules* 23 (9).
- Sahai, E., 2005. Mechanisms of cancer cell invasion. *Curr. Opin. Genet. Dev.* 15 (1), 87–96.
- Brasseur, K., Gevry, N., Asselin, E., 2017. Chemoresistance and targeted therapies in ovarian and endometrial cancers. *Oncotarget* 8 (3), 4008–4042.
- Cho, R.W., Clarke, M.F., 2008. Recent advances in cancer stem cells. *Curr. Opin. Genet. Dev.* 18 (1), 48–53.
- Tefas, L.R., Barbalata, C., Tefas, C., Tomuta, I., 2021. Salinomycin-based drug delivery systems: overcoming the hurdles in cancer therapy. *Pharmaceutics* 13 (8).
- Essex, A., Pineda, J., Acharya, G., Xin, H., Evans, J., 2019. B. reproducibility project: cancer, replication study: wnt activity defines colon cancer stem cells and is regulated by the microenvironment. *eLife* 8.
- Borah, A., Raveendran, S., Rochani, A., Maekawa, T., Kumar, D.S., 2015. Targeting self-renewal pathways in cancer stem cells: clinical implications for cancer therapy. *Oncogenesis* 4 (11), e177.

- Farnie, G., Clarke, R.B., 2007. Mammary stem cells and breast cancer—role of Notch signalling. *Stem Cell Rev.* 3 (2), 169–175.
- Clement, V., Sanchez, P., de Tribolet, N., Radovanovic, I., Ruiz i Altaba, A., 2007. HEDGEHOG-Gli1 signaling regulates human glioma growth, cancer stem cell self-renewal, and tumorigenicity. *Curr. Biol.* 17 (2), 165–172.
- Yang, L., Shi, P., Zhao, G., Xu, J., Peng, W., Zhang, J., Zhang, G., Wang, X., Dong, Z., Chen, F., Cui, H., 2020. Targeting cancer stem cell pathways for cancer therapy. *Signal Transduct. Target. Ther.* 5 (1), 8.
- Ajani, J.A., Song, S., Hochster, H.S., Steinberg, I.B., 2015. Cancer stem cells: the promise and the potential. *Semin. Oncol.* 42 (S3–17), Suppl 1.
- Kroon, P., Berry, P.A., Stower, M.J., Rodrigues, G., Mann, V.M., Simms, M., Bhasin, D., Chettiar, S., Li, C., Li, P.K., Maitland, N.J., Collins, A.T., 2013. JAK-STAT blockade inhibits tumor initiation and clonogenic recovery of prostate cancer stem-like cells. *Cancer Res.* 73 (16), 5288–5298.
- Plaks, V., Kong, N., Werb, Z., 2015. The cancer stem cell niche: how essential is the niche in regulating stemness of tumor cells? *Cell Stem Cell* 16 (3), 225–238.
- Abbaszadegan, M.R., Bagheri, V., Razavi, M.S., Momtazi, A.A., Sahebkar, A., Gholamin, M., 2017. Isolation, identification, and characterization of cancer stem cells: A review. *J. Cell. Physiol.* 232 (8), 2008–2018.
- Kukul, S., Guin, D., Rawat, C., Bora, S., Mishra, M.K., Sharma, P., Paul, P.R., Kanojia, N., Grewal, G.K., Kukreti, S., Saso, L., Kukreti, R., 2021. Multidrug efflux transporter ABCG2: expression and regulation. *Cell. Mol. Life Sci.* 78 (21–22), 6887–6939.
- Abdullah, L.N., Chow, E.K., 2013. Mechanisms of chemoresistance in cancer stem cells. *Clin. Transl. Med.* 2 (1), 3.
- Olcina, M., Lecane, P.S., Hammond, E.M., 2010. Targeting hypoxic cells through the DNA damage response. *Clin. Cancer Res.* 16 (23), 5624–5629.
- Quintana, E., Shackleton, M., Sabel, M.S., Fullen, D.R., Johnson, T.M., Morrison, S.J., 2008. Efficient tumour formation by single human melanoma cells. *Nature* 456 (7222), 593–598.
- Hosen, N., Park, C.Y., Tatsumi, N., Oji, Y., Sugiyama, H., Gramatzki, M., Krensky, A.M., Weissman, I.L., 2007. CD96 is a leukemic stem cell-specific marker in human acute myeloid leukemia. *Proc. Natl. Acad. Sci. U. S. A.* 104 (26), 11008–11013.
- Zhao, W., Li, Y., Zhang, X., 2017. Stemness-related markers in cancer. *Cancer Transl. Med.* 3 (3), 87–95.
- Eun, K., Ham, S.W., Kim, H., 2017. Cancer stem cell heterogeneity: origin and new perspectives on CSC targeting. *BMB Rep.* 50 (3), 117–125.
- Hejmady, S., Pradhan, R., Alexander, A., Agrawal, M., Singhvi, G., Gorain, B., Tiwari, S., Kesharwani, P., Dubey, S.K., 2020. Recent advances in targeted nanomedicine as promising antitumor therapeutics. *Drug Discov Today* 25 (12), 2227–2244.
- Hanahan, D., Coussens, L.M., 2012. Accessories to the crime: functions of cells recruited to the tumor microenvironment. *Cancer Cell* 21 (3), 309–322.
- Albini, A., Bruno, A., Gallo, C., Pajardi, G., Noonan, D.M., Dallaglio, K., 2015. Cancer stem cells and the tumor microenvironment: interplay in tumor heterogeneity. *Connect. Tissue Res.* 56 (5), 414–425.
- Thankamony, A.P., Saxena, K., Murali, R., Jolly, M.K., Nair, R., 2020. Cancer stem cell plasticity - a deadly deal. *Front. Mol. Biosci.* 7, 79.
- Tredan, O., Galmarini, C.M., Patel, K., Tannock, I.F., 2007. Drug resistance and the solid tumor microenvironment. *J. Natl. Cancer Inst.* 99 (19), 1441–1454.
