

Review

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Review **Revolutionizing Cardiovascular Health with Nano Encapsulated Omega-3 Fatty Acids: A Nano-Solution Approach**

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Abstract: Omega-3 polyunsaturated fatty acids (ω-3 PUFAs) offer diverse health benefits, such as supporting cardiovascular health, improving cognitive function, promoting joint and musculoskeletal health, and contributing to healthy aging. Despite their advantages, challenges like oxidation susceptibility, low bioavailability, and potential adverse effects at high doses persist. Nanoparticle encapsulation emerges as a promising avenue to address these limitations while preserving stability, enhanced bioavailability, and controlled release. This comprehensive review explores the therapeutic roles of omega-3 fatty acids, critically appraising their shortcomings and delving into modern encapsulation strategies. Furthermore, it explores the potential advantages of metal–organic framework nanoparticles (MOF NPs) compared to other commonly utilized nanoparticles in improving the therapeutic effectiveness of omega-3 fatty acids within drug delivery systems (DDSs). Additionally, it outlines future research directions to fully exploit the therapeutic benefits of these encapsulated omega-3 formulations for cardiovascular disease treatment.

Keywords: omega-3 fatty acids; cardiovascular disease; nanoparticle encapsulation; metal–organic framework; drug delivery systems

1. Introduction

Cardiovascular diseases (CVDs) remain the world's leading cause of mortality, claiming a staggering 695,000 lives in the United States alone in 2021 [\[1\]](#page-18-0), translating to roughly one death every 30 s [\[2\]](#page-18-1). According to a 2019 report by the World Health Organization, CVD contributes to 32% of total worldwide fatalities, with 85% of these fatalities attributed to heart attacks or strokes [\[3\]](#page-18-2). Major contributors and risk factors for CVD include elevated blood pressure, diabetes, and cholesterol levels, as well as smoking, unhealthy diet, obesity, and physical inactivity [\[4\]](#page-18-3).

According to a recent report from the American Heart Association, an intake of approximately 3 g of omega-3 fatty acids daily, whether obtained from food or supplements, appears to be the optimal amount for reducing high blood pressure and preventing cardiovascular disease, as indicated by a review of multiple research studies [\[5\]](#page-18-4). Omega-3 fatty acids (ω -3 FAs) are categorized as polyunsaturated fatty acids (PUFAs) that include a minimum of a single double bond (C=C) between the carbon atoms in the third and fourth positions from the methyl end of the fatty acid [\[6](#page-18-5)[,7\]](#page-18-6). PUFAs are long-chain fatty acids (LC-FAs) found in oily fish like sardines, tuna, and salmon, and other seafood like shellfish, algae, and shrimp, as well as particular plants and nut-based oils [\[8\]](#page-18-7). The most

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common bioactive ω-3 FAs are eicosapentaenoic acid (EPA) (C20:5 ω-3), docosahexaenoic acid (DHA) (C22:6 ω-3), and α-linolenic acid (ALA) (C18:3 ω) [\[9\]](#page-18-8). ALA can undergo various elongation and desaturation processes within the body to be converted into EPA and DHA [\[10\]](#page-18-9). However, the conversion rates of ALA to EPA and DHA may be relatively low, potentially insufficient to confer health benefits. Therefore, it is essential to ensure adequate intake of EPA and DHA for various aspects of health, including cardiovascular health, brain function, and inflammation regulation. PUFAs are essential nutrients that must be obtained from the diet because the body cannot synthesize them. ω <u>και του κατά</u> του και του κατά του και του κατά του και του κατά του κατά του κατά του κατά του κατά του και τ
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PUFAs play numerous physiological roles involving cell signaling and transmission, intercellular contact, membrane fluidity, phospholipid membrane maintenance, reduced inflammation [\[11](#page-18-10)[,12\]](#page-18-11), and improved fatty acid oxidation [\[13\]](#page-18-12). Hence, a deficiency in ω-3 FAs can deteriorate bone health [\[14–](#page-18-13)[16\]](#page-18-14), cardiovascular health [\[17\]](#page-18-15), and skin and neurological health [\[13,](#page-18-12)[18\]](#page-18-16) (Figure [1\)](#page-2-0). Along with this, ω-3 FA ingestion is associated ω with a lowered prevalence of inflammation [\[19\]](#page-18-17) through the lowering of pro-inflammatory cytokines triggered by oxidative stress in macrophages isolated from female mice [\[20\]](#page-18-18) and r_1 various studies have also assessed the anti-inhibitory effect of ω-3 FAs on cancers in female mice [\[22\]](#page-18-20) and cancer cell lines [\[23\]](#page-18-21). Over the decades, various preclinical and clinical studies have been conducted using ω -3 FAs to show their efficacy in reducing cardiovascular ailments. This review paper will provide an overview of ω -3 FAs with a focus on their effect on the cardiovascular system, along with shortcomings in the delivery of ω -3 FAs to which nanoparticles could be potentially a viable solution.

Figure 1. Types, sources, and health benefits of omega-3 fatty acids. Three types of n-3 fatty acids are ALA (plant-based sources), DHA (animal-based sources), and EPA (animal-based sources). As mentioned earlier, n-3 fatty acids have a wide range of health benefits related to cardiovascular, ocular, cognitive, and dermal health.

1.1. Preclinical Studies

mental units have proved the effectiveness of ω -3 FAs in cardioprotection and decreasing Over the years, various pre-clinical studies conducted on a range of animal experimarkers of cardiovascular stress, as summarized in Table [1.](#page-3-0)

Table 1. Summary of outcomes of pre-clinical animal studies published between 1992 and 2022 on cardiovascular health utilizing diets rich in omega-3 fatty acids (ω -3 FAs). (\downarrow indicates a positive decrease in outcome). REF = reference or control diet, SF = sheep fat diet, PUFA = polyunsaturated fatty acids, SSO = sunflower seed oil diet, TFO = tuna fish oil diet, EPA = eicosapentaenoic acid, DHA = docosahexaenoic acid, LNA = Linoleic acid.

1.2. Clinical Studies

Several significant meta-analyses of ω-3 FAs and their connection to cardiovascular morbidity and mortality have been published in the last four decades. Some of these studies concluded that fish oil supplementation lowers the risk of cardiovascular events and the mortality rates from CVD, while others did not support this conclusion, as seen in Table [2.](#page-4-0)

Table 2. Summary of outcomes of randomized clinical trials (RCTs) published between 1989 and 2021 on cardiovascular health utilizing diets rich in ω-3 FAs. (↓ indicates a positive decrease in outcome, ↓ indicates a negative decrease in outcome, and ↑ indicates a negative increase in outcome).

Table 2. *Cont.*

Table 2. *Cont.*

The differences in research findings might be attributed to the various fatty acids employed in these studies, which ranged fromo-3 carboxylic acid formulation to ethyl esters of EPA, DHA, and ALA. The omega-3 CA employed in the STRENGTH study was EPA as a free fatty carboxylic acid, which has greater absorption in a diet low in fat than EPA and DHA ethyl esters, comparable to the EPA ethyl ester used in the REDUCE-IT study. Consequently, omega-3 CA increases blood levels of EPA and DHA when combined with a diet low in fat, but not with a standard diet. In the STRENGTH study, omega-3 CA was administered independent of eating habits or dietary fat composition, which might culminate in fluctuation in EPA and DHA levels in the blood and perhaps reduce the impact on cardiovascular events. Icosapent ethyl additionally contains approximately 25% higher EPA in each dose than omega-3 CA, resulting in 61% greater EPA levels in blood in REDUCE-IT versus STRENGTH from comparable baseline values [\[41\]](#page-19-15).

1.3. Epidemiological Studies

Apart from clinical studies, epidemiological studies were also conducted on the effect of ω -[3](#page-7-0) FAs on the reduction of cardiovascular events. The results of the studies in Table 3 were mixed, with some claiming that ω -3 FAs decrease cardiovascular events and others claiming no such outcome.

Table 3. Summary of outcomes of cohort studies published between 2000 and 2021 on cardiovascular health utilizing diets rich in ω-3 FAs. (↓ indicates a positive decrease in outcome, ↓ indicates a negative decrease in outcome, and ↑ indicates a negative increase in outcome).

Table 3. *Cont.*

1.4. Case-Control Studies

The number of case-control studies measuring EPA and DHA levels in plasma as a biomarker of ω -3 FA after 2004 declined due to the introduction of the new concept of the omega-3 index (O3I) [\[50\]](#page-20-2). The omega-3 index is defined as the proportion of EPA and DHA in a total of 26 distinct fatty acids in the membrane of red blood cells (RBCs), with many studies claiming that an O3I of 8% is required to elicit the cardioprotective effect of ω -3 FAs [\[51\]](#page-20-3). There are many reasons why measuring EPA and DHA in erythrocyte membranes is a more accurate method of assessing ω -3 FA intake in the diet. RBC fatty acid content has relatively little biological variability in comparison to plasma. Erythrocyte lipids are nearly entirely composed of phospholipids, and RBC fatty acid content represents tissue fatty acid composition [\[52\]](#page-20-4). Table [4](#page-8-0) shows the results of a few of the earliest case-control studies.

Table 4. Summary of outcomes of case-control studies published between 1990 and 2010 on cardiovascular health utilizing diets rich in ω -3 FAs. (\downarrow indicates a positive decrease in outcome.)

Table 4. *Cont.*

2. Nanoparticles and Administration of Omega-3

2.1. Obstacles in the Effective Administration of Omega-3 PUFAs

There have been concerns regarding the optimal source and route of administration of ω-3 FAs for human consumption. Even though the simplest method of ω-3 FA intake is through eating fish, decreasing fish stocks worldwide and biomagnification of toxic trace elements due to water pollution are proving to be valid concerns [\[58\]](#page-20-10). An alternate method is consuming ω -3 FA supplements. The issue with supplements is that they may cause undesired side effects, such as stomach upsets, stale breath, nausea, etc. (Figure [2\)](#page-10-0). Moreover, the release of ω -3 FAs in supplements occurs rapidly, simultaneously delivering the entire amount [\[59\]](#page-20-11).

Undesirable side effects of Omega-3 fatty acid capsule supplementation

Figure 2. Common undesirable side effects of omega-3 fish oil capsule supplements.