- Halder, J., Pradhan, D., Kar, B., Ghosh, G., Rath, G., 2022. Nanotherapeutics approaches to overcome P-glycoprotein-mediated multi-drug resistance in cancer. *Nanomedicine* 40, 102494.
- Bruno, E., Manacorda, M., Verga Loiaconi, E., Villa, G., 1987. [Hyperbaric oxygen therapy in the therapy of mandibular osteoradionecrosis]. *Dent. Cadmos* 55 (6), 67–75.
- Ediriwickrema, A., Saltzman, W.M., 2015. Nanotherapy for cancer: targeting and multifunctionality in the future of cancer therapies. *ACS Biomater. Sci. Eng.* 1 (2), 64–78.
- McCarthy, T.D., Karellas, P., Henderson, S.A., Giannis, M., O'Keefe, D.F., Heery, G., Paull, J.R., Matthews, B.R., Holan, G., 2005. Dendrimers as drugs: discovery and preclinical and clinical development of dendrimer-based microbicides for HIV and STI prevention. *Mol. Pharm.* 2 (4), 312–318.
- Brigger, I., Dubernet, C., Couvreur, P., 2002. Nanoparticles in cancer therapy and diagnosis. *Adv. Drug. Deliv. Rev.* 54 (5), 631–651.
- Davis, M.E., Zuckerman, J.E., Choi, C.H., Seligson, D., Tolcher, A., Alabi, C.A., Yen, Y., Heidel, J.D., Ribas, A., 2010. Evidence of RNAi in humans from systemically administered siRNA via targeted nanoparticles. *Nature* 464 (7291), 1067–1070.
- Mendes, B.B., Sousa, D.P., Connot, J., Conde, J., 2021. Nanomedicine-based strategies to target and modulate the tumor microenvironment. *Trends Cancer* 7 (9), 847–862.
- Bedhiafi, T., Idoudi, S., Alhams, A.A., Fernandes, Q., Iqbal, H., Basinani, R., Uddin, S., Dermime, S., Merhi, M., Billa, N., 2023. Applications of polydopaminic nanomaterials in mucosal drug delivery. *J. Control. Release* 353, 842–849.
- Bedhiafi, T., Idoudi, S., Fernandes, Q., Al-Zaidan, L., Uddin, S., Dermime, S., Billa, N., Merhi, M., 2023. Nano-vitamin C: a promising candidate for therapeutic applications. *Biomed. Pharmacother.* 158, 114093.
- Gindy, M.E., Prud'homme, R.K., 2009. Multifunctional nanoparticles for imaging, delivery and targeting in cancer therapy. *Expert Opin. Drug Deliv.* 6 (8), 865–878.
- Kim, B.Y., Rutka, J.T., Chan, W.C., 2010. Nanomedicine. *N. Engl. J. Med.* 363 (25), 2434–2443.
- Hobbs, S.K., Monsky, W.L., Yuan, F., Roberts, W.G., Griffith, L., Torchilin, V.P., Jain, R. K., 1998. Regulation of transport pathways in tumor vessels: role of tumor type and microenvironment. *Proc. Natl. Acad. Sci. U. S. A.* 95 (8), 4607–4612.
- Prencipe, G., Tabakman, S.M., Welsher, K., Liu, Z., Goodwin, A.P., Zhang, L., Henry, J., Dai, H., 2009. PEG branched polymer for functionalization of nanomaterials with ultralong blood circulation. *J. Am. Chem. Soc.* 131 (13), 4783–4787.
- Alexis, F., Pridgen, E., Molnar, L.K., Farokhzad, O.C., 2008. Factors affecting the clearance and biodistribution of polymeric nanoparticles. *Mol. Pharm.* 5 (4), 505–515.
- Xu, M., Han, X., Xiong, H., Gao, Y., Xu, B., Zhu, G., Li, J., 2023. Cancer nanomedicine: emerging strategies and therapeutic potentials. *Molecules* 28 (13), 5145.
- Libutti, S.K., Paciotti, G.F., Byrnes, A.A., Alexander Jr., H.R., Gannon, W.E., Walker, M., Seidel, G.D., Yuldasheva, N., Tamarkin, L., 2010. Phase I and pharmacokinetic studies of CYT-6091, a novel PEGylated colloidal gold-rhTNF nanomedicine. *Clin. Cancer Res.* 16 (24), 6139–6149.
- Visaria, R.K., Griffin, R.J., Williams, B.W., Ebbini, E.S., Paciotti, G.F., Song, C.W., Bischof, J.C., 2006. Enhancement of tumor thermal therapy using gold nanoparticle-assisted tumor necrosis factor-alpha delivery. *Mol. Cancer Ther.* 5 (4), 1014–1020.
- Kim, T.Y., Kim, D.W., Chung, J.Y., Shin, S.G., Kim, S.C., Heo, D.S., Kim, N.K., Bang, Y.J., 2004. Phase I and pharmacokinetic study of Genexol-PM, a cremophor-free, polymeric micelle-formulated paclitaxel, in patients with advanced malignancies. *Clin. Cancer Res.* 10 (11), 3708–3716.
- Sun, X., Zhao, P., Lin, J., Chen, K., Shen, J., 2023. Recent advances in access to overcome cancer drug resistance by nanocarrier drug delivery system. *Cancer Drug Resist.* 6 (2), 390–415.
- Han, Y., Wen, P., Li, J., Kataoka, K., 2022. Targeted nanomedicine in cisplatin-based cancer therapeutics. *J. Control. Release* 345, 709–720.
- Ghosh, S., Javia, A., Shetty, S., Bardoliwala, D., Maiti, K., Banerjee, S., Khopade, A., Misra, A., Sawant, K., Bhowmick, S., 2021. Triple negative breast cancer and non-small cell lung cancer: Clinical challenges and nano-formulation approaches. *J. Control. Release* 337, 27–58.
- Araujo, F., Shrestha, N., Granja, P.L., Hirvonen, J., Santos, H.A., Sarmiento, B., 2015. Safety and toxicity concerns of orally delivered nanoparticles as drug carriers. *Expert Opin. Drug Metab. Toxicol.* 11 (3), 381–393.
- Arora, S., Rajwade, J.M., Paknikar, K.M., 2012. Nanotoxicology and *in vitro* studies: the need of the hour. *Toxicol. Appl. Pharmacol.* 258 (2), 151–165.
- Baldric, P., 2010. The safety of chitosan as a pharmaceutical excipient. *Regul. Toxicol. Pharmacol.* 56 (3), 290–299.
- Bar-Zeev, M., Livnev, Y.D., Assaraf, Y.G., 2017. Targeted nanomedicine for cancer therapeutics: towards precision medicine overcoming drug resistance. *Drug Resist. Update* 31, 15–30.
- Harisinghani, M.G., Barentsz, J., Hahn, P.F., Deserno, W.M., Tabatabaei, S., van de Kaa, C.H., de la Rosette, J., Weisleder, R., 2003. Noninvasive detection of clinically occult lymph-node metastases in prostate cancer. *N. Engl. J. Med.* 348 (25), 2491–2499.
- Enochs, W.S., Harsh, G., Hochberg, F., Weissleder, R., 1999. Improved delineation of human brain tumors on MR images using a long-circulating, superparamagnetic iron oxide agent. *J. Magn. Reson. Imaging* 9 (2), 228–232.
- Perrault, S.D., Walkey, C., Jennings, T., Fischer, H.C., Chan, W.C., 2009. Mediating tumor targeting efficiency of nanoparticles through design. *Nano Lett.* 9 (5), 1909–1915.
- Li, L., Jiang, W., Luo, K., Song, H., Lan, F., Wu, Y., Gu, Z., 2013. Superparamagnetic iron oxide nanoparticles as MRI contrast agents for non-invasive stem cell labeling and tracking. *Theranostics* 3 (8), 595–615.
- Cormode, D.P., Naha, P.C., Fayad, Z.A., 2014. Nanoparticle contrast agents for computed tomography: a focus on micelles. *Contrast Media Mol. Imaging* 9 (1), 37–52.
- Garbayo, E., Pascual-Gil, S., Rodriguez-Nogales, C., Saludas, L., Estella-Hermoso de Mendoza, A., Blanco-Prieto, M.J., 2020. Nanomedicine and drug delivery systems in cancer and regenerative medicine. *Wiley Interdiscip. Rev. Nanomed. Nanobiotechnol.* 12 (5), e1637.
- Lyakhovich, A., Leonart, M.E., 2016. Bypassing mechanisms of mitochondria-mediated cancer stem cells resistance to chemo- and radiotherapy. *Oxidative Med. Cell. Longev.* 2016, 1716341.
- Senbanjo, L.T., Chellaiah, M.A., 2017. CD44: a multifunctional cell surface adhesion receptor is a regulator of progression and metastasis of cancer cells. *Front. Cell Dev. Biol.* 5, 18.
- Brown, R.L., Reinke, L.M., Damerow, M.S., Perez, D., Chodosh, L.A., Yang, J., Cheng, C., 2011. CD44 splice isoform switching in human and mouse epithelium is essential for epithelial-mesenchymal transition and breast cancer progression. *J. Clin. Invest.* 121 (3), 1064–1074.
- Su, Y.J., Lai, H.M., Chang, Y.W., Chen, G.Y., Lee, J.L., 2011. Direct reprogramming of stem cell properties in colon cancer cells by CD44. *EMBO J.* 30 (15), 3186–3199.
- Dubey, P., Gupta, R., Mishra, A., Kumar, V., Bhaduria, S., Bhatt, M.L.B., 2022. Evaluation of correlation between CD44, radiotherapy response, and survival rate in patients with advanced stage of head and neck squamous cell carcinoma (HNSCC). *Cancer Med.* 11 (9), 1937–1947.
- Martincuks, A., Li, P.C., Zhao, Q., Zhang, C., Li, Y.J., Yu, H., Rodriguez-Rodriguez, L., 2020. CD44 in ovarian cancer progression and therapy resistance—a critical role for STAT3. *Front. Oncol.* 10, 589601.
- Al-Hajj, M., Wicha, M.S., Benito-Hernandez, A., Morrison, S.J., Clarke, M.F., 2003. Prospective identification of tumorigenic breast cancer cells. *Proc. Natl. Acad. Sci. U. S. A.* 100 (7), 3983–3988.
- Gener, P., Gouveia, L.P., Sabat, G.R., de Sousa Rafael, D.F., Fort, N.B., Arranja, A., Fernandez, Y., Prieto, R.M., Ortega, J.S., Arango, D., Abasolo, I., Videira, M., Schwartz Jr., S., 2015. Fluorescent CSC models evidence that targeted nanomedicines improve treatment sensitivity of breast and colon cancer stem cells. *Nanomedicine* 11 (8), 1883–1892.
- Xu, H., Niu, M., Yuan, X., Wu, K., Liu, A., 2020. CD44 as a tumor biomarker and therapeutic target. *Exp. Hematol. Oncol.* 9 (1), 36.
- Aires, A., Ocampo, S.M., Simoes, B.M., Rodriguez, M.J., Cadenas, J.F., Couleaud, P., Spence, K., Latorre, A., Miranda, R., Somoza, A., Clarke, R.B., Carrascosa, J.L.,

- Cortajarena, A.L., 2016. Multifunctionalized iron oxide nanoparticles for selective drug delivery to CD44-positive cancer cells. *Nanotechnology* 27 (6), 065103.