The bioavailability of long-chain ω -3 PUFA is another complex issue that needs to be ω ω-3 PUFAs, when ingested as pure oil, cannot be completely absorbed by the cells in ω consummated in an emulsified way rather than in a pure form [\[65\]](#page-20-17). Widely ingested edible tackled. Bioavailability is a relative term that refers to both the rate of absorption and the amount of the substance ingested [\[60\]](#page-20-12). Bioavailability encompasses both absorption speed and quantity absorbed. It is influenced by gastrointestinal absorption and transport rates to the portal system. In a broader sense, it considers the amount reaching systemic circulation or the intended destination. Understanding this aids in pharmacokinetics and dietary planning. NPs can significantly improve the pharmacokinetics of drugs by increasing their solubility [\[61\]](#page-20-13), stability [\[62\]](#page-20-14), permeability [\[63\]](#page-20-15), half-life, and residence time [\[64\]](#page-20-16). The major cause of the limited bioavailability of long-chain n-3 PUFAs is low solubility in the aqueous gastrointestinal fluids of the GI tract, alongside their vulnerability to chemical breakdown during transit through the stomach [\[65\]](#page-20-17). Long-chain omega-3 PUFAs are mostly present in fish as triacylglycerides (TAGs), phospholipids (PLs), and free fatty acids (FFAs) [\[66\]](#page-20-18). our intestines, resulting in reduced bioavailability [\[67\]](#page-20-19). Fish oils are transformed into oil-in-water emulsions within the mouth and stomach, while emulsified oils are colloidal before consumption. Lipases in the stomach and pancreas subsequently attach to the surface of the lipid drop and begin the process of lipid digestion, which converts TAGs into FFAs and monoacylglycerols (MAGs). The digestive products, FFAs and MAGs, subsequently combine with bile and PLs to produce blended micelles that transport the lipids across the mucus membrane to the epithelial cell surfaces, where they are absorbed. Investigations have demonstrated that the bioavailability of ω -3 PUFAs is enhanced when oils and products have less ω -3 PUFAs compared to the suggested daily guideline; also, the minimal quantity of PUFAs taken is poorly absorbed. The transformation rate of plantsourced ω -3 FAs, namely, ALA into EPA and DHA, the primary ω -3 PUFAs accountable for the reported benefits, is only 3–6%, with DHA having an inadequate transformation rate maxing out at 1% [\[68\]](#page-20-20).

Apart from these complications, lipid oxidation is an additional problem that needs to be addressed. Among the different types of ω-3 FAs, EPA and DHA are extremely vulnerable to the oxidation of lipids due to the presence of numerous double bonds [\[69\]](#page-20-21). The oxidative degradation of lipids in fish oil, along with other PUFA-rich and fortified foods is a severe issue that frequently results in a decrease in shelf-life, customer acceptance, performance, nutrient content, and quality [\[70\]](#page-20-22). The oxidation of these ω -3 FAs results in aldehydes that are toxic to proteins and nucleic acids in the human body, namely 4hydroxy-2-nonenal and malondialdehyde [\[71\]](#page-20-23). The toxicity stems from their capacity to crosslink proteins and attach covalently to nucleic acids.

Lipid oxidation in fish oils is heightened by exposure to light, oxygen, and heat. Lipid oxidation creates three major issues: (i) it produces disagreeable unpleasant flavors, (ii) it ω decreases the nutritional content of lipid-containing food items, and (iii) free radicals generated during oxidation may contribute to the occurrence of atherosclerosis in the system [\[72\]](#page-20-24). ω Nevertheless, the low oxidative resistance of ω-3 PUFAs makes these oil-enriched foods ω require potent antioxidant defense to avert oxidative degradation and undesirable flavor formation [\[73\]](#page-20-25). Some solutions to stabilize ω-3 PUFAs include providing antioxidants for ω oxidative stability, mixing and blending with other oils, hydrogenation, and interesterifi-ω cation [\[68\]](#page-20-20). However, another innovative solution might be ω -3 FA encapsulation using nanoparticles as part of nanotechnology.

2.2. Nanoencapsulation of Omega-3 PUFAs

Nanoparticles (NPs) are the focal point of nanomedicine, which is a branch of medicine that relies on smart drug delivery technology that can boost the biological action, pharmacological index, and physiological half-life of the drug loaded inside the body [\[74\]](#page-21-0). Nanoparticles are increasingly becoming popular as drug delivery systems, especially to treat CVDs [\[75\]](#page-21-1) and metabolic disorders [\[76\]](#page-21-2) due to their ability to have a comparatively big surface area that can attach to, adsorb, and transport molecules, including drugs, probes, and protein molecules [\[77\]](#page-21-3). Furthermore, for the administration of medication, engineered nanoparticles, as well as the drug itself, can be synthesized at nanoscale and operate as a carrier for themselves [\[78\]](#page-21-4). There exists an array of nanoparticles, each having distinct features, advantages, disadvantages, and applications (Table [5\)](#page-11-0).

Table 5. Characteristics, uses, advantages, and disadvantages of different classes of nanoparticles.

Table 5. *Cont.*

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tested a nanotechnology-focused method for administering ω-3 FAs to the walls of vascular ω vessels alongside other drugs to avoid occlusive vasculopathy after vascular injury [\[107\]](#page-22-4). In the past few years, multiple studies have been performed that utilized encapsulated ω-3 FAs within NPs as a drug delivery method. An in vitro study by Deshpande et al. The researchers created an ω-3-FA-rich oil-in-water nanoemulsion composition using

flax seed oil naturally rich in ALA, that was administered to cultured vascular cell lines. Combining the administration of 17-βE and CER-laden nanoemulsions had a stronger antiproliferative impact on vascular smooth muscle cells as compared to endothelial cells [\[108\]](#page-22-5). In 2016, the same researchers set out to examine the effectiveness of an ω -3 FA containing 17-βE nanodelivery setup in treating induced atherosclerosis. The study found that a 3-week 17-βE therapy administered in an ω-3-PUFA-encapsulated nanoemulsion setup improved acute vascular damage with just 30% arterial stenosis [109].

Another study conducted by a separate group in 2019 focused on utilizing atorvastatin nano lipid carriers preloaded with ω -3 PUFAs to lower hyperlipidemia. When compared with the commercial formulation, orally administered ω-3-FA-based Atorvastatin reloaded nano lipid carriers resulted in a substantial decrease in low-density lipoprotein and triglyc-− and the blood in the blood [\[110\]](#page-22-7). Through various studies, nanoencapsulation has been shown to increase the bioavailability of ω -3 PUFAs in the body. Through their studies, Wakil et al., 2010 and Sanguansri et al., 2013 proved clearly that microencapsulation can improve the availability of FAs [\[111,](#page-22-8)[112\]](#page-22-9).

Nanofibers like zein have also been used to encapsulate ω-3 PUFAs recently, such as in one study by Busolo et al., where 85 wt% DHA-supplemented fish oil was encapsulated in zein through electrospraying assisted by pressurized gas technology (EAPG). The average particle size and encapsulation efficiency were 3.7 ± 1.8 µm and 84% , respectively. The fortified reconstituted milk with zein/DHA-enriched fish oil microcapsules showed no signs of oxidation even after 45 days in an oxidation test [\[113\]](#page-22-10).

Whey microgels loaded with ω -3 PUFAs were also tested in a study in 2022 where 85 wt% DHA-enriched algal oil was loaded into whey protein microgels by ball milling. The end product had an average particle size of 250 nm, an average diameter of 380 nm, and a polydispersity index of 0.26, indicating the zeta potential. These protein microgels loaded with omega-3 PUFAs addressed several obstacles in the development and storage of omega-3 PUFA oils, such as long-term oxidative resistance and better sensory and textural qualities [\[114\]](#page-22-11).

A summary of studies utilizing unique nanoparticles for the encapsulation of ω -3 FAs, their mechanism of production, physiochemical properties, and observed effect is listed below in Table [6.](#page-14-0)

The most common mechanism for nanoencapsulation of ω -3 FAs in the literature involves physical methods, as illustrated below in Figure [3.](#page-13-0)

Figure 3. Common physical mechanisms for nanoencapsulation of omega-3 PUFAs.

Table 6. Summary of studies encapsulating omega-3 fatty acids using different types of nanoparticles between 2013 and 2022 prepared by a plethora of different techniques and having varying physiochemical properties.

However, lipid-based NPs have their drawbacks as well. Due to their precise crystalline form, they display limited drug loading capacity and the likelihood of drug ejection owing to crystallization during storage [\[115\]](#page-22-12) along with an initial burst discharge of the drug instead of slow controlled drug release [\[116\]](#page-22-13). Other disadvantages during oral administration of lipid-based NPs are the formation of gel of hydrophobic lipid dispersion, restricted loading quantity for hydrophilic formulations, and polymorphic transformation [\[117\]](#page-22-14). Liposomes are another type of lipid-based NPs that are employed for drug delivery; however, they tend to have a reduced solubility window [\[91\]](#page-21-14), problems with drug incorporation and encapsulation, high manufacturing costs, and trouble preserving drug integrity and bioactivity during conjugation [\[75\]](#page-21-1). Microgels, on the other hand, are complicated and time-consuming to mass produce on a large scale, as the yield and stability of individual microgels is highly variable using the currently available technology [\[118\]](#page-22-15). Nanofibers, although super effective in encapsulating omega-3 PUFAs, have their shortcomings too. Their disadvantages include quick disintegration, low mechanical durability, and full dissolution. Therefore, such fibers must be cross-linked to limit their solubility [\[119\]](#page-22-16).

An alternative to avoid this uncontrolled release would be to use metal–organic framework (MOF) NPs, a novel group of composite nanomaterials, consisting of a combination of inorganic and crystalline organic components [\[120\]](#page-22-17). A few of the properties of MOF NPs are their extensive surface area (hollowed-out interior structure) [\[121\]](#page-22-18), a high degree of porosity, configurable pore dimensions, heat resistance, chemical stability [\[122\]](#page-22-19), and post-synthesis alterations; these features elevate MOF NPs over lipid-based NPs when it comes to versatility, adaptability, and customizability [\[123,](#page-22-20)[124\]](#page-22-21). MOF NPs, such as Material Institute Lavoisier 89 nanoparticles (nanoMIL-89), due to their vast list of perks (Figure [4\)](#page-15-0), can mitigate non-specific drug administration to unsuitable sites, pre-activation

of therapeutic agents before reaching the targeted tissue, early immune system approval, and, in certain instances, potentially enhance the pharmacokinetics of drugs at the level of permeability, intake, and dispersion of a drug in the tissue layers [\[125\]](#page-22-22) (Table [7\)](#page-15-1).

Figure 4. Advantageous features of a type of MOF; Material Institute Lavoisier 89 nanoparticles (nanoMIL-89). Note: these advantages were based on the study by Al-Ansari et al. [\[124\]](#page-22-21) that addressed the internalization of MOF NPs in human vascular cells.

Physiochemical Properties of MOF NPs	Related Biomedical Advantages	References
High surface area compared to volume	Ability to make post-synthesis surface modifications	[121, 126]
Biocompatibility	Reduced physiological toxicity	$[127]$
Small size	Higher bioavailability and higher penetrability	[125, 128]
A high degree of porosity	Higher drug loading capability	[122, 129]
Presence of various functional groups	Increased mechanical stability and increased thermal stability	[123, 130]

Table 7. Physiochemical properties and related biomedical advantages of MOF NPs.

Keeping the hydrophobic nature of omega-3 FAs in mind, the most crucial design consideration in MOF NPs is to modify the interior of the NPs from hydrophilic to hydrophobic. There are two main approaches to solving the issue of interior hydrophilicity. The first is grafting hydrophobic polymers into the interior wall of the NP, which involves linking hydrophobic polymer chains to the interior of MOF pores to alter the interior for improved encapsulation of omega-3 FAs [\[131\]](#page-23-5). The second approach incorporates hydrophobic functional groups within the interior, wherein the MOF linkers are chemically modified to contain hydrophobic groups such as isobutyl, alkyl, and isopropyl functional groups [\[132\]](#page-23-6).