- Arabi, L., Badiee, A., Mosaffa, F., Jaafari, M.R., 2015. Targeting CD44 expressing cancer cells with anti-CD44 monoclonal antibody improves cellular uptake and antitumor efficacy of liposomal doxorubicin. *J. Control. Release* 220 (Pt A), 275–286.
- Qiu, J., Cheng, R., Zhang, J., Sun, H., Deng, C., Meng, F., Zhong, Z., 2017. Glutathione-sensitive hyaluronic acid-mercaptopurine prodrug linked via carbonyl vinyl sulfide: a robust and CD44-targeted nanomedicine for leukemia. *Biomacromolecules* 18 (10), 3207–3214.
- Chen, C., Zhao, S., Karnad, A., Freeman, J.W., 2018. The biology and role of CD44 in cancer progression: therapeutic implications. *J. Hematol. Oncol.* 11 (1), 64.
- Vinogradov, S., Wei, X., 2012. Cancer stem cells and drug resistance: the potential of nanomedicine. *Nanomedicine* 7 (4), 597–615 (Lond).
- Hassn Mesrati, M., Syafruddin, S.E., Mohtar, M.A., Syahir, A., 2021. CD44: a multifunctional mediator of cancer progression. *Biomolecules* 11 (12), 1850.
- Glumac, P.M., LeBeau, A.M., 2018. The role of CD133 in cancer: a concise review. *Clin. Transl. Med.* 7 (1), 18.
- Taib, N., Merhi, M., Inchakalody, V., Mestiri, S., Hydrose, S., Makni-Maalej, K., Raza, A., Sahir, F., Azizi, F., Nizamuddin, P.B., Fernandes, Q., Yoosuf, Z., Almoghrabi, S., Al-Zaidan, L., Shablak, A., Uddin, S., Maccalli, C., Al Homsy, M.U., Dermime, S., 2023. Treatment with decitabine induces the expression of stemness markers, PD-L1 and NY-ESO-1 in colorectal cancer: potential for combined chemoimmunotherapy. *J. Transl. Med.* 21 (1), 235.
- Tian, H., Zhang, T., Qin, S., Huang, Z., Zhou, L., Shi, J., Nice, E.C., Xie, N., Huang, C., Shen, Z., 2022. Enhancing the therapeutic efficacy of nanoparticles for cancer treatment using versatile targeted strategies. *J. Hematol. Oncol.* 15 (1), 132.
- Ni, M., Xiong, M., Zhang, X., Cai, G., Chen, H., Zeng, Q., Yu, Z., 2015. Poly(lactic-co-glycolic acid) nanoparticles conjugated with CD133 aptamers for targeted salinomycin delivery to CD133<sup>+</sup> osteosarcoma cancer stem cells. *Int. J. Nanomed.* 10, 2537–2554.
- Qi, X., Yu, D., Jia, B., Jin, C., Liu, X., Zhao, X., Zhang, G., 2016. Targeting CD133<sup>+</sup> laryngeal carcinoma cells with chemotherapeutic drugs and siRNA against ABCG2 mediated by thermo/pH-sensitive mesoporous silica nanoparticles. *Tumour Biol.* 37 (2), 2209–2217.
- Si, L., Zhang, L., Xing, S., Fang, P., Tian, X., Liu, X., Xv, X., 2023. Curcumin as a therapeutic agent in cancer therapy: Focusing on its modulatory effects on circular RNAs. *Phytother. Res.*
- Yang, S.J., Pai, J.A., Yao, C.J., et al., 2023. SN38-loaded nanomedicine mediates chemoradiotherapy against CD44-expressing cancer growth. *Cancer Nano* 14, 1–20.
- Mishra, A.P., Swetanshu, P.S., Yadav, S., Nigam, M., Seidel, V., Rodrigues, C.F., 2023. Role of the dietary phytochemical curcumin in targeting cancer cell signalling pathways. *Plants* 12 (9), 1782.
- Dytrych, P., Kejik, Z., Hajdudch, J., Kaplanek, R., Vesela, K., Kucnirova, K., Skaliczkova, M., Venhauerova, A., Hoskovec, D., Martasek, P., Jakubek, M., 2023. Therapeutic potential and limitations of curcumin as antimetastatic agent. *Biomed. Pharmacother.* 163, 114758.
- Zhou, J., Zhang, L., Yan, J., Hou, A., Sui, W., Sun, M., 2023. Curcumin induces ferroptosis in A549 CD133<sup>+</sup> cells through the GSH-GPX4 and FSP1-CoQ10-NAPH pathways. *Discov. Med.* 35 (176), 251–263.
- Li, Y., Zhang, T., 2014. Targeting cancer stem cells by curcumin and clinical applications. *Cancer Lett.* 346 (2), 197–205.
- Lim, K.J., Bisht, S., Bar, E.E., Maitra, A., Eberhart, C.G., 2011. A polymer nanoparticle formulation of curcumin inhibits growth, clonogenicity and stem-like fraction in malignant brain tumors. *Cancer Biol. Ther.* 11 (5), 464–473.
- Poonaki, E., Nickel, A.C., Shafiee Ardestani, M., Rademacher, L., Kaul, M., Apartsin, E., Meuth, S.G., Gorji, A., Janiak, C., Kahler, U.D., 2022. CD133-functionalized gold nanoparticles as a carrier platform for telaglenastat (CB-839) against tumor stem cells. *Int. J. Mol. Sci.* 23 (10).
- Swaminathan, S.K., Roger, E., Toti, U., Niu, L., Ohlfest, J.R., Panyam, J., 2013. CD133-targeted paclitaxel delivery inhibits local tumor recurrence in a mouse model of breast cancer. *J. Control. Release* 171 (3), 280–287.
- Wang, Z., Sun, M., Li, W., Fan, L., Zhou, Y., Hu, Z., 2020. A novel CD133- and EpCAM-targeted liposome with redox-responsive properties capable of synergistically eliminating liver cancer stem cells. *Front. Chem.* 8, 649.