The possible administration routes of MOF NPs loaded with omega-3 FAs are specific to the system of choice, which, in this review paper, is the cardiovascular system. There are three possible administration routes to focus on for drug delivery to the cardiovascular system: oral, intravenous, and inhalation routes. However, since inhalation relates more to pulmonary ailments [\[133\]](#page-23-7) and not directly to cardiovascular disease, the former two routes are preferable. Muchow et al., in their study, developed lipid-based omega-3 FA NPs that proved to be patient-friendly and were administered orally [\[134\]](#page-23-8). Another consideration for the oral administration of NPs is coating the NP with polymer substances to protect it from the harsh and acidic gastric environment, for instance, coating NPs with poly lactic-coglycolic acid (PLGA) [\[135\]](#page-23-9) and chitosan [\[136\]](#page-23-10). Omega-3 FA NPs have also successfully been delivered intravenously in some studies [\[110\]](#page-22-7), resulting in increased hyperlipidemic action.

Based on the toxicology of metal-organic framework MIL-89 nanoparticles on embryonic zebrafish development, Al-Ansari et al. said, "The investigation demonstrates that nanoMIL-89 has no developmental harm on zebrafish embryos at low doses (1–10 µM). μ μ High concentrations of nanoMIL-89 ($>$ 30 μ M) significantly impacted hatching time and heart development. The study proves the safety of nanoMIL89 in biological, environmental, and medicinal applications without cytotoxicity beyond a certain concentration of μ 30µM" [\[124\]](#page-22-21).

Other NPs similar to nanoMIL-89, such as MIL88A and MIL101, are also used for the biomedical application of drug delivery. Since both MIL88A and MIL101 share similar features such as small size, high biocompatibility, etc. [\[137\]](#page-23-11), to nanoMIL-89, they have also been extensively studied for carrying anti-cancer drugs such as curcumin [\[138\]](#page-23-12) and doxorubicin [\[139\]](#page-23-13).

SEM and TEM analysis is commonly used to characterize the shape and size of nanoparticles after production, as in the case of nanoMIL-89 below (Figure [5\)](#page-16-0).

Figure 5. NanoMIL-89 (iron oxide nanoparticles), a subcategory of MOF nanoparticles. (**a**) A scanning electron microscope captured an image of nanoMIL-89 (50,000×). (**b**) A transmission electron microscope captured an image of nanoMIL-89 (5000 \times).

Paclitaxel (PTX) in microneedle arrays (PTX-MNAs) has higher anticancer efficacy than free PTX (Taxol) in both in vitro and in vivo experiments [\[140\]](#page-23-14). Chen and Feng's research shows that uncoated gold nanoparticles (GNPs) have the potential for skin applications such as penetration, medication loading/release, and combination with physical procedures to treat skin ailments [\[141\]](#page-23-15). Gao et al. show that copper sulfide nanoparticles (CuS-TRPV1) can operate as a photothermal switch to minimize atherosclerosis, assisting in cardiac imaging and lowering plaque development in mice, with no long-term damage [\[142\]](#page-23-16). Spivak et al. utilized gold NPs loaded with levosimendan (Simdax[®]) in an in vivo study with Wistar rats and concluded that conjugated AuNPs-Simdax[®] had a favorable impact on cardiac contractile capacity. Additionally, IV administration of 30 nm AuNPs resulted in accumulation in the endothelial cells of infarcted arteries and capillaries. Necrobiosis and fibrosis were greatly reduced following all treatments. Conjugate (Simdax + AuNPs) injections resulted in a considerably larger hydrothorax reduction compared to Simdax injections alone $(p < 0.01)$, along with improved cardiac contractile ability. Interestingly, AuNP administration yielded results similar to that of the conjugate [\[143\]](#page-23-17). In 2020, Li et al. successfully demonstrated that a gold-nanorod-based NP that catalyzes continuous NO production safeguards against cardiovascular damage in vitro [\[144\]](#page-23-18). Some studies employ a hybrid NP that combines liposomes coated with metal NPs, as in the case of Bejarano et al., who developed a gold NP-based nanosystem and employed it to optimize the distribution of angiotensin-1–9, which is a cardioprotectant peptide, to the myocardium, helping both

hypertension and myocardial remodeling [\[145\]](#page-23-19). Hussein et al. were successful in producing ZnO and gum NPs loaded with 400 mg of DHA solution using a one-step solid-state process [\[146\]](#page-23-20). A total of 250 mg of DHA were loaded into ZnO NPs using ultrasonication and homogenization methods [\[147\]](#page-23-21). Later, the same researcher also synthesized DHAloaded AgNPs through nanoprecipitation for antidiabetic drug testing [\[148\]](#page-23-22). In a study by El-Daly et al., both DHA-Ag NPs and DHA-ZnO NPs were tested side by side to study the expression of the glucose transport cascade [\[149,](#page-23-23)[150\]](#page-23-24). Lastly, another study implied that an administration of 400 μ g/kg/day gold NPs helps improve myocardial damage induced by isoproterenol in male albino rats [\[151\]](#page-24-0).

Even though MOF NPs have an array of advantages in their applications, they also come with some drawbacks, such as some of the techniques used to synthesize NPs. The solvothermal and microemulsion techniques are highly expensive. The microemulsion technique uses certain surfactants that are classified as pollutants in the environment. The mechanochemical technique requires a large amount of energy and yields amorphous, not crystalline, NPs, which further cannot be used in X-ray crystal structure analysis. Synthetic techniques for the synthesis of MOFs with effective carbon dioxide absorption capability, such as amine scrubbing, include drawbacks such as high energy usage [\[152\]](#page-24-1). Rare-earthbased MOF NPs, namely uranium-, cerium-, lanthanum-, and yttrium-based NPs, are highly expensive, difficult to access, and radioactive, hence dangerous [\[153\]](#page-24-2). To conclude, while MOF NPs provide appealing advantages in a variety of applications, their possible downsides highlight the need for additional research and careful evaluation of their usage to maximize their benefits while mitigating related impediments.

3. Conclusions

In conclusion, encapsulating omega-3 fatty acids with nanoparticles offers a potential strategy for improving the vital nutrients' durability, bioavailability, and targeted administration. Investigators have effectively mitigated oxidation, undesirable taste, and restricted GI absorption linked to free omega-3 fatty acids using various encapsulating strategies, from nanoemulsions to solid lipid and polymeric nanoparticles. Nanoparticle containment has various benefits, including environmental protection, precise release dynamics, and the opportunity to integrate other bioactive substances for synergistic effects. Without a doubt, nanoparticles also have drawbacks that need to be addressed to improve acceptability. Furthermore, the nanoscale dimension of these delivery methods facilitates effective cellular absorption and transit beyond biological barriers, resulting in better therapeutic effects.

Nonetheless, additional investigation is needed to optimize the formulation characteristics of nanoparticles, such as structure, dimension, surface characteristics, and encapsulation efficacy, to maximize the bioavailability and effectiveness of omega-3s. Furthermore, long-term stability trials and in vivo tests are required to evaluate the safety, pharmaceutical kinetics, and medicinal potential of nanoparticle-based omega-3 nanoparticles in a variety of clinical contexts. In summary, applying nanoparticles to encapsulate omega-3 fatty acids shows considerable potential for resolving present problems in omega-3 supplementation, while opening up new options for personalized nutrition and preventative healthcare.

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References

- 1. Tsao, C.W.; Aday, A.W.; Almarzooq, Z.I.; Anderson, C.A.M.; Arora, P.; Avery, C.L.; Baker-Smith, C.M.; Beaton, A.Z.; Boehme, A.K.; Buxton, A.E.; et al. Heart Disease and Stroke Statistics—2023 Update: A Report from the American Heart Association. *Circulation* **2023**, *147*, e153–e639. [\[CrossRef\]](https://doi.org/10.1161/CIR.0000000000001123) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36695182)
- 2. National Center for Health Statistics. Multiple Cause of Death Data on CDC WONDER. 2023. Available online: [https://wonder.](https://wonder.cdc.gov/mcd.html) [cdc.gov/mcd.html](https://wonder.cdc.gov/mcd.html) (accessed on 2 January 2024).
- 3. World Health Organization. Cardiovascular Diseases (CVDs). 2021. Available online: [https://www.who.int/news-room/fact](https://www.who.int/news-room/fact-sheets/detail/cardiovascular-diseases-(cvds))[sheets/detail/cardiovascular-diseases-\(cvds\)](https://www.who.int/news-room/fact-sheets/detail/cardiovascular-diseases-(cvds)) (accessed on 2 January 2024).
- 4. Centers for Disease Control and Prevention. Heart Disease and Stroke. 2020. Available online: [https://www.cdc.gov/heart](https://www.cdc.gov/heart-disease/data-research/facts-stats/index.html)[disease/data-research/facts-stats/index.html](https://www.cdc.gov/heart-disease/data-research/facts-stats/index.html) (accessed on 2 January 2024).
- 5. American Heart Association. Consuming about 3 Grams of Omega-3 Fatty Acids a Day May Lower Blood Pressure. Available online: [https://www.heart.org/en/news/2022/06/01/consuming-about-3-grams-of-omega-3-fatty-acids-a-day-may-lower](https://www.heart.org/en/news/2022/06/01/consuming-about-3-grams-of-omega-3-fatty-acids-a-day-may-lower-blood-pressure)[blood-pressure](https://www.heart.org/en/news/2022/06/01/consuming-about-3-grams-of-omega-3-fatty-acids-a-day-may-lower-blood-pressure) (accessed on 2 January 2024).
- 6. von Schacky, C.; Harris, W. Cardiovascular benefits of omega-3 fatty acids. *Cardiovasc. Res.* **2007**, *73*, 310–315. [\[CrossRef\]](https://doi.org/10.1016/j.cardiores.2006.08.019) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16979604)
- 7. Higdon, J. Essential Fatty Acids, Micronutrient Information Center 179. 2019. Available online: [https://lpi.oregonstate.edu/mic/](https://lpi.oregonstate.edu/mic/other-nutrients/essential-fatty-acids) [other-nutrients/essential-fatty-acids](https://lpi.oregonstate.edu/mic/other-nutrients/essential-fatty-acids) (accessed on 2 January 2024).
- 8. Ulven, S.M.; Kirkhus, B.; Lamglait, A.; Basu, S.; Elind, E.; Haider, T.; Berge, K.; Vik, H.; Pedersen, J.I. Metabolic Effects of Krill Oil are Essentially Similar to Those of Fish Oil but at Lower Doses of EPA and DHA, in Healthy Volunteers. *Lipids* **2010**, *46*, 37–46. [\[CrossRef\]](https://doi.org/10.1007/s11745-010-3490-4) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21042875)
- 9. Gammone, M.; Riccioni, G.; Parrinello, G.; D'Orazio, N. Omega-3 Polyunsaturated Fatty Acids: Benefits and Endpoints in Sport. *Nutrients* **2018**, *11*, 46. [\[CrossRef\]](https://doi.org/10.3390/nu11010046) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30591639)
- 10. Østbye, T.-K.K.; Gudbrandsen, O.A.; Drotningsvik, A.; Ruyter, B.; Berge, G.M.; Vogt, G.; Nilsson, A. Different Dietary Ratios of Camelina Oil to Sandeel Oil Influence the Capacity to Synthesise and Deposit EPA and DHA in Zucker Fa/Fa Rats. *Nutrients* **2023**, *15*, 2344. [\[CrossRef\]](https://doi.org/10.3390/nu15102344) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37242227)
- 11. Halade, G.V.; Rahman, M.M.; Bhattacharya, A.; Barnes, J.L.; Chandrasekar, B.; Fernandes, G. Docosahexaenoic Acid-Enriched Fish Oil Attenuates Kidney Disease and Prolongs Median and Maximal Life Span of Autoimmune Lupus-Prone Mice. *J. Immunol.* **2010**, *184*, 5280–5286. [\[CrossRef\]](https://doi.org/10.4049/jimmunol.0903282) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20368275)
- 12. Fernandes, G.; Bhattacharya, A.; Rahman, M.; Zaman, K.; Banu, J. Effects of n-3 fatty acids on autoimmunity and osteoporosis. *Front. Biosci.* **2008**, *13*, 4015–4020. [\[CrossRef\]](https://doi.org/10.2741/2989) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18508495)
- 13. Bentsen, H. Dietary polyunsaturated fatty acids, brain function, and mental health. *Microb. Ecol. Health Dis.* **2017**, *28* (Suppl. 1), 1281916. [\[CrossRef\]](https://doi.org/10.1080/16512235.2017.1281916)
- 14. Rennie, K.L.; Hughes, J.; Lang, R.; Jebb, S.A. Nutritional management of rheumatoid arthritis: A review of the evidence. *J. Hum. Nutr. Diet.* **2003**, *16*, 97–109. [\[CrossRef\]](https://doi.org/10.1046/j.1365-277x.2003.00423.x)
- 15. Bhattacharya, A.; Rahman, M.M.; Banu, J.; Lawrence, R.; McGuff, H.S.; Garrett, I.R.; Fischbach, M.; Fernandes, G. Inhibition of Osteoporosis in Autoimmune Disease Prone MRL/Mpj-FaslprMice by N-3 Fatty Acids. *J. Am. Coll. Nutr.* **2005**, *24*, 200–209. [\[CrossRef\]](https://doi.org/10.1080/07315724.2005.10719466)
- 16. Abou-Saleh, H.; Ouhtit, A.; Halade, G.V.; Rahman, M.M. Bone Benefits of Fish Oil Supplementation Depend on its EPA and DHA Content. *Nutrients* **2019**, *11*, 2701. [\[CrossRef\]](https://doi.org/10.3390/nu11112701) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31717258)
- 17. Holm, T. Omega-3 fatty acids improve blood pressure control and preserve renal function in hypertensive heart transplant recipients. *Eur. Heart J.* **2001**, *22*, 428–436. [\[CrossRef\]](https://doi.org/10.1053/euhj.2000.2369) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11207085)
- 18. Freeman, M.P. Omega-3 Fatty Acids in Psychiatry: A Review. *Ann. Clin. Psychiatry* **2000**, *12*, 159–165. [\[CrossRef\]](https://doi.org/10.1023/A:1009069002816) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/10984006)
- 19. Yaghmur, A.; Ghayas, S.; Jan, H.; Kalaycioglu, G.D.; Moghimi, S.M. Omega-3 fatty acid nanocarriers: Characterization and potential applications. *Curr. Opin. Colloid Interface Sci.* **2023**, *67*, 101728. [\[CrossRef\]](https://doi.org/10.1016/j.cocis.2023.101728)
- 20. Bhattacharya, A.; Sun, D.; Rahman, M.; Fernandes, G. Different ratios of eicosapentaenoic and docosahexaenoic omega-3 fatty acids in commercial fish oils differentially alter pro-inflammatory cytokines in peritoneal macrophages from C57BL/6 female mice. *J. Nutr. Biochem.* **2007**, *18*, 23–30. [\[CrossRef\]](https://doi.org/10.1016/j.jnutbio.2006.02.005) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16563716)
- 21. Kesavalu, L.; Bakthavatchalu, V.; Rahman, M.M.; Su, J.; Raghu, B.; Dawson, D.; Fernandes, G.; Ebersole, J.L. Omega-3 fatty acid regulates inflammatory cytokine/mediator messenger RNA expression in Porphyromonas gingivalis-induced experimental periodontal disease. *Oral Microbiol. Immunol.* **2007**, *22*, 232–239. [\[CrossRef\]](https://doi.org/10.1111/j.1399-302x.2007.00346.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17600534)
- 22. Rahman, M.M.; Veigas, J.M.; Williams, P.J.; Fernandes, G. DHA is a more potent inhibitor of breast cancer metastasis to bone and related osteolysis than EPA. *Breast Cancer Res. Treat.* **2013**, *141*, 341–352. [\[CrossRef\]](https://doi.org/10.1007/s10549-013-2703-y) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24062211)
- 23. Crovella, S.; Ouhtit, A.; Rahman, S.M.; Rahman, M.M. Docosahexaenoic Acid, a Key Compound for Enhancing Sensitization to Drug in Doxorubicin-Resistant MCF-7 Cell Line. *Nutrients* **2023**, *15*, 1658. [\[CrossRef\]](https://doi.org/10.3390/nu15071658)
- 24. McLennan, P.L.; Bridle, T.M.; Abeywardena, M.Y.; Charnock, J.S. Dietary lipid modulation of ventricular fibrillation threshold in the marmoset monkey. *Am. Heart J.* **1992**, *123*, 1555–1561. [\[CrossRef\]](https://doi.org/10.1016/0002-8703(92)90809-a)
- 25. Madingou, N.; Gilbert, K.; Tomaro, L.; Touchette, C.P.H.; Trudeau, F.; Fortin, S.; Rousseau, G. Comparison of the effects of EPA and DHA alone or in combination in a murine model of myocardial infarction. *Prostaglandins Leukot. Essent. Fat. Acids/Prostaglandins Leukot. Essent. Fat. Acids* **2016**, *111*, 11–16. [\[CrossRef\]](https://doi.org/10.1016/j.plefa.2016.06.001)
- 26. Baum, J.R.; Dolmatova, E.; Tan, A.; Duffy, H.S. Omega 3 fatty acid inhibition of inflammatory cytokine-mediated Connexin43 regulation in the heart. *Front. Physiol.* **2012**, *3*, 272. [\[CrossRef\]](https://doi.org/10.3389/fphys.2012.00272)
- 27. Kalish, B.T.; Matte, A.; Andolfo, I.; Iolascon, A.; Weinberg, O.; Ghigo, A.; Cimino, J.; Siciliano, A.; Hirsch, E.; Federti, E.; et al. Dietary ω-3 fatty acids protect against vasculopathy in a transgenic mouse model of sickle cell disease. *Haematologica* **2015**, *100*, 870–880. [\[CrossRef\]](https://doi.org/10.3324/haematol.2015.124586) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25934765)
- 28. Angelotti, A.; Snoke, D.B.; Ormiston, K.; Cole, R.M.; Borkowski, K.; Newman, J.W.; Orchard, T.S.; Belury, M.A. Potential Cardioprotective Effects and Lipid Mediator Differences in Long-Chain Omega-3 Polyunsaturated Fatty Acid Supplemented Mice Given Chemotherapy. *Metabolites* **2022**, *12*, 782. [\[CrossRef\]](https://doi.org/10.3390/metabo12090782) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36144189)
- 29. Burr, M.L.; Gilbert, J.F.; Holliday, R.M.; Elwood, P.C.; Fehily, A.M.; Rogers, S.; Sweetnam, P.M.; Deadman, N.M. Effects of changes in fat, fish, and fiber intakes on death and myocardial reinfarction: Diet and reinfarction trial (DART). *Lancet* **1989**, *334*, 757–761. [\[CrossRef\]](https://doi.org/10.1016/s0140-6736(89)90828-3)