- Yang, Z.F., Ngai, P., Ho, D.W., Yu, W.C., Ng, M.N., Lau, C.K., Li, M.L., Tam, K.H., Lam, C. T., Poon, R.T., Fan, S.T., 2008. Identification of local and circulating cancer stem cells in human liver cancer. *Hepatology* 47 (3), 919–928.
- Yang, Z.F., Ho, D.W., Ng, M.N., Lau, C.K., Yu, W.C., Ngai, P., Chu, P.W., Lam, C.T., Poon, R.T., Fan, S.T., 2008. Significance of CD90<sup>+</sup> cancer stem cells in human liver cancer. *Cancer Cell* 13 (2), 153–166.
- Bakalova, R., Ohba, H., Zhelev, Z., Ishikawa, M., Baba, Y., 2004. Quantum dots as photosensitizers? *Nat. Biotechnol.* 22 (11), 1360–1361.
- Li, D., Zhang, T., Gu, W., Li, P., Cheng, X., Tong, T., Wang, W., 2015. The ALDH1<sup>+</sup> subpopulation of the human NMPF-1 cell line exhibits cancer stem-like characteristics. *Oncol. Rep.* 33 (5), 2291–2298.
- Chen, C.H., Ferreira, J.C., Gross, E.R., Mochly-Rosen, D., 2014. Targeting aldehyde dehydrogenase 2: new therapeutic opportunities. *Physiol. Rev.* 94 (1), 1–34.
- Therachiyil, L., Krishnankutty, R., Ahmad, F., Mateo, J.M., Uddin, S., Korashy, H.M., 2022. Aryl hydrocarbon receptor promotes cell growth, stemness like characteristics, and metastasis in human ovarian cancer via activation of PI3K/Akt, beta-catenin, and epithelial to mesenchymal transition pathways. *Int. J. Mol. Sci.* 23 (12).
- Shao, C., Sullivan, J.P., Girard, L., Augustyn, A., Yenerall, P., Rodriguez-Canales, J., Liu, H., Behrens, C., Shay, J.W., Wistuba, I.I., Minna, J.D., 2014. Essential role of aldehyde dehydrogenase 1A3 for the maintenance of non-small cell lung cancer stem cells is associated with the STAT3 pathway. *Clin. Cancer Res.* 20 (15), 4154–4166.
- Liu, S.Y., Zheng, P.S., 2013. High aldehyde dehydrogenase activity identifies cancer stem cells in human cervical cancer. *Oncotarget* 4 (12), 2462–2475.
- Vira, D., Basak, S.K., Veena, M.S., Wang, M.B., Batra, R.K., Srivatsan, E.S., 2012. Cancer stem cells, microRNAs, and therapeutic strategies including natural products. *Cancer Metastasis Rev.* 31 (3–4), 733–751.
- Sher, G., Masoodi, T., Patil, K., Akhtar, S., Kuttikrishnan, S., Ahmad, A., Uddin, S., 2022. Dysregulated FOXM1 signaling in the regulation of cancer stem cells. *Semin. Cancer Biol.* 86 (Pt 3), 107–121.
- Dastidar, D.G., Das, A., Datta, S., Ghosh, S., Pal, M., Thakur, N.S., Banerjee, U.C., Chakrabarti, G., 2019. Paclitaxel-encapsulated core-shell nanoparticle of cetyl alcohol for active targeted delivery through oral route. *Nanomedicine* 14 (16), 2121–2150 (Lond).
- Hothi, P., Martins, T.J., Chen, L., Deleyrolle, L., Yoon, J.G., Reynolds, B., Foltz, G., 2012. High-throughput chemical screens identify disulfiram as an inhibitor of human glioblastoma stem cells. *Oncotarget* 3 (10), 1124–1136.
- Liu, P., Brown, S., Goktug, T., Channathodiyil, P., Kannappan, V., Hugnot, J.P., Guichet, P.O., Bian, X., Armesilla, A.L., Darling, J.L., Wang, W., 2012. Cytotoxic effect of disulfiram/copper on human glioblastoma cell lines and ALDH-positive cancer-stem-like cells. *Br. J. Cancer* 107 (9), 1488–1497.
- Yip, N.C., Fombon, I.S., Liu, P., Brown, S., Kannappan, V., Armesilla, A.L., Xu, B., Cassidy, J., Darling, J.L., Wang, W., 2011. Disulfiram modulated ROS-MAPK and NFκB pathways and targeted breast cancer cells with cancer stem cell-like properties. *Br. J. Cancer* 104 (10), 1564–1574.
- Lipsky, J.J., Shen, M.L., Naylor, S., 2001. *In vivo* inhibition of aldehyde dehydrogenase by disulfiram. *Chem. Biol. Interact.* 130–132 (1–3), 93–102.
- Wang, Z., Tan, J., McConville, C., Kannappan, V., Tawari, P.E., Brown, J., Ding, J., Armesilla, A.L., Irache, J.M., Mei, Q.B., Tan, Y., Liu, Y., Jiang, W., Bian, X.W., Wang, W., 2017. Poly lactic-co-glycolic acid controlled delivery of disulfiram to target liver cancer stem-like cells. *Nanomedicine* 13 (2), 641–657.
- Yang, L., Sun, J., Li, M., Long, Y., Zhang, D., Guo, H., Huang, R., Yan, J., 2021. Oxidized low-density lipoprotein links hypercholesterolemia and bladder cancer aggressiveness by promoting cancer stemness. *Cancer Res.* 81 (22), 5720–5732.
- Charbe, N.B., Lagos, C.F., Ortiz, C.A.V., Tambuwala, M., Palakurthi, S.S., Zacconi, F.C., 2022. PCSK9 conjugated liposomes for targeted delivery of paclitaxel to the cancer cell: a proof-of-concept study. *Biomed. Pharmacother.* 153, 113428.