- 30. Jialal, I.; Devaraj, S.; Huet, B.; Traber, M. GISSI-Prevenzione trial. *Lancet* **1999**, *354*, 1554. [\[CrossRef\]](https://doi.org/10.1016/s0140-6736(99)90191-5)
- 31. Yokoyama, M.; Origasa, H.; Matsuzaki, M.; Matsuzawa, Y.; Saito, Y.; Ishikawa, Y.; Oikawa, S.; Sasaki, J.; Hishida, H.; Itakura, H.; et al. Effects of eicosapentaenoic acid on major coronary events in hypercholesterolaemic patients (JELIS): A randomized open-label, blinded endpoint analysis. *Lancet* **2007**, *369*, 1090–1098. [\[CrossRef\]](https://doi.org/10.1016/s0140-6736(07)60527-3) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17398308)
- 32. Einvik, G.; Ole Klemsdal, T.; Sandvik, L.; Hjerkinn, E.M. A randomized clinical trial on n-3 polyunsaturated fatty acids supplementation and all-cause mortality in elderly men at high cardiovascular risk. *Eur. J. Cardiovasc. Prev. Rehabil.* **2010**, *17*, 588–592. [\[CrossRef\]](https://doi.org/10.1097/hjr.0b013e328339cc70)
- 33. Rauch, B.; Schiele, R.; Schneider, S.; Diller, F.; Victor, N.; Gohlke, H.; Gottwik, M.; Steinbeck, G.; Del Castillo, U.; Sack, R.; et al. OMEGA, a Randomized, Placebo-Controlled Trial to Test the Effect of Highly Purified Omega-3 Fatty Acids on Top of Modern Guideline-Adjusted Therapy After Myocardial Infarction. *Circulation* **2010**, *122*, 2152–2159. [\[CrossRef\]](https://doi.org/10.1161/circulationaha.110.948562)
- 34. Kromhout, D.; Giltay, E.J.; Geleijnse, J.M. n–3 Fatty Acids and Cardiovascular Events after Myocardial Infarction. *N. Engl. J. Med.* **2010**, *363*, 2015–2026. [\[CrossRef\]](https://doi.org/10.1056/nejmoa1003603)
- 35. Galan, P.; Kesse-Guyot, E.; Czernichow, S.; Briancon, S.; Blacher, J.; Hercberg, S. Effects of B vitamins and omega 3 fatty acids on cardiovascular diseases: A randomized placebo-controlled trial. *BMJ* **2010**, *341*, c6273. [\[CrossRef\]](https://doi.org/10.1136/bmj.c6273)
- 36. Bowman, L.; Mafham, M.; Stevens, W.; Haynes, R.; Aung, T.; Chen, F.; Buck, G.; Collins, R.; Armitage, J.; ASCEND Study Collaborative Group. ASCEND: A Study of Cardiovascular Events in Diabetes: Characteristics of a randomized trial of aspirin and of omega-3 fatty acid supplementation in 15,480 people with diabetes. *Am. Heart J.* **2018**, *198*, 135–144. [\[CrossRef\]](https://doi.org/10.1016/j.ahj.2017.12.006) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29653635)
- 37. Manson, J.E.; Cook, N.R.; Lee, I.-M.; Christen, W.; Bassuk, S.S.; Mora, S.; Gibson, H.; Albert, C.M.; Gordon, D.; Copeland, T.; et al. Marine n−3 Fatty Acids and Prevention of Cardiovascular Disease and Cancer. *N. Engl. J. Med.* **2019**, *380*, 23–32. [\[CrossRef\]](https://doi.org/10.1056/nejmoa1811403) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30415637)
- 38. Bhatt, D.L.; Steg, P.G.; Miller, M.; Brinton, E.A.; Jacobson, T.A.; Ketchum, S.B.; Doyle, R.T.; Juliano, R.A.; Jiao, L.; Granowitz, C.; et al. Cardiovascular Risk Reduction with Icosapent Ethyl for Hypertriglyceridemia. *N. Engl. J. Med.* **2019**, *380*, 11–22. [\[CrossRef\]](https://doi.org/10.1056/nejmoa1812792) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30415628)
- 39. Nicholls, S.J.; Lincoff, A.M.; Garcia, M.; Bash, D.; Ballantyne, C.M.; Barter, P.J.; Davidson, M.H.; Kastelein, J.J.P.; Koenig, W.; McGuire, D.K.; et al. Effect of High-Dose Omega-3 Fatty Acids vs Corn Oil on Major Adverse Cardiovascular Events in Patients at High Cardiovascular Risk. *JAMA* **2020**, *324*, 2268. [\[CrossRef\]](https://doi.org/10.1001/jama.2020.22258) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33190147)
- 40. Kalstad, A.A.; Myhre, P.L.; Laake, K.; Tveit, S.H.; Schmidt, E.B.; Smith, P.; Nilsen, D.W.T.; Tveit, A.; Fagerland, M.W.; Solheim, S.; et al. Effects of n-3 Fatty Acid Supplements in Elderly Patients After Myocardial Infarction. *Circulation* **2021**, *143*, 528–539. [\[CrossRef\]](https://doi.org/10.1161/circulationaha.120.052209) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33191772)
- 41. Welty, F.; Bistrian, B.; Driscoll, D. Omega-3 Fatty Acids Effect on Major Cardiovascular Events in Patients at High Cardiovascular Risk. *JAMA* **2021**, *325*, 1333. [\[CrossRef\]](https://doi.org/10.1001/jama.2021.0830)
- 42. Gillum, R.F.; Mussolino, M.; Madans, J.H. The relation between fish consumption, death from all causes, and incidence of coronary heart disease. *J. Clin. Epidemiol.* **2000**, *53*, 237–244. [\[CrossRef\]](https://doi.org/10.1016/s0895-4356(99)00149-3) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/10760632)
- 43. Iso, H.; Kobayashi, M.; Ishihara, J.; Sasaki, S.; Okada, K.; Kita, Y.; Kokubo, Y.; Tsugane, S. Intake of Fish and n3 Fatty Acids and Risk of Coronary Heart Disease Among Japanese. *Circulation* **2006**, *113*, 195–202. [\[CrossRef\]](https://doi.org/10.1161/circulationaha.105.581355)
- 44. Kühn, T.; Teucher, B.; Kaaks, R.; Boeing, H.; Weikert, C.; Buijsse, B. Fish consumption and the risk of myocardial infarction and stroke in the German arm of the European Prospective Investigation into Cancer and Nutrition (EPIC-Germany). *Br. J. Nutr.* **2013**, *110*, 1118–1125. [\[CrossRef\]](https://doi.org/10.1017/s0007114513000202)
- 45. Nahab, F.; Pearson, K.; Frankel, M.R.; Ard, J.; Safford, M.M.; Kleindorfer, D.; Howard, V.J.; Judd, S. Dietary fried fish intake increases the risk of CVD: The reasons for Geographic and Racial Differences in Stroke (REGARDS) study. *Public Health Nutr.* **2016**, *19*, 3327–3336. [\[CrossRef\]](https://doi.org/10.1017/s136898001600152x)
- 46. Bonaccio, M.; Ruggiero, E.; Di Castelnuovo, A.; Costanzo, S.; Persichillo, M.; De Curtis, A.; Cerletti, C.; Donati, M.B.; de Gaetano, G.; Iacoviello, L.; et al. Fish intake is associated with lower cardiovascular risk in a Mediterranean population: Prospective results from the Moli-sani study. *Nutr. Metab. Cardiovasc. Dis.* **2017**, *27*, 865–873. [\[CrossRef\]](https://doi.org/10.1016/j.numecd.2017.08.004) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28967596)
- 47. Hengeveld, L.M.; Praagman, J.; Beulens, J.W.J.; Brouwer, I.A.; van der Schouw, Y.T.; Sluijs, I. Fish consumption and risk of stroke, coronary heart disease, and cardiovascular mortality in a Dutch population with low fish intake. *Eur. J. Clin. Nutr.* **2018**, *72*, 942–950. [\[CrossRef\]](https://doi.org/10.1038/s41430-018-0190-2)
- 48. Ward, R.E.; Cho, K.; Nguyen, X.M.T.; Vassy, J.L.; Ho, Y.L.; Quaden, R.M.; Gagnon, D.R.; Wilson, P.W.F.; Gaziano, J.M.; Djoussé, L. Omega-3 supplement use, fish intake, and risk of non-fatal coronary artery disease and ischemic stroke in the Million Veteran Program. *Clin. Nutr.* **2020**, *39*, 574–579. [\[CrossRef\]](https://doi.org/10.1016/j.clnu.2019.03.005)
- 49. Pertiwi, K.; Küpers, L.K.; de Goede, J.; Zock, P.L.; Kromhout, D.; Geleijnse, J.M. Dietary and Circulating Long-Chain Omega-3 Polyunsaturated Fatty Acids and Mortality Risk After Myocardial Infarction: A Long-Term Follow-Up of the Alpha Omega Cohort. *J. Am. Heart Assoc.* **2021**, *10*, e022617. [\[CrossRef\]](https://doi.org/10.1161/jaha.121.022617)
- 50. Harris, W.S.; von Schacky, C. The Omega-3 Index: A new risk factor for death from coronary heart disease? *Prev. Med.* **2004**, *39*, 212–220. [\[CrossRef\]](https://doi.org/10.1016/j.ypmed.2004.02.030)
- 51. Harris, W. Omega-3 fatty acids and cardiovascular disease: A case for omega-3 index as a new risk factor. *Pharmacol. Res.* **2007**, *55*, 217–223. [\[CrossRef\]](https://doi.org/10.1016/j.phrs.2007.01.013) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17324586)
- 52. Harris, W.S.; Thomas, R.M. Biological variability of blood omega-3 biomarkers. *Clin. Biochem.* **2010**, *43*, 338–340. [\[CrossRef\]](https://doi.org/10.1016/j.clinbiochem.2009.08.016) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/19733159)
- 53. Gramenzi, A.; Gentile, A.; Fasoli, M.; Negri, E.; Parazzini, F.; La Vecchia, C. Association between certain foods and risk of acute myocardial infarction in women. *BMJ* **1990**, *300*, 771–773. [\[CrossRef\]](https://doi.org/10.1136/bmj.300.6727.771)
- 54. Siscovick, D.S. Dietary Intake and Cell Membrane Levels of Long-Chain n-3 Polyunsaturated Fatty Acids and the Risk of Primary Cardiac Arrest. *JAMA J. Am. Med. Assoc.* **1995**, *274*, 1363. [\[CrossRef\]](https://doi.org/10.1001/jama.1995.03530170043030)
- 55. Hallgren, C.G.; Göran Hallmans Jansson, J.O.; Marklund, S.L.; Huhtasaari, F.; Schütz, A.; Strömberg, U.; Vessby, B.; Skerfving, S. Markers of high fish intake are associated with a decreased risk of a first myocardial infarction. *Br. J. Nutr.* **2001**, *86*, 397–404. [\[CrossRef\]](https://doi.org/10.1079/bjn2001415)
- 56. Panagiotakos, D.B.; Pitsavos, C.; Zampelas, A.; Chrysohoou, C.; Griffin, B.A.; Stefanadis, C.; Toutouzas, P. Fish consumption and the risk of developing acute coronary syndromes: The CARDIO2000 study. *Int. J. Cardiol.* **2005**, *102*, 403–409. [\[CrossRef\]](https://doi.org/10.1016/j.ijcard.2004.05.043) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16004884)
- 57. Amani, R.; Noorizadeh, M.; Rahmanian, S.; Afzali, N.; Haghighizadeh, M.H. Nutritional related cardiovascular risk factors in patients with coronary artery disease in IRAN: A case-control study. *Nutr. J.* **2010**, *9*, 70. [\[CrossRef\]](https://doi.org/10.1186/1475-2891-9-70) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21184687)
- 58. Venegas-Calerón, M.; Napier, J.A. New alternative sources of omega-3 fish oil. *Adv. Food Nutr. Res.* **2023**, *1*, 343–398.
- 59. Tur, J.A.; Bibiloni, M.M.; Sureda, A.; Pons, A. Dietary sources of omega 3 fatty acids: Public health risks and benefits. *Br. J. Nutr.* **2012**, *107* (Suppl. 2), S23–S52. [\[CrossRef\]](https://doi.org/10.1017/s0007114512001456) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22591897)
- 60. Cholewski, M.; Tomczykowa, M.; Tomczyk, M. A Comprehensive Review of Chemistry, Sources and Bioavailability of Omega-3 Fatty Acids. *Nutrients* **2018**, *10*, 1662. [\[CrossRef\]](https://doi.org/10.3390/nu10111662) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30400360)
- 61. Khan, K.U.; Minhas, M.U.; Badshah, S.F.; Suhail, M.; Ahmad, A.; Ijaz, S. Overview of nanoparticulate strategies for solubility enhancement of poorly soluble drugs. *Life Sci.* **2022**, *291*, 120301. [\[CrossRef\]](https://doi.org/10.1016/j.lfs.2022.120301) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34999114)
- 62. Borges, A.; de Freitas, V.; Mateus, N.; Fernandes, I.; Oliveira, J. Solid Lipid Nanoparticles as Carriers of Natural Phenolic Compounds. *Antioxidants* **2020**, *9*, 998. [\[CrossRef\]](https://doi.org/10.3390/antiox9100998) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33076501)
- 63. Lamson, N.G.; Berger, A.; Fein, K.C.; Whitehead, K.A. Anionic nanoparticles enable the oral delivery of proteins by enhancing intestinal permeability. *Nat. Biomed. Eng.* **2019**, *4*, 84–96. [\[CrossRef\]](https://doi.org/10.1038/s41551-019-0465-5)
- 64. Tekie, F.S.M.; Hajiramezanali, M.; Geramifar, P.; Raoufi, M.; Dinarvand, R.; Soleimani, M.; Atyabi, F. Controlling evolution of protein corona: A prosperous approach to improve chitosan-based nanoparticle biodistribution and half-life. *Sci. Rep.* **2020**, *10*, 9664. [\[CrossRef\]](https://doi.org/10.1038/s41598-020-66572-y)
- 65. McClements, D.J. Advances in edible nanoemulsions: Digestion, bioavailability, and potential toxicity. *Prog. Lipid Res.* **2021**, *81*, 101081. [\[CrossRef\]](https://doi.org/10.1016/j.plipres.2020.101081)
- 66. Schuchardt, J.P.; Hahn, A. Bioavailability of long-chain omega-3 fatty acids. *Prostaglandins Leukot. Essent. Fat. Acids* **2013**, *89*, 1–8. [\[CrossRef\]](https://doi.org/10.1016/j.plefa.2013.03.010) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23676322)
- 67. Li, J.; Pora, B.L.R.; Dong, K.; Hasjim, J. Health benefits of docosahexaenoic acid and its bioavailability: A review. *Food Sci. Nutr.* **2021**, *9*, 5229–5243. [\[CrossRef\]](https://doi.org/10.1002/fsn3.2299) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34532031)
- 68. Homroy, S.; Chopra, R.N.; Singh, P.K.; Dhiman, A.; Rama, S.; Talwar, B. Role of encapsulation on the bioavailability of omega-3 fatty acids. *Compr. Rev. Food Sci. Food Saf.* **2023**, *23*, e13272. [\[CrossRef\]](https://doi.org/10.1111/1541-4337.13272) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38284597)
- 69. Dacaranhe, C.D.; Terao, J. Effect of Phosphatidic Acid and Phosphatidylserine on Lipid Oxidation in Beef Homogenate During Storage and in Emulsified Sardine Oil. *J. Food Sci.* **2001**, *66*, 422–427. [\[CrossRef\]](https://doi.org/10.1111/j.1365-2621.2001.tb16121.x)
- 70. Arab-Tehrany, E.; Jacquot, M.; Gaiani, C.; Imran, M.; Desobry, S.; Linder, M. Beneficial effects and oxidative stability of omega-3 long-chain polyunsaturated fatty acids. *Trends Food Sci. Technol.* **2012**, *25*, 24–33. [\[CrossRef\]](https://doi.org/10.1016/j.tifs.2011.12.002)
- 71. Nair, V.; Cooper, C.S.; Vietti, D.E.; Turner, G.A. The chemistry of lipid peroxidation metabolites: Crosslinking reactions of malondialdehyde. *Lipids* **1986**, *21*, 6–10. [\[CrossRef\]](https://doi.org/10.1007/bf02534294) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/3959768)
- 72. Jacobsen, C.; Hartvigsen, K.; Lund, P.; Meyer, A.S.; Nissen, J.A.; Holstborg, J.; Hølmer, G. Oxidation in fish-oil-enriched mayonnaise. *Eur. Food Res. Technol.* **1999**, *210*, 13–30. [\[CrossRef\]](https://doi.org/10.1007/s002170050526)
- 73. Jacobsen, C.; Hartvigsen, K.; Lund, P.; Thomsen, M.; Skibsted, L.H.; Hølmer, G.; Adler-Nissen, J.; Meyer, A.S. Oxidation in fish oil-enriched mayonnaise: 4. Effect of tocopherol concentration on oxidative deterioration. *Eur. Food Res. Technol.* **2001**, *212*, 308–318. [\[CrossRef\]](https://doi.org/10.1007/s002170000251)
- 74. Omer, A.M.; Ziora, Z.M.; Tamer, T.M.; Khalifa, R.E.; Hassan, M.A.; Mohy-Eldin, M.S.; Blaskovich, M.A.T. Formulation of Quaternized Aminated Chitosan Nanoparticles for Efficient Encapsulation and Slow Release of Curcumin. *Molecules* **2021**, *26*, 449. [\[CrossRef\]](https://doi.org/10.3390/molecules26020449)
- 75. Mohamed, N.A.; Saleh, H.A.; Kameno, Y.; Marei, I.; de Nucci, G.; Ahmetaj-Shala, B.; Shala, F.; Kirkby, N.S.; Jennings, L.; Davies, R.P.; et al. Metal-organic framework (MOF) nanomedicine preparations of sildenafil designed for the future treatment of pulmonary arterial hypertension. *bioRxiv* **2019**. [\[CrossRef\]](https://doi.org/10.1101/718478)
- 76. Mohamed, H.; Mohamed, N.; Macasa, S.; Basha, H.; Adan, A.; Marei, I.; Ding, H.; Triggle, C.; Crovella, S.; Abou-Saleh, H. Managing diabetes with nanomedicine: nanoMIL-89 as a promising drug delivery system for metformin. *Res. Sq.* 2024, *preprint*. [\[CrossRef\]](https://doi.org/10.21203/rs.3.rs-3893992/v1)
- 77. Mohamed, N.A.; Marei, I.; Crovella, S.; Abou-Saleh, H. Recent Developments in Nanomaterials-Based Drug Delivery and Upgrading Treatment of Cardiovascular Diseases. *Int. J. Mol. Sci.* **2022**, *23*, 1404. [\[CrossRef\]](https://doi.org/10.3390/ijms23031404) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35163328)
- 78. De Jong, W.H.; Borm, P.J. Drug delivery and nanoparticles: Applications and hazards. *Int. J. Nanomed.* **2008**, *3*, 133. [\[CrossRef\]](https://doi.org/10.2147/ijn.s596) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18686775)
- 79. Preethi, R.; Dutta, S.; Moses, J.A.; Anandharamakrishnan, C. *Green Nanomaterials and Nanotechnology for the Food Industry*; Elsevier: Amsterdam, The Netherlands, 2022; pp. 215–256. [\[CrossRef\]](https://doi.org/10.1016/b978-0-12-823137-1.00008-7)
- 80. Khalid, M.; El-Sawy, H.S. Polymeric nanoparticles: Promising platform for drug delivery. *Int. J. Pharm.* **2017**, *528*, 675–691. [\[CrossRef\]](https://doi.org/10.1016/j.ijpharm.2017.06.052) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28629982)
- 81. Kahraman, E.; Güngör, S.; Özsoy, Y. Potential enhancement and targeting strategies of polymeric and lipid-based nanocarriers in dermal drug delivery. *Ther. Deliv.* **2017**, *8*, 967–985. [\[CrossRef\]](https://doi.org/10.4155/tde-2017-0075) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29061106)
- 82. Klinkova, A.; Thérien-Aubin, H. Chapter 6—Polymer nanoparticles. In *Nanochemistry: Chemistry of Nanoparticle Formation and Interactions*; Klinkova, A., Thérien-Aubin, H., Eds.; Elsevier: Amsterdam, The Netherlands, 2024; pp. 167–215. Available online: <https://www.sciencedirect.com/science/article/abs/pii/B9780443214479000023> (accessed on 18 March 2024).
- 83. Perumal, S.; Atchudan, R.; Lee, W. A Review of Polymeric Micelles and Their Applications. *Polymers* **2022**, *14*, 2510. [\[CrossRef\]](https://doi.org/10.3390/polym14122510) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35746086)
- 84. Wakaskar, R.R. Polymeric Micelles and their Properties. *J. Nanomed. Nanotechnol.* **2017**, *8*, 433. [\[CrossRef\]](https://doi.org/10.4172/2157-7439.1000433)
- 85. Mittal, P.; Saharan, A.; Verma, R.; Altalbawy, F.M.A.; Alfaidi, M.A.; Batiha, G.E.S.; Akter, W.; Gautam, R.K.; Uddin, M.S.; Rahman, M.S. Dendrimers: A New Race of Pharmaceutical Nanocarriers. *BioMed Res. Int.* **2021**, *2021*, e8844030. [\[CrossRef\]](https://doi.org/10.1155/2021/8844030)
- 86. Chis, A.A.; Dobrea, C.; Morgovan, C.; Arseniu, A.M.; Rus, L.L.; Butuca, A.; Juncan, A.M.; Totan, M.; Vonica-Tincu, A.L.; Cormos, G.; et al. Applications and Limitations of Dendrimers in Biomedicine. *Molecules* **2020**, *25*, 3982. [\[CrossRef\]](https://doi.org/10.3390/molecules25173982)
- 87. Santos, A.; Veiga, F.; Figueiras, A. Dendrimers as Pharmaceutical Excipients: Synthesis, Properties, Toxicity and Biomedical Applications. *Materials* **2019**, *13*, 65. [\[CrossRef\]](https://doi.org/10.3390/ma13010065)
- 88. Cavalli, R.; Caputo, O.; Gasco, M.R. Solid lipospheres of doxorubicin and idarubicin. *Int. J. Pharm.* **1993**, *89*, R9–R12. [\[CrossRef\]](https://doi.org/10.1016/0378-5173(93)90313-5)
- 89. Nguyen, T.T.L.; Duong, V.A. Solid Lipid Nanoparticles. *Encyclopedia* **2022**, *2*, 952–973. [\[CrossRef\]](https://doi.org/10.3390/encyclopedia2020063)
- 90. Jha, S.; KSharma, P.; Malviya, R. Liposomal Drug Delivery System for Cancer Therapy: Advancement and Patents. *Recent Pat. Drug Deliv. Formul.* **2016**, *10*, 177–183. [\[CrossRef\]](https://doi.org/10.2174/1872211310666161004155757) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27712569)
- 91. Akbarzadeh, A.; Rezaei-Sadabady, R.; Davaran, S.; Joo, S.W.; Zarghami, N.; Hanifehpour, Y.; Samiei, M.; Kouhi, M.; Nejati-Koshki, K. Liposome: Classification, preparation, and applications. *Nanoscale Res. Lett.* **2013**, *8*, 102. [\[CrossRef\]](https://doi.org/10.1186/1556-276x-8-102) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23432972)