- Zhan, T., Rindtorff, N., Boutros, M., 2017. Wnt signaling in cancer. *Oncogene* 36 (11), 1461–1473.
- Nasrolahi, A., Azizdoost, S., Radoszkiewicz, K., Najafi, S., Ghaedrahmati, F., Anbiyaee, O., Khoshnam, S.E., Farzaneh, M., Uddin, S., 2023. Signaling pathways governing glioma cancer stem cells behavior. *Cell Signal.* 101, 110493.
- Ram Makena, M., Gatla, H., Verlekar, D., Sukhavas, S., Manoj, K.P., Kartick, C.P., 2019. Wnt/beta-catenin signaling: the culprit in pancreatic carcinogenesis and therapeutic resistance. *Int. J. Mol. Sci.* 20 (17), 4242.
- Valcourt, D.M., Dang, M.N., Wang, J., Day, E.S., 2020. Nanoparticles for manipulation of the developmental wnt, hedgehog, and notch signaling pathways in cancer. *Ann. Biomed. Eng.* 48 (7), 1864–1884.
- Liu, Q., Zhu, H., Tiruthani, K., Shen, L., Chen, F., Gao, K., Zhang, X., Hou, L., Wang, D., Liu, R., Huang, L., 2018. Nanoparticle-mediated trapping of wnt family member 5A in tumor microenvironments enhances immunotherapy for B-Raf proto-oncogene mutant melanoma. *ACS Nano* 12 (2), 1250–1261.
- Bhattacharyya, J., Ren, X.R., Mook, R.A., Wang, J., Spasojevic, I., Premont, R.T., Li, X., Chilkoti, A., Chen, W., 2017. Niclosamide-conjugated polypeptide nanoparticles inhibit Wnt signaling and colon cancer growth. *Nanoscale* 9 (34), 12709–12717.
- Motawi, T.K., El-Maraghy, S.A., ElMeshad, A.N., Nady, O.M., Hammam, O.A., 2017. Cromolyn chitosan nanoparticles as a novel protective approach for colorectal cancer. *Chem. Biol. Interact.* 275, 1–12.
- Chen, M., Wang, J., Lu, J., Bond, M.C., Ren, X.R., Lyerly, H.K., Barak, L.S., Chen, W., 2009. The anti-helminthic niclosamide inhibits Wnt/Frizzled1 signaling. *Biochemistry* 48 (43), 10267–10274.
- Ganesh, S., Koser, M.L., Cyr, W.A., Chopda, G.R., Tao, J., Shui, X., Ying, B., Chen, D., Pandya, P., Chipumuro, E., Siddiquee, Z., Craig, K., Lai, C., Dudek, H., Monga, S.P., Wang, W., Brown, B.D., Abrams, M.T., 2016. Direct pharmacological inhibition of beta-catenin by RNA interference in tumors of diverse origin. *Mol. Cancer Ther.* 15 (9), 2143–2154.
- Riley, R.S., Day, E.S., 2017. Frizzled7 antibody-functionalized nanoshells enable multivalent binding for wnt signaling inhibition in triple negative breast cancer cells. *Small* 13 (26).
- Miller-Kleinhenz, J., Guo, X., Qian, W., Zhou, H., Bozeman, E.N., Zhu, L., Ji, X., Wang, Y. A., Styblo, T., O'Regan, R., Mao, H., Yang, L., 2018. Dual-targeting Wnt and uPA receptors using peptide conjugated ultra-small nanoparticle drug carriers inhibited cancer stem-cell phenotype in chemo-resistant breast cancer. *Biomaterials* 152, 47–62.
- Mumm, J.S., Kopan, R., 2000. Notch signaling: from the outside in. *Dev. Biol.* 228 (2), 151–165.
- Matsui, W.H., 2016. Cancer stem cell signaling pathways. *Medicine* 95 (1 Suppl 1), S8–S19 (Baltimore).
- Hill, R., Wu, H., 2009. PTEN, stem cells, and cancer stem cells. *J. Biol. Chem.* 284 (18), 11755–11759.
- Espinoza, I., Miele, L., 2013. Notch inhibitors for cancer treatment. *Pharmacol. Ther.* 139 (2), 95–110.
- Mamaeva, V., Niemi, R., Beck, M., Ozliseli, E., Desai, D., Landor, S., Gronroos, T., Kronqvist, P., Pettersen, I.K., McCormack, E., Rosenholm, J.M., Linden, M., Sahlgren, C., 2016. Inhibiting notch activity in breast cancer stem cells by glucose functionalized nanoparticles carrying gamma-secretase inhibitors. *Mol. Ther.* 24 (5), 926–936.

- Wan, X., Liu, C., Lin, Y., Fu, J., Lu, G., Lu, Z., 2019. pH sensitive peptide functionalized nanoparticles for co-delivery of erlotinib and DAPT to restrict the progress of triple negative breast cancer. *Drug Deliv.* 26 (1), 470–480.
- Yang, H., Li, Y., Li, T., Xu, M., Chen, Y., Wu, C., Dang, X., Liu, Y., 2014. Multifunctional core/shell nanoparticles cross-linked polyetherimide-folic acid as efficient Notch-1 siRNA carrier for targeted killing of breast cancer. *Sci. Rep.* 4, 7072.
- Deng, X., Cao, M., Zhang, J., Hu, K., Yin, Z., Zhou, Z., Xiao, X., Yang, Y., Sheng, W., Wu, Y., Zeng, Y., 2014. Hyaluronic acid-chitosan nanoparticles for co-delivery of miR-34a and doxorubicin in therapy against triple negative breast cancer. *Biomaterials* 35 (14), 4333–4344.
- Purow, B., 2012. Notch inhibition as a promising new approach to cancer therapy. *Adv. Exp. Med. Biol.* 727, 305–319.
- Ruiz i Altaba, A., Sanchez, P., Dahmane, N., 2002. Gli and hedgehog in cancer: tumours, embryos and stem cells. *Nat. Rev. Cancer* 2 (5), 361–372.
- Cochrane, C.R., Szczepny, A., Watkins, D.N., Cain, J.E., 2015. Hedgehog signaling in the maintenance of cancer stem cells. *Cancers* 7 (3), 1554–1585 (Basel).
- Melamed, J.R., Morgan, J.T., Ioele, S.A., Gleghorn, J.P., Sims-Mourtada, J., Day, E.S., 2018. Investigating the role of Hedgehog/GLI1 signaling in glioblastoma cell response to temozolomide. *Oncotarget* 9 (43), 27000–27015.
- Rubin, L.L., de Sauvage, F.J., 2006. Targeting the Hedgehog pathway in cancer. *Nat. Rev. Drug Discov.* 5 (12), 1026–1033.
- Pasca di Magliano, M., Hebrok, M., 2003. Hedgehog signalling in cancer formation and maintenance. *Nat. Rev. Cancer* 3 (12), 903–911.
- Kiesslich, T., Berr, F., Alinger, B., Kemmerling, R., Pichler, M., Ocker, M., Neureiter, D., 2012. Current status of therapeutic targeting of developmental signalling pathways in oncology. *Curr. Pharm. Biotechnol.* 13 (11), 2184–2220.
- Gonnissen, A., Isebaert, S., Haustermans, K., 2013. Hedgehog signaling in prostate cancer and its therapeutic implication. *Int. J. Mol. Sci.* 14 (7), 13979–14007.
- Bhateja, P., Cherian, M., Majumder, S., Ramaswamy, B., 2019. The hedgehog signaling pathway: a viable target in breast cancer? *Cancers* 11 (8), 1126.
- Verma, R.K., Yu, W., Singh, S.P., Shankar, S., Srivastava, R.K., 2015. Anthrocol-encapsulated PLGA nanoparticles inhibit pancreatic cancer stem cell growth by modulating sonic hedgehog pathway. *Nanomedicine* 11 (8), 2061–2070.
- Verma, R.K., Yu, W., Shrivastava, A., Shankar, S., Srivastava, R.K., 2016. alpha-Mangostin-encapsulated PLGA nanoparticles inhibit pancreatic carcinogenesis by targeting cancer stem cells in human, and transgenic (Kras(G12D), and Kras(G12D)/tp53R270H) mice. *Sci. Rep.* 6, 32743.
- Ingallina, C., Costa, P.M., Ghirga, F., Klippstein, R., Wang, J.T., Berardozi, S., Hodgins, N., Infante, P., Pollard, S.M., Botta, B., Al-Jamal, K.T., 2017. Polymeric glabrescione B nanocapsules for passive targeting of Hedgehog-dependent tumor therapy *in vitro*. *Nanomedicine* 12 (7), 711–728 (Lond).
- Hu, K., Zhou, H., Liu, Y., Liu, Z., Liu, J., Tang, J., Li, J., Zhang, J., Sheng, W., Zhao, Y., Wu, Y., Chen, C., 2015. Hyaluronic acid functional amphiphatic and redox-responsive polymer particles for the co-delivery of doxorubicin and cyclopamine to eradicate breast cancer cells and cancer stem cells. *Nanoscale* 7 (18), 8607–8618.
- Zhao, J., Wang, H., Hsiao, C.H., Chow, D.S., Koay, E.J., Kang, Y., Wen, X., Huang, Q., Ma, Y., Bankson, J.A., Ullrich, S.E., Overwijk, W., Maitra, A., Piwnica-Worms, D., Fleming, J.B., Li, C., 2018. Simultaneous inhibition of hedgehog signaling and tumor proliferation remodels stroma and enhances pancreatic cancer therapy. *Biomaterials* 159, 215–228.
- Yang, R., Mondal, G., Wen, D., Mahato, R.I., 2017. Combination therapy of paclitaxel and cyclopamine polymer-drug conjugates to treat advanced prostate cancer. *Nanomedicine* 13 (2), 391–401.
- Xu, Y., Chenna, V., Hu, C., Sun, H.X., Khan, M., Bai, H., Yang, X.R., Zhu, Q.F., Sun, Y.F., Maitra, A., Fan, J., Anders, R.A., 2012. Polymeric nanoparticle-encapsulated hedgehog pathway inhibitor HPI-1 (NanoHHI) inhibits systemic metastases in an orthotopic model of human hepatocellular carcinoma. *Clin. Cancer Res.* 18 (5), 1291–1302.
- Chenna, V., Hu, C., Pramanik, D., Aftab, B.T., Karikari, C., Campbell, N.R., Hong, S.M., Zhao, M., Rudek, M.A., Khan, S.R., Rudin, C.M., Maitra, A., 2012. A polymeric nanoparticle encapsulated small-molecule inhibitor of Hedgehog signaling (NanoHHI) bypasses secondary mutational resistance to Smoothed antagonists. *Mol. Cancer Ther.* 11 (1), 165–173.
- Melamed, J.R., Ioele, S.A., Hannum, A.J., Ullman, V.M., Day, E.S., 2018. Polyethylenimine-spherical nucleic acid nanoparticles against Gli1 reduce the chemoresistance and stemness of glioblastoma cells. *Mol. Pharm.* 15 (11), 5135–5145.
- Zhou, P., Cao, Y., Liu, X., Yu, T., Xu, Q., You, C., Gao, X., Wei, Y., 2018. Delivery siRNA with a novel gene vector for glioma therapy by targeting Gli1. *Int. J. Nanomed.* 13, 4781–4793.
- Botchkina, L.L., Rowehl, R.A., Rivadeneira, D.E., Karpeh Jr., M.S., Crawford, H., Dufour, A., Ju, J., Wang, Y., Leyfman, Y., Botchkina, G.I., 2009. Phenotypic subpopulations of metastatic colon cancer stem cells: genomic analysis. *Cancer Genom. Proteom.* 6 (1), 19–29.