- 92. Gupta, A.; Eral, H.B.; Hatton, T.A.; Doyle, P.S. Nanoemulsions: Formation, properties and applications. *Soft Matter* **2016**, *12*, 2826–2841. [\[CrossRef\]](https://doi.org/10.1016/b978-0-12-818392-2.00007-x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26924445)
- 93. Bhosale, R.R.; Osmani, R.A.; Ghodake, P.P.; Shaikh, S.M.; Chavan, S.R. Nanoemulsion: A Review on Novel Profusion in Advanced Drug Delivery. *Indian J. Pharm. Biol. Res.* **2014**, *2*, 122. [\[CrossRef\]](https://doi.org/10.30750/ijpbr.2.1.19)
- 94. Yeh, Y.C.; Creran, B.; Rotello, V.M. Gold Nanoparticles: Preparation, Properties, and Applications in Bionanotechnology. *Nanoscale* **2012**, *4*, 1871–1880. [\[CrossRef\]](https://doi.org/10.1039/c1nr11188d) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22076024)
- 95. Kesharwani, P.; Ma, R.; Liang, S.; Fatima, M.; Sheikh, A.; Abourehab, M.A.S.; Gupta, N.; Chen, Z.-S.; Zhou, Y. Gold nanoparticles and gold nanorods in the landscape of cancer therapy. *Mol. Cancer* **2023**, *22*, 98. [\[CrossRef\]](https://doi.org/10.1186/s12943-023-01798-8)
- 96. Almatroudi, A. Silver nanoparticles: Synthesis, characterization and biomedical applications. *Open Life Sci.* **2020**, *15*, 819–839. [\[CrossRef\]](https://doi.org/10.1515/biol-2020-0094)
- 97. Ferdous, Z.; Nemmar, A. Health Impact of Silver Nanoparticles: A Review of the Biodistribution and Toxicity Following Various Routes of Exposure. *Int. J. Mol. Sci.* **2020**, *21*, 2375. [\[CrossRef\]](https://doi.org/10.3390/ijms21072375)
- 98. Ali, A.; Zafar, H.; Zia, M.; ul Haq, I.; Phull, A.R.; Ali, J.S.; Hussain, A. Synthesis, characterization, applications, and challenges of iron oxide nanoparticles. *Nanotechnol. Sci. Appl.* **2016**, *9*, 49–67. [\[CrossRef\]](https://doi.org/10.2147/nsa.s99986)
- 99. Laurent, S.; Forge, D.; Port, M.; Roch, A.; Robic, C.; Vander Elst, L.; Muller, R.N. Magnetic Iron Oxide Nanoparticles: Synthesis, Stabilization, Vectorization, Physicochemical Characterizations, and Biological Applications. *Chem. Rev.* **2008**, *108*, 2064–2110. [\[CrossRef\]](https://doi.org/10.1021/cr068445e) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18543879)
- 100. Attia, N.F.; El-Monaem, E.M.A.; El-Aqapa, H.G.; Elashery, S.E.A.; Eltaweil, A.S.; El Kady, M.; Khalifa, S.A.M.; Hawash, H.B.; El-Seedi, H.R. Iron oxide nanoparticles and their pharmaceutical applications. *Appl. Surf. Sci. Adv.* **2022**, *11*, 100284. [\[CrossRef\]](https://doi.org/10.1016/j.apsadv.2022.100284)
- 101. Sahoo, J.K.; Sahoo, S.K. *Applications of Magnetic Nanocomposites in Wastewater Treatment*; Elsevier: Amsterdam, The Netherlands, 2023; pp. 47–63. [\[CrossRef\]](https://doi.org/10.1016/b978-0-323-99344-9.00011-6)
- 102. Leong, K.H.; Chin, Y.H.; Sim, L.C.; Tan, B.; Dai, C.; Saravanan, P. *Physical Properties of Quantum Dots*; Elsevier: Amsterdam, The Netherlands, 2022; pp. 687–709. [\[CrossRef\]](https://doi.org/10.1016/b978-0-323-85457-3.00017-7)
- 103. Drbohlavova, J.; Adam, V.; Kizek, R.; Hubalek, J. Quantum Dots—Characterization, Preparation and Usage in Biological Systems. *Int. J. Mol. Sci.* **2009**, *10*, 656–673. [\[CrossRef\]](https://doi.org/10.1021/es00005a015)
- 104. Pednekar, P.P.; Godiyal, S.C.; Jadhav, K.R.; Kadam, V.J. Chapter 23—Mesoporous silica nanoparticles: A promising multifunctional drug delivery system. In *Nanostructures for Cancer Therapy*; Elsevier: Amsterdam, The Netherlands, 2017; pp. 593–621. Available online: <https://www.sciencedirect.com/science/article/abs/pii/B9780323461443000234> (accessed on 4 February 2024).
- 105. Frickenstein, A.N.; Hagood, J.M.; Britten, C.N.; Abbott, B.S.; McNally, M.W.; Vopat, C.A.; Patterson, E.G.; MacCuaig, W.M.; Jain, A.; Walters, K.B.; et al. Mesoporous Silica Nanoparticles: Properties and Strategies for Enhancing Clinical Effect. *Pharmaceutics* **2021**, *13*, 570. [\[CrossRef\]](https://doi.org/10.3390/pharmaceutics13040570) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33920503)
- 106. Bharti, C.; Nagaich, U.; Pal, A.K.; Gulati, N. Mesoporous silica nanoparticles in target drug delivery system: A review. *Int. J. Pharm. Investig.* **2015**, *5*, 124–133. [\[CrossRef\]](https://doi.org/10.4103/2230-973X.160844)
- 107. Serini, S.; Cassano, R.; Trombino, S.; Calviello, G. Nanomedicine-based formulations containing ω-3 polyunsaturated fatty acids: Potential application in cardiovascular and neoplastic diseases. *Int. J. Nanomed.* **2019**, *14*, 2809–2828. [\[CrossRef\]](https://doi.org/10.2147/ijn.s197499)
- 108. Deshpande, D.; Janero, D.R.; Amiji, M. Engineering of an ω-3 polyunsaturated fatty acid-containing nanoemulsion system for combination C6-ceramide and 17β-estradiol delivery and bioactivity in human vascular endothelial and smooth muscle cells. *Nanomed. Nanotechnol. Biol. Med.* **2013**, *9*, 885–894. [\[CrossRef\]](https://doi.org/10.1016/j.nano.2013.02.007)
- 109. Deshpande, D.; Kethireddy, S.; Janero, D.R.; Amiji, M.M. Therapeutic Efficacy of an ω-3-Fatty Acid-Containing 17-β Estradiol Nano-Delivery System against Experimental Atherosclerosis. Zhu X, editor. *PLoS ONE* **2016**, *11*, e0147337. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0147337)
- 110. Sreedhar, R.; Kumar, V.S.; Bhaskaran Pillai, A.K.; Mangalathillam, S. Omega-3 Fatty Acid Based Nanolipid Formulation of Atorvastatin for Treating Hyperlipidemia. *Adv. Pharm. Bull.* **2019**, *9*, 271–280. [\[CrossRef\]](https://doi.org/10.15171/apb.2019.031)
- 111. Wakil, A.; Mackenzie, G.; Diego-Taboada, A.; Bell, J.G.; Atkin, S.L. Enhanced Bioavailability of Eicosapentaenoic Acid from Fish Oil After Encapsulation Within Plant Spore Exines as Microcapsules. *Lipids* **2010**, *45*, 645–649. [\[CrossRef\]](https://doi.org/10.1007/s11745-010-3427-y) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20495964)
- 112. Sanguansri, L.; Shen, Z.; Weerakkody, R.; Barnes, M.; Lockett, T.; Augustin, M.A. Omega-3 fatty acids in ileal effluent after consuming different foods containing microencapsulated fish oil powder—An ileostomy study. *Food Funct.* **2013**, *4*, 74–82. [\[CrossRef\]](https://doi.org/10.1039/c2fo30133d) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22992723)
- 113. Busolo, M.A.; Torres-Giner, S.; Prieto, C.; Lagaron, J.M. Electrospraying assisted by pressurized gas as an innovative highthroughput process for the microencapsulation and stabilization of docosahexaenoic acid-enriched fish oil in zein prolamine. *Innov. Food Sci. Emerg. Technol.* **2019**, *51*, 12–19. [\[CrossRef\]](https://doi.org/10.1016/j.ifset.2018.04.007)
- 114. Wang, G.S.; Chen, H.; Feng, G.; Yuan, Y.; Wan, Z.; Guo, J.; Wang, J.; Yang, X. Polyphenol-Enriched Protein Oleogels as Potential Delivery Systems of Omega-3 Fatty Acids. *J. Agric. Food Chem.* **2022**, *71*, 749–759. [\[CrossRef\]](https://doi.org/10.1021/acs.jafc.2c06348) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36534616)
- 115. Yoon, G.; Park, J.W.; Yoon, I.S. Solid lipid nanoparticles (SLNs) and nanostructured lipid carriers (NLCs): Recent advances in drug delivery. *J. Pharm. Investig.* **2013**, *43*, 353–362. [\[CrossRef\]](https://doi.org/10.1007/s40005-013-0087-y)
- 116. Makwana, V.; Jain, R.; Patel, K.; Nivsarkar, M.; Joshi, A. Solid lipid nanoparticles (SLN) of Efavirenz as lymph targeting drug delivery system: Elucidation of mechanism of uptake using chylomicron flow blocking approach. *Int. J. Pharm.* **2015**, *495*, 439–446. [\[CrossRef\]](https://doi.org/10.1016/j.ijpharm.2015.09.014) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26367780)
- 117. Tran, T.H.; Ramasamy, T.; Truong, D.H.; Choi, H.G.; Yong, C.S.; Kim, J.O. Preparation and Characterization of Fenofibrate-Loaded Nanostructured Lipid Carriers for Oral Bioavailability Enhancement. *AAPS PharmSciTech* **2014**, *15*, 1509–1515. [\[CrossRef\]](https://doi.org/10.1208/s12249-014-0175-y) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25035071)
- 118. Feng, Q.; Li, D.; Li, Q.; Cao, X.; Dong, H. Microgel assembly: Fabrication, characteristics and application in tissue engineering and regenerative medicine. *Bioact. Mater.* **2022**, *9*, 105–119. [\[CrossRef\]](https://doi.org/10.1016/j.bioactmat.2021.07.020) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34820559)
- 119. El-Seedi, H.R.; Said, N.S.; Yosri, N.; BIHawash, H.; El-Sherif, D.M.; Abouzid, M.; Abdel-Daim, M.M.; Yaseen, M.; Omar, H.; Shou, Q.; et al. Gelatin nanofibers: Recent insights in synthesis, bio-medical applications, and limitations. *Heliyon* **2023**, *9*, e16228. [\[CrossRef\]](https://doi.org/10.1016/j.heliyon.2023.e16228)
- 120. Yaghi, O.M.; O'Keeffe, M.; Ockwig, N.W.; Chae, H.K.; Eddaoudi, M.; Kim, J. Reticular synthesis and the design of new materials. *Nature* **2003**, *423*, 705–714. [\[CrossRef\]](https://doi.org/10.1038/nature01650)
- 121. Baumann, A.E.; Burns, D.A.; Liu, B.; Thoi, V.S. Metal-organic framework functionalization and design strategies for advanced electrochemical energy storage devices. *Commun. Chem.* **2019**, *2*, 86. [\[CrossRef\]](https://doi.org/10.1038/s42004-019-0184-6)
- 122. Hirschle, P.; Preiß, T.; Auras, F.; Pick, A.; Völkner, J.; Valdepérez, D.; Witte, G.; Parak, W.J.; Rädler, J.O.; Wuttke, S. Exploration of MOF nanoparticle sizes using various physical characterization methods—Is what you measure what you get? *CrystEngComm* **2016**, *18*, 4359–4368. [\[CrossRef\]](https://doi.org/10.1039/c6ce00198j)
- 123. Liu, L.; Zhou, Y.; Liu, S.; Xu, M. The Applications of Metal−Organic Frameworks in Electrochemical Sensors. *ChemElectroChem* **2018**, *5*, 6–19. [\[CrossRef\]](https://doi.org/10.1002/celc.201700931)
- 124. Al-Ansari, D.E.; Mohamed, N.A.; Marei, I.; Zekri, A.; Kameno, Y.; Davies, R.P.; Lickiss, P.D.; Rahman, M.M.; Abou-Saleh, H. Internalization of Metal-Organic Framework Nanoparticles in Human Vascular Cells: Implications for Cardiovascular Disease Therapy. *Nanomaterials.* **2020**, *10*, 1028. [\[CrossRef\]](https://doi.org/10.3390/nano10061028)
- 125. Mohamed, N.A. *Metal-Organic Frameworks, Their Properties and Future Promises in the Medical Field*; Nova Science Publishers, Inc.: Hauppauge, NY, USA, 2021. Available online: <https://qspace.qu.edu.qa/handle/10576/48948> (accessed on 12 April 2024).
- 126. Ahangaran, F.; Navarchian, A.H. Recent advances in chemical surface modification of metal oxide nanoparticles with silane coupling agents: A review. *Adv. Colloid Interface Sci.* **2020**, *286*, 102298. [\[CrossRef\]](https://doi.org/10.1016/j.cis.2020.102298)
- 127. Ranjha, M.M.A.N.; Shafique, B.; Rehman, A.; Mehmood, A.; Ali, A.; Zahra, S.M.; Roobab, U.; Singh, A.; Ibrahim, S.A.; Siddiqui, S.A. Biocompatible Nanomaterials in Food Science, Technology, and Nutrient Drug Delivery: Recent Developments and Applications. *Front. Nutr.* **2022**, *8*, 778155. [\[CrossRef\]](https://doi.org/10.3389/fnut.2021.778155)
- 128. Morshed, M.; Chowdhury, E.H. *Gene Delivery and Clinical Applications*; Elsevier: Amsterdam, The Netherlands, 2019; pp. 345–351. [\[CrossRef\]](https://doi.org/10.1016/b978-0-12-801238-3.99883-0)
- 129. Farzan, M.; Roth, R.; Schoelkopf, J.; Huwyler, J.; Puchkov, M. The processes behind drug loading and release in porous drug delivery systems. *Eur. J. Pharm. Biopharm.* **2023**, *189*, 133–151. [\[CrossRef\]](https://doi.org/10.1016/j.ejpb.2023.05.019) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37230292)
- 130. Sharma, P.R.; Varma, A.J. Thermal stability of cellulose and their nanoparticles: Effect of incremental increases in carboxyl and aldehyde groups. *Carbohydr. Polym.* **2014**, *114*, 339–343. [\[CrossRef\]](https://doi.org/10.1016/j.carbpol.2014.08.032)
- 131. Sun, D.; Kim, D.P. Hydrophobic MOFs@Metal Nanoparticles@COFs for Interfacially Confined Photocatalysis with High Efficiency. *ACS Appl. Mater. Interfaces* **2020**, *12*, 20589–20595. [\[CrossRef\]](https://doi.org/10.1021/acsami.0c04537)
- 132. Yang, S.; Peng, L.; Sun, D.T.; Asgari, M.; Oveisi, E.; Trukhina, O.; Bulut, S.; Jamali, A.; Queen, W.L. A new post-synthetic polymerization strategy makes metal-organic frameworks more stable. *Chem. Sci.* **2019**, *10*, 4542–4549. [\[CrossRef\]](https://doi.org/10.1039/c9sc00135b)
- 133. Paranjpe, M.; Müller-Goymann, C. Nanoparticle-Mediated Pulmonary Drug Delivery: A Review. *Int. J. Mol. Sci.* **2014**, *15*, 5852–5873. [\[CrossRef\]](https://doi.org/10.3390/ijms15045852) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24717409)
- 134. Muchow, M.; Schmitz, E.I.; Despatova, N.; Maincent, P.; Müller, R.H. Omega-3 fatty acids-loaded lipid nanoparticles for patient-convenient oral bioavailability enhancement. *Pharm.-Int. J. Pharm. Sci.* **2009**, *64*, 499–504.
- 135. Guo, X.; Zuo, X.; Zhou, Z.; Gu, Y.; Zheng, H.; Wang, X.; Wang, G.; Xu, C.; Wang, F. PLGA-Based Micro/Nanoparticles: An Overview of Their Applications in Respiratory Diseases. *Int. J. Mol. Sci.* **2023**, *24*, 4333. [\[CrossRef\]](https://doi.org/10.3390/ijms24054333) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36901762)
- 136. Frank, L.A.; Onzi, G.R.; Morawski, A.S.; Pohlmann, A.R.; Guterres, S.S.; Contri, R.V. Chitosan as a coating material for nanoparticles intended for biomedical applications. *React. Funct. Polym.* **2020**, *147*, 104459. [\[CrossRef\]](https://doi.org/10.1016/j.reactfunctpolym.2019.104459)
- 137. Horcajada, P.; Chalati, T.; Serre, C.; Gillet, B.; Sebrie, C.; Baati, T.; Eubank, J.F.; Heurtaux, D.; Clayette, P.; Kreuz, C.; et al. Porous metal–metal-organic-framework nanoscale carriers as a potential platform for drug delivery and imaging. *Nat. Mater.* **2009**, *9*, 172–178. [\[CrossRef\]](https://doi.org/10.1038/nmat2608) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20010827)
- 138. Dehghani, S.; Hosseini, M.; Haghgoo, S.; Changizi, V.; Akbari Javar, H.; Khoobi, M.; Riahi Alam, N. Multifunctional MIL-Cur@FC as a theranostic agent for magnetic resonance imaging and targeting drug delivery: In vitro and in vivo study. *J. Drug Target.* **2020**, *28*, 668–680. [\[CrossRef\]](https://doi.org/10.1080/1061186x.2019.1710839) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31886726)
- 139. Karimi Alavijeh, R.; Akhbari, K. Cancer therapy by nano MIL-n series of metal-organic frameworks. *Coord. Chem. Rev.* **2024**, *503*, 215643. [\[CrossRef\]](https://doi.org/10.1016/j.ccr.2023.215643)
- 140. Xiong, F.; Chen, Y.; Chen, J.; Yang, B.; Zhang, Y.; Gao, H.; Hua, Z.; Gu, N. Rubik-like magnetic nanoassemblies as an efficient drug multifunctional carrier for cancer theranostics. *J. Control. Release* **2013**, *172*, 993–1001. [\[CrossRef\]](https://doi.org/10.1016/j.jconrel.2013.09.023) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24096016)
- 141. Chen, Y.; Feng, X. Gold nanoparticles for skin drug delivery. *Int. J. Pharm.* **2022**, *625*, 122122. [\[CrossRef\]](https://doi.org/10.1016/j.ijpharm.2022.122122)
- 142. Gao, W.; Sun, Y.; Cai, M.; Zhao, Y.; Cao, W.; Liu, Z.; Cui, G.; Tang, B. Copper sulfide nanoparticles as a photothermal switch for TRPV1 signaling to attenuate atherosclerosis. *Nat. Commun.* **2018**, *9*, 231. [\[CrossRef\]](https://doi.org/10.1038/s41467-017-02657-z)
- 143. Ya Spivak, M.; Bubnov, R.V.; Yemets, I.M.; Lazarenko, L.M.; Tymoshok, N.O.; Ul'berg, Z.R. Development and testing of gold nanoparticles for drug delivery and treatment of heart failure: A theranostic potential for PPP cardiology. *EPMA J.* **2013**, *4*, 20. [\[CrossRef\]](https://doi.org/10.1186/1878-5085-4-20) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23889805)
- 144. Li, H.; Yan, J.; Meng, D.; Cai, R.; Gao, X.; Ji, Y.; Wang, L.; Chen, C.; Wu, X. Gold Nanorod-Based Nanoplatform Catalyzes Constant NO Generation and Protects from Cardiovascular Injury. *ACS Nano* **2020**, *14*, 12854–12865. [\[CrossRef\]](https://doi.org/10.1021/acsnano.0c03629) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32955857)
- 145. Bejarano, J.; Rojas, A.; Ramírez-Sagredo, A.; Riveros, A.L.; Morales-Zavala, F.; Flores, Y.; Riquelme, J.A.; Guzman, F.; Araya, E.; Chiong, M.; et al. Light-induced release of the cardioprotective peptide angiotensin-(1–9) from thermosensitive liposomes with gold nanoclusters. *J. Control. Release* **2020**, *328*, 859–872. [\[CrossRef\]](https://doi.org/10.1016/j.jconrel.2020.11.002) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33160006)
- 146. Hussein, J.; Attia, M.F.; El Bana, M.; El-Daly, S.M.; Mohamed, N.; El-Khayat, Z.; El-Naggar, M.E. Solid state synthesis of docosahexaenoic acid-loaded zinc oxide nanoparticles as a potential antidiabetic agent in rats. *Int. J. Biol. Macromol.* **2019**, *140*, 1305–1314. [\[CrossRef\]](https://doi.org/10.1016/j.ijbiomac.2019.08.201) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31449866)
- 147. Hussein, J.; El-Naggar, M.E.; Badawy, E.; El-Laithy, N.; El-Waseef, M.; Hassan, H.; Abdel-Latif, Y. Homocysteine and Asymmetrical Dimethylarginine in Diabetic Rats Treated with Docosahexaenoic Acid–Loaded Zinc Oxide Nanoparticles. *Appl. Biochem. Biotechnol.* **2020**, *191*, 1127–1139. [\[CrossRef\]](https://doi.org/10.1007/s12010-020-03230-z) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31960366)
- 148. Hussein, J.; Rasheed, W.; Ramzy, T.; Nabeeh, M.; Harvy, M.; El-Toukhy, S.; Ali, O.; Raafat, J.; El-Naggar, M. Synthesis of docosahexaenoic acid–loaded silver nanoparticles for improving endothelial dysfunctions in experimental diabetes. *Hum. Exp. Toxicol.* **2019**, *38*, 962–973. [\[CrossRef\]](https://doi.org/10.1177/0960327119843586) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31018711)
- 149. El-Daly, S.M.; Medhat, D.; El-Bana, M.; Abdel-Latif, Y.; El-Naggar, M.E.; Omara, E.A.; Morsy, S.M.; Hussein, J. Stimulatory effect of docosahexaenoic acid alone or loaded in zinc oxide or silver nanoparticles on the expression of glucose transport pathway. *Prostaglandins Other Lipid Mediat.* **2021**, *155*, 106566. [\[CrossRef\]](https://doi.org/10.1016/j.prostaglandins.2021.106566) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34048868)
- 150. Ali, A.H.; Hachem, M.; Ahmmed, M.K. Docosahexaenoic acid-loaded nanoparticles: A state-of-the-art of preparation methods, characterization, functionality, and therapeutic applications. *Heliyon* **2024**, *10*, e30946. [\[CrossRef\]](https://doi.org/10.1016/j.heliyon.2024.e30946)
- 151. Ahmed, S.M.; Abdelrahman, S.A.; Salama, A.E. Efficacy of gold nanoparticles against isoproterenol-induced acute myocardial infarction in adult male albino rats. *Ultrastruct. Pathol.* **2017**, *41*, 168–185. [\[CrossRef\]](https://doi.org/10.1080/01913123.2017.1281367)
- 152. Raptopoulou, C.P. Metal-Organic Frameworks: Synthetic Methods and Potential Applications. *Materials* **2021**, *14*, 310. [\[CrossRef\]](https://doi.org/10.3390/ma14020310)
- 153. Shahini, M.H.; Mohammadloo, H.E.; Ramezanzadeh, M.; Ramezanzadeh, B. Recent innovations in synthesis/characterization of advanced nanoporous metal-organic frameworks (MOFs); current/future trends with a focus on the smart anti-corrosion features. *Mater. Chem. Phys.* **2022**, *276*, 125420. [\[CrossRef\]](https://doi.org/10.1016/j.matchemphys.2021.125420)

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