- Turdo, A., Veschi, V., Gaggianesi, M., Chinnici, A., Bianca, P., Todaro, M., Stassi, G., 2019. Meeting the challenge of targeting cancer stem cells. *Front. Cell Dev. Biol.* 7, 16.
- Shah, V., Taratula, O., Garbuzenko, O.B., Taratula, O.R., Rodriguez-Rodriguez, L., Minko, T., 2013. Targeted nanomedicine for suppression of CD44 and simultaneous cell death induction in ovarian cancer: an optimal delivery of siRNA and anticancer drug. *Clin. Cancer Res.* 19 (22), 6193–6204.
- Yang, S.J., Pai, J.A., Yao, C.J., Huang, C.H., Chen, J.L.Y., Wang, C.H., Chen, K.C., Shieh, M.J., 2023. SN38-loaded nanomedicine mediates chemo-radiotherapy against CD44-expressing cancer growth. *Cancer Nanotechnol.* 14 (1), 1.
- Asghari, F., Khademi, R., Esmaili Ranjbar, F., Veisi Malekshahi, Z., Faridi Majidi, R., 2019. Application of nanotechnology in targeting of cancer stem cells: a review. *Int. J. Stem Cells* 12 (2), 227–239.
- Tangudu, N.K., Verma, V.K., Clemons, T.D., Beevi, S.S., Hay, T., Mahidhara, G., Raja, M., Nair, R.A., Alexander, L.E., Patel, A.B., Jose, J., Smith, N.M., Zdyrko, B., Bourdoncle, A., Luzinov, I., Iyer, K.S., Clarke, A.R., Kumar, L.D., 2015. RNA interference using c-myc-conjugated nanoparticles suppresses breast and colorectal cancer models. *Mol. Cancer Ther.* 14 (5), 1259–1269.
- Nunes, T., Pons, T., Hou, X., Van Do, K., Caron, B., Rigal, M., Di Benedetto, M., Palpat, B., Leboeuf, C., Janin, A., Bousquet, G., 2019. Pulsed-laser irradiation of multifunctional gold nanoshells to overcome trastuzumab resistance in HER2-overexpressing breast cancer. *J. Exp. Clin. Cancer Res.* 38 (1), 306.
- Murase, K., Nishimoto, K., Mimura, A., Aoki, M., Hamakawa, K., Banura, N., 2015. Application of magnetic particle imaging to pulmonary imaging using nebulized magnetic nanoparticles: Phantom and small animal experiments. In: *Proceedings of the 5th International Workshop on Magnetic Particle Imaging (IWMP)*. IEEE, p. 1–1.
- Ulbrich, K., Hola, K., Subr, V., Bakandritsos, A., Tucek, J., Zboril, R., 2016. Targeted drug delivery with polymers and magnetic nanoparticles: covalent and noncovalent approaches, release control, and clinical studies. *Chem. Rev.* 116 (9), 5338–5431.
- Fu, N., Hu, Y., Shi, S., Ren, S., Liu, W., Su, S., Zhao, B., Weng, L., Wang, L., 2018. Au nanoparticles on two-dimensional MoS<sub>2</sub> nanosheets as a photoanode for efficient photoelectrochemical miRNA detection. *Analyst* 143 (7), 1705–1712.
- Parungo, C.P., Ohnishi, S., De Grand, A.M., Laurence, R.G., Soltesz, E.G., Colson, Y.L., Kang, P.M., Mihajlic, T., Cohn, L.H., Frangioni, J.V., 2004. *In vivo* optical imaging of pleural space drainage to lymph nodes of prognostic significance. *Ann. Surg. Oncol.* 11 (12), 1085–1092.
- Gao, X., Cui, Y., Levenson, R.M., Chung, L.W., Nie, S., 2004. *In vivo* cancer targeting and imaging with semiconductor quantum dots. *Nat. Biotechnol.* 22 (8), 969–976.
- Dubertret, B., Skourides, P., Norris, D.J., Noireaux, V., Brivanlou, A.H., Libchaber, A., 2002. *In vivo* imaging of quantum dots encapsulated in phospholipid micelles. *Science* 298 (5599), 1759–1762.
- Aswathy, R.G., Yoshida, Y., Maekawa, T., Kumar, D.S., 2010. Near-infrared quantum dots for deep tissue imaging. *Anal Bioanal Chem* 397 (4), 1417–1435.
- Savic, R., Luo, L., Eisenberg, A., Maysinger, D., 2003. Micellar nanocontainers distribute to defined cytoplasmic organelles. *Science* 300 (5619), 615–618.
- Ghezzi, M., Pescina, S., Padula, C., Santi, P., Del Favero, E., Cantu, L., Nicoli, S., 2021. Polymeric micelles in drug delivery: an insight of the techniques for their characterization and assessment in biorelevant conditions. *J. Control. Release* 332, 312–336.
- Bozzuto, G., Molinari, A., 2015. Liposomes as nanomedical devices. *Int. J. Nanomed.* 10, 975–999.
- Burlaka, A., Lukin, S., Prylutska, S., Remeniak, O., Prylutsky, Y., Shuba, M., Maksimenko, S., Ritter, U., Scharff, P., 2010. Hyperthermic effect of multi-walled carbon nanotubes stimulated with near infrared irradiation for anticancer therapy: *in vitro* studies. *Exp. Oncol.* 32 (1), 48–50.
- Batra, S., Sharma, S., Mehra, N.K., 2022. Carbon Nanotubes For Drug Delivery Applications, *Handbook of Carbon Nanotubes*. Springer, pp. 1–14.
- Borah, A., P.V., Girija, A.R., Balasubramanian, S., Rochani, A.K., 2017. Poly-lactic-co-glycolic acid nanoformulation of small molecule antagonist GANT61 for cancer annihilation by modulating hedgehog pathway. *NanoWorld J.* 3 (1), 1–10